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Attorney Docket No.: DIVER1280-3

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Presented for filing is a new continuation-in-part patent application of:

APPLICANTS: JAY M. SHORT and MARTIN KELLER

**TITLE: HIGH THROUGHPUT SCREENING FOR SEQUENCES OF
INTEREST**

Under 35 USC § 120, this application claims the benefit of prior U.S. applications Serial No. 09/444,112, filed November 22, 1999, which is a continuation-in-part of U.S. Patent Application Serial No. 09/098,206, filed June 16, 1998, which is a continuation-in-part of U.S. Patent Application Serial No. 08/876,276, filed June 16, 1997, the contents of all of which are incorporated by reference in their entirety herein.

Enclosed are the following papers, including all those required for a filing date under 37 CFR § 1.53(b):

Pages of Specification	112
Pages of Claims	9
Pages of Abstract	1
Pages of Declaration	3 (unsigned)
Sheets of Drawings	5

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CIP APPLICATION

for

UNITED STATES LETTERS PATENT

on

HIGH THROUGHPUT SCREENING FOR SEQUENCES OF INTEREST

by

Jay M. Short

Martin Keller

Sheets of Drawings: Five (5)
 Docket No.: DIVER1280-3

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HIGH THROUGHPUT SCREENING FOR SEQUENCES OF INTEREST

CROSS REFERENCE TO RELATED APPLICATIONS

This application is a continuation-in-part of U.S. Patent Application Serial No. 09/444,112, filed November 22, 1999 (pending), which is a continuation-in-part of 5 U.S. Patent Application Serial No. 09/098,206, filed June 16, 1998 (pending), which application is a continuation-in-part of U.S. Patent Application Serial No. 08/876,276, filed June 16, 1997 (pending), the contents of which are all incorporated by reference in their entirety herein.

FIELD OF THE INVENTION

10 The present invention relates generally to screening of mixed populations of organisms or nucleic acids and more specifically to the identification of bioactive molecules and bioactivities by using high throughput screening techniques, including fluorescence activated cell sorting (FACS).

BACKGROUND

15 There is a critical need in the chemical industry for efficient catalysts for the practical synthesis of optically pure materials; enzymes can provide the optimal solution. All classes of molecules and compounds that are utilized in both established and emerging chemical, pharmaceutical, textile, food and feed, detergent markets must meet stringent economical and environmental standards. The synthesis of 20 polymers, pharmaceuticals, natural products and agrochemicals is often hampered by expensive processes which produce harmful byproducts and which suffer from low enantioselectivity (Faber, 1995; Tonkovich and Gerber, U.S. Dept of Energy study, 1995). Enzymes have a number of remarkable advantages which can overcome these problems in catalysis: they act on single functional groups, they distinguish between 25 similar functional groups on a single molecule, and they distinguish between enantiomers. Moreover, they are biodegradable and function at very low mole fractions in reaction mixtures. Because of their chemo-, regio- and stereospecificity, enzymes present a unique opportunity to optimally achieve desired selective transformations. These are often extremely difficult to duplicate chemically,

especially in single-step reactions. The elimination of the need for protection groups, selectivity, the ability to carry out multi-step transformations in a single reaction vessel, along with the concomitant reduction in environmental burden, has led to the increased demand for enzymes in chemical and pharmaceutical industries (Faber, 5 1995). Enzyme-based processes have been gradually replacing many conventional chemical-based methods (Wrotnowski, 1997). A current limitation to more widespread industrial use is primarily due to the relatively small number of commercially available enzymes. Only ~300 enzymes (excluding DNA modifying enzymes) are at present commercially available from the > 3000 non DNA-modifying 10 enzyme activities thus far described.

The use of enzymes for technological applications also may require performance under demanding industrial conditions. This includes activities in environments or on substrates for which the currently known arsenal of enzymes was 15 not evolutionarily selected. Enzymes have evolved by selective pressure to perform very specific biological functions within the milieu of a living organism, under conditions of mild temperature, pH and salt concentration. For the most part, the non-DNA modifying enzyme activities thus far described (Enzyme Nomenclature, 1992) have been isolated from mesophilic organisms, which represent a very small fraction 20 of the available phylogenetic diversity (Amann *et al.*, 1995). The dynamic field of biocatalysis takes on a new dimension with the help of enzymes isolated from microorganisms that thrive in extreme environments. Such enzymes must function at temperatures above 100 °C in terrestrial hot springs and deep sea thermal vents, at temperatures below 0 °C in arctic waters, in the saturated salt environment of the 25 Dead Sea, at pH values around 0 in coal deposits and geothermal sulfur-rich springs, or at pH values greater than 11 in sewage sludge (Adams and Kelly, 1995). Enzymes obtained from these extremophilic organisms open a new field in biocatalysis.

For example, several esterases and lipases cloned and expressed from 30 extremophilic organisms are remarkably robust, showing high activity throughout a wide range of temperatures and pHs. The fingerprints of five of these esterases show a diverse substrate spectrum, in addition to differences in the optimum reaction

temperature. As seen in Figure 1, esterase #5 recognizes only short chain substrates while #2 only acts on long chain substrates in addition to a huge difference in the optimal reaction temperature. These results suggest that more diverse enzymes fulfilling the need for new biocatalysts can be found by screening biodiversity.

5 Substrates upon which enzymes act are herein defined as bioactive substrates.

Furthermore, virtually all of the enzymes known so far have come from cultured organisms, mostly bacteria and more recently archaea (Enzyme Nomenclature, 1992). Traditional enzyme discovery programs rely solely on cultured 10 microorganisms for their screening programs and are thus only accessing a small fraction of natural diversity. Several recent studies have estimated that only a small percentage, conservatively less than 1%, of organisms present in the natural environment have been cultured (see Table I, Amann *et al.*, 1995, Barns et. al 1994, Torsvik, 1990). For example, Norman Pace's laboratory recently reported intensive 15 untapped diversity in water and sediment samples from the "Obsidian Pool" in Yellowstone National Park, a spring which has been studied since the early 1960 s by microbiologists (Barns, 1994). Amplification and cloning of 16S rRNA encoding sequences revealed mostly unique sequences with little or no representation of the organisms which had previously been cultured from this pool. This suggests 20 substantial diversity of archaea with so far unknown morphological, physiological and biochemical features which may be useful in industrial processes. David Ward's laboratory in Bozeman, Montana has performed similar studies on the cyanobacterial mat of Octopus Spring in Yellowstone Park and came to the same conclusion, namely, tremendous uncultured diversity exists (Bateson *et al.*, 1989). Giovannoni *et al.* 25 (1990) reported similar results using bacterioplankton collected in the Sargasso Sea while Torsvik *et al.* (1990) have shown by DNA reassociation kinetics that there is considerable diversity in soil samples. Hence, this vast majority of microorganisms represents an untapped resource for the discovery of novel biocatalysts. In order to 30 access this potential catalytic diversity, recombinant screening approaches are required.

The discovery of novel bioactive molecules other than enzymes is also afforded by the present invention. For instance, antibiotics, antivirals, antitumor agents and regulatory proteins can be discovered utilizing the present invention.

5 Bacteria and many eukaryotes have a coordinated mechanism for regulating genes whose products are involved in related processes. The genes are clustered, in structures referred to as "gene clusters," on a single chromosome and are transcribed together under the control of a single regulatory sequence, including a single promoter which initiates transcription of the entire cluster. The gene cluster, the promoter, and
10 additional sequences that function in regulation altogether are referred to as an "operon" and can include up to 20 or more genes, usually from 2 to 6 genes. Thus, a gene cluster is a group of adjacent genes that are either identical or related, usually as to their function.

15 Some gene families consist of one or more identical members. Clustering is a prerequisite for maintaining identity between genes, although clustered genes are not necessarily identical. Gene clusters range from extremes where a duplication is generated of adjacent related genes to cases where hundreds of identical genes lie in a tandem array. Sometimes no significance is discernable in a repetition of a particular
20 gene. A principal example of this is the expressed duplicate insulin genes in some species, whereas a single insulin gene is adequate in other mammalian species.

It is important to further research gene clusters and the extent to which the full length of the cluster is necessary for the expression of the proteins resulting
25 therefrom. Gene clusters undergo continual reorganization and, thus, the ability to create heterogeneous libraries of gene clusters from, for example, bacterial or other prokaryote sources is valuable in determining sources of novel proteins, particularly including enzymes such as, for example, the polyketide synthases that are responsible for the synthesis of polyketides having a vast array of useful activities. As indicated,
30 other types of proteins that are the product(s) of gene clusters are also contemplated, including, for example, antibiotics, antivirals, antitumor agents and regulatory proteins, such as insulin.

Polyketides are molecules which are an extremely rich source of bioactivities, including antibiotics (such as tetracyclines and erythromycin), anti-cancer agents (daunomycin), immunosuppressants (FK506 and rapamycin), and veterinary products (monensin). Many polyketides (produced by polyketide synthases) are valuable as therapeutic agents. Polyketide synthases are multifunctional enzymes that catalyze the biosynthesis of a huge variety of carbon chains differing in length and patterns of functionality and cyclization. Polyketide synthase genes fall into gene clusters and at least one type (designated type I) of polyketide synthases have large size genes and encoded enzymes, complicating genetic manipulation and *in vitro* studies of these genes/proteins. The method(s) of the present invention facilitate the rapid discovery of these gene clusters in gene expression libraries.

Of particular interest are cellular "switches" known as receptors which interact with a variety of biomolecules, such as hormones, growth factors, and neurotransmitters, to mediate the transduction of an "external" cellular signaling event into an "internal" cellular signal. External signaling events include the binding of a ligand to the receptor, and internal events include the modulation of a pathway in the cytoplasm or nucleus involved in the growth, metabolism or apoptosis of the cell. Internal events also include the inhibition or activation of transcription of certain nucleic acid sequences, resulting in the increase or decrease in the production or presence of certain molecules (such as nucleic acid, proteins, and/or other molecules affected by this increase or decrease in transcription). Drugs to cure disease or alleviate its symptoms can activate or block any of these events to achieve a desired pharmaceutical effect.

Transduction can be accomplished by a transducing protein in the cell membrane which is activated upon an allosteric change the receptor may undergo upon binding to a specific biomolecule. The "active" transducing protein activates production of so-called "second messenger" molecules within the cell, which then activate certain regulatory proteins within the cell that regulate gene expression or alter some metabolic process. Variations on the theme of this "cascade" of events

occur. For example, a receptor may act as its own transducing protein, or a transducing protein may act directly on an intracellular target without mediation by a second messenger.

5 Signal transduction is a fundamental area of inquiry in biology. For instance, ligand/receptor interactions and the receptor/effectector coupling mediated by Guanine nucleotide-binding proteins (G-proteins) are of interest in the study of disease. A large number of G protein-linked receptors funnel extracellular signals as diverse as hormones, growth factors, neurotransmitters, primary sensory stimuli, and other 10 signals through a set of G proteins to a small number of second-messenger systems. The G proteins act as molecular switches with an "on" and "off" state governed by a GTPase cycle. Mutations in G proteins may result in either constitutive activation or loss of expression mutations.

15 Many receptors convey messages through heterotrimeric G proteins, of which at least 17 distinct forms have been isolated. Additionally, there are several different G protein-dependent effectors. The signals transduced through the heterotrimeric G proteins in mammalian cells influence intracellular events through the action of 20 effector molecules.

25 Given the variety of functions subserved by G protein-coupled signal transduction, it is not surprising that abnormalities in G protein-coupled pathways can lead to diseases with manifestations as dissimilar as blindness, hormone resistance, precocious puberty and neoplasia. G-protein-coupled receptors are extremely 30 important to drug research efforts. It is estimated that up to 60% of today's prescription drugs work by somehow interacting with G protein-coupled receptors. However, these drugs were developed using classical medicinal chemistry and without a knowledge of the molecular mechanism of action. A more efficient drug discovery program could be deployed by targeting individual receptors and making use of information on gene sequence and biological function to develop effective 35 therapeutics. The present invention allows one to, for example, study molecules which affect the interaction of G proteins with receptors, or of ligands with receptors.

Several groups have reported cells which express mammalian G proteins or subunits thereof, along with mammalian receptors which interact with these molecules. For example, WO92/05244 (April 2, 1992) describes a transformed yeast 5 cell which is incapable of producing a yeast G protein subunit, but which has been engineered to produce both a mammalian G protein subunit and a mammalian receptor which interacts with the subunit. The authors found that a modified version of a specific mammalian receptor integrated into the membrane of the cell, as shown by studies of the ability of isolated membranes to interact properly with various 10 known agonists and antagonists of the receptor. Ligand binding resulted in G protein-mediated signal transduction.

Another group has described the functional expression of a mammalian adenylyl cyclase in yeast, and the use of the engineered yeast cells in identifying 15 potential inhibitors or activators of the mammalian adenylyl cyclase (WO 95/30012). Adenylyl cyclase is among the best studied of the effector molecules which function in mammalian cells in response to activated G proteins. "Activators" of adenylyl cyclase cause the enzyme to become more active, elevating the cAMP signal of the yeast cell to a detectable degree. "Inhibitors" cause the cyclase to become less active, 20 reducing the cAMP signal to a detectable degree. The method describes the use of the engineered yeast cells to screen for drugs which activate or inhibit adenylyl cyclase by their action on G protein-coupled receptors.

When attempting to identify genes encoding bioactivities of interest from 25 complex environmental expression libraries, the rate limiting steps in discovery occur at the both DNA cloning level and at the screening level. Screening of complex environmental libraries which contain, for example, 100 s of different organisms requires the analysis of several million clones to cover this genomic diversity. An extremely high-throughput screening method has been developed to handle the 30 enormous numbers of clones present in these libraries.

In traditional flow cytometry, it is common to analyze very large numbers of eukaryotic cells in a short period of time. Newly developed flow cytometers can analyze and sort up to 20,000 cells per second. In a typical flow cytometer, individual particles pass through an illumination zone and appropriate detectors, gated

5 electronically, measure the magnitude of a pulse representing the extent of light scattered. The magnitude of these pulses are sorted electronically into "bins" or "channels", permitting the display of histograms of the number of cells possessing a certain quantitative property versus the channel number (Davey and Kell, 1996). It was recognized early on that the data accruing from flow cytometric measurements

10 could be analyzed (electronically) rapidly enough that electronic cell-sorting procedures could be used to sort cells with desired properties into separate "buckets", a procedure usually known as fluorescence-activated cell sorting (Davey and Kell, 1996).

15 Fluorescence-activated cell sorting has been primarily used in studies of human and animal cell lines and the control of cell culture processes. Fluorophore labeling of cells and measurement of the fluorescence can give quantitative data about specific target molecules or subcellular components and their distribution in the cell population. Flow cytometry can quantitate virtually any cell-associated property or

20 cell organelle for which there is a fluorescent probe (or natural fluorescence). The parameters which can be measured have previously been of particular interest in animal cell culture.

Flow cytometry has also been used in cloning and selection of variants from

25 existing cell clones. This selection, however, has required stains that diffuse through cells passively, rapidly and irreversibly, with no toxic effects or other influences on metabolic or physiological processes. Since, typically, flow sorting has been used to study animal cell culture performance, physiological state of cells, and the cell cycle, one goal of cell sorting has been to keep the cells viable during and after sorting.

30

There currently are no reports in the literature of screening and discovery of recombinant enzymes in *E. coli* expression libraries by fluorescence activated cell

sorting of single cells. Furthermore there are no reports of recovering DNA encoding bioactivities screened by expression screening in *E. coli* using a FACS machine. The present invention provides these methods to allow the extremely rapid screening of viable or non-viable cells to recover desirable activities and the nucleic acid encoding those activities.

A limited number of papers describing various applications of flow cytometry in the field of microbiology and sorting of fluorescence activated microorganisms have, however, been published (Davey and Kell, 1996). Fluorescence and other forms of staining have been employed for microbial discrimination and identification, and in the analysis of the interaction of drugs and antibiotics with microbial cells. Flow cytometry has been used in aquatic biology, where autofluorescence of photosynthetic pigments are used in the identification of algae or DNA stains are used to quantify and count marine populations (Davey and Kell, 1996). Thus, Diaper and Edwards used flow cytometry to detect viable bacteria after staining with a range of fluorogenic esters including fluorescein diacetate (FDA) derivatives and CemChrome B, a proprietary stain sold commercially for the detection of viable bacteria in suspension (Diaper and Edwards, 1994). Labeled antibodies and oligonucleotide probes have also been used for these purposes.

20

Papers have also been published describing the application of flow cytometry to the detection of native and recombinant enzymatic activities in eukaryotes. Betz *et al.* studied native (non-recombinant) lipase production by the eukaryote, *Rhizopus arrhizus* with flow cytometry. They found that spore suspensions of the mold were heterogeneous as judged by light-scattering data obtained with excitation at 633 nm, and they sorted clones of the subpopulations into the wells of microtiter plates. After germination and growth, lipase production was automatically assayed (turbidimetrically) in the microtiter plates, and a representative set of the most active were reisolated, cultured, and assayed conventionally (Betz *et al.*, 1984).

25
30

Scrienc *et al.* have reported a flow cytometric method for detecting cloned - galactosidase activity in the eukaryotic organism, *S. cerevisiae*. The ability of flow

cytometry to make measurements on single cells means that individual cells with high levels of expression (e.g., due to gene amplification or higher plasmid copy number) could be detected. In the method reported, a non-fluorescent compound α -naphthol- β -galactopyranoside) is cleaved by β -galactosidase and the liberated naphthol is 5 trapped to form an insoluble fluorescent product. The insolubility of the fluorescent product is of great importance here to prevent its diffusion from the cell. Such diffusion would not only lead to an underestimation of β -galactosidase activity in highly active cells but could also lead to an overestimation of enzyme activity in inactive cells or those with low activity, as they may take up the leaked fluorescent 10 compound, thus reducing the apparent heterogeneity of the population.

One group has described the use of a FACS machine in an assay detecting fusion proteins expressed from a specialized transducing bacteriophage in the prokaryote *Bacillus subtilis* (Chung, et.al., *J. of Bacteriology*, Apr. 1994, p. 1977-15 1984; Chung, et.al., *Biotechnology and Bioengineering*, Vol. 47, pp. 234-242 (1995)). This group monitored the expression of a lacZ gene (encodes β -galactosidase) fused to the sporulation loci in *subtilis* (spo). The technique used to monitor β -galactosidase expression from spo-lacZ fusions in single cells involved taking samples from a sporulating culture, staining them with a commercially available fluorogenic substrate 20 for β -galactosidase called C8-FDG, and quantitatively analyzing fluorescence in single cells by flow cytometry. In this study, the flow cytometer was used as a detector to screen for the presence of the spo gene during the development of the cells. The device was not used to screen and recover positive cells from a gene expression library or nucleic acid for the purpose of discovery.

25

Another group has utilized flow cytometry to distinguish between the developmental stages of the delta-proteobacteria *Myxococcus xanthus* (F. Russo-Marie, et.al., *PNAS*, Vol. 90, pp.8194-8198, September 1993). As in the previously 30 described study, this study employed the capabilities of the FACS machine to detect and distinguish genotypically identical cells in different development regulatory states. The screening of an enzymatic activity was used in this study as an indirect measure of developmental changes.

The lacZ gene from *E. coli* is often used as a reporter gene in studies of gene expression regulation, such as those to determine promoter efficiency, the effects of trans-acting factors, and the effects of other regulatory elements in bacterial, yeast, 5 and animal cells. Using a chromogenic substrate, such as ONPG (o-nitrophenyl-(D-galactopyranoside), one can measure expression of β -galactosidase in cell cultures; but it is not possible to monitor expression in individual cells and to analyze the heterogeneity of expression in cell populations. The use of fluorogenic substrates, however, makes it possible to determine β -galactosidase activity in a large number of 10 individual cells by means of flow cytometry. This type of determination can be more informative with regard to the physiology of the cells, since gene expression can be correlated with the stage in the mitotic cycle or the viability under certain conditions. In 1994, Plovins *et al.*, reported the use of fluorescein-Di- β -D-galactopyranoside (FDG) and C12-FDG as substrates for β -galactosidase detection in animal, bacterial, 15 and yeast cells. This study compared the two molecules as substrates for β -galactosidase, and concluded that FDG is a better substrate for β -galactosidase detection by flow cytometry in bacterial cells. The screening performed in this study was for the comparison of the two substrates. The detection capabilities of a FACS machine were employed to perform the study on viable bacterial cells.

20 Cells with chromogenic or fluorogenic substrates yield colored and fluorescent products, respectively. Previously, it had been thought that the flow cytometry-fluorescence activated cell sorter approaches could be of benefit only for the analysis of cells that contain intracellularly, or are normally physically associated 25 with, the enzymatic activity of small molecule of interest. On this basis, one could only use fluorogenic reagents which could penetrate the cell and which are thus potentially cytotoxic. To avoid clumping of heterogeneous cells, it is desirable in flow cytometry to analyze only individual cells, and this could limit the sensitivity and therefore the concentration of target molecules that can be sensed. Weaver and his 30 colleagues at MIT and others have developed the use of gel microdroplets containing (physically) single cells which can take up nutrients, secret products, and grow to form colonies. The diffusional properties of gel microdroplets may be made such that

sufficient extracellular product remains associated with each individual gel microdroplet, so as to permit flow cytometric analysis and cell sorting on the basis of concentration of secreted molecule within each microdroplet. Beads have also been used to isolate mutants growing at different rates, and to analyze antibody secretion 5 by hybridoma cells and the nutrient sensitivity of hybridoma cells. The gel microdroplet method has also been applied to the rapid analysis of mycobacterial growth and its inhibition by antibiotics.

10 The gel microdroplet technology has had significance in amplifying the signals available in flow cytometric analysis, and in permitting the screening of microbial strains in strain improvement programs for biotechnology. Wittrup *et al.*, (Biotechnolo.Bioeng. (1993) 42:351-356) developed a microencapsulation selection method which allows the rapid and quantitative screening of >106 yeast cells for enhanced secretion of *Aspergillus awamori* glucoamylase. The method provides a 15 400-fold single-pass enrichment for high-secretion mutants.

20 Gel microdroplet or other related technologies can be used in the present invention to localize as well as amplify signals in the high throughput screening of recombinant libraries. Cell viability during the screening is not an issue or concern since nucleic acid can be recovered from the microdroplet.

25 Different types of encapsulation strategies and compounds or polymers can be used with the present invention. For instance, high temperature agaroses can be employed for making microdroplets stable at high temperatures, allowing stable encapsulation of cells subsequent to heat kill steps utilized to remove all background activities when screening for thermostable bioactivities.

30 There are several hurdles which must be overcome when attempting to detect and sort *E. coli* expressing recombinant enzymes, and recover encoding nucleic acids. FACS systems have typically been based on eukaryotic separations and have not been refined to accurately sort single *E. coli* cells; the low forward and sideward scatter of small particles like *E. coli*, reduces the ability of accurate sorting; enzyme substrates

typically used in automated screening approaches, such as umbelliferyl based substrates, diffuse out of *E. coli* at rates which interfere with quantitation. Further, recovery of very small amounts of DNA from sorted organisms can be problematic. The present invention addresses and overcomes these hurdles and offers a novel 5 screening approach.

There has been a dramatic increase in the need for bioactive compounds with novel activities. This demand has arisen largely from changes in worldwide 10 demographics coupled with the clear and increasing trend in the number of pathogenic organisms that are resistant to currently available antibiotics as well as the need for new industrial processes for synthesis of compounds. For example, while there has been a surge in demand for antibacterial drugs in emerging nations with young 15 populations, countries with aging populations, such as the U.S., require a growing repertoire of drugs against cancer, diabetes, arthritis and other debilitating conditions. The death rate from infectious diseases has increased 58% between 1980 and 1992 and it has been estimated that the emergence of antibiotic resistant microbes has added in excess of \$30 billion annually to the cost of health care in the U.S. alone. 20 (Adams *et al.*, *Chemical and Engineering News*, 1995; Amann *et al.*, *Microbiological Reviews*, 59, 1995). As a response to this trend, pharmaceutical companies have significantly increased their screening of microbial diversity for compounds with unique activities or specificities.

The majority of bioactive compounds currently in use are derived from soil 25 microorganisms. Many microbes inhabiting soils and other complex ecological communities produce a variety of compounds that increase their ability to survive and proliferate. These compounds are generally thought to be nonessential for growth of the organism and are synthesized with the aid of genes involved in intermediary metabolism. Such secondary metabolites that influence the growth or survival of other organisms are known as "bioactive" compounds and serve as key components of 30 the chemical defense arsenal of both micro- and macroorganisms. Humans have exploited these compounds for use as antibiotics, antiinfectives and other bioactive

compounds with activity against a broad range of prokaryotic and eukaryotic pathogens (Barnes *et al.*, *Proc. Nat. Acad. Sci. U.S.A.*, **91**, 1994).

5 The approach currently used to screen microbes for new bioactive compounds has been largely unchanged since the inception of the field. New isolates of bacteria, particularly gram positive strains from soil environments, are collected and their metabolites tested for pharmacological activity.

10 There is still tremendous biodiversity that remains untapped as the source of lead compounds. However, the currently available methods for screening and producing lead compounds cannot be applied efficiently to these under-explored resources. For instance, it is estimated that at least 99% of marine bacteria species do not survive on laboratory media, and commercially available fermentation equipment is not optimal for use in the conditions under which these species will grow, hence 15 these organisms are difficult or impossible to culture for screening or re-supply. Recollection, growth, strain improvement, media improvement and scale-up production of the drug-producing organisms often pose problems for synthesis and development of lead compounds. Furthermore, the need for the interaction of specific organisms to synthesize some compounds makes their use in discovery extremely 20 difficult. New methods to harness the genetic resources and chemical diversity of these untapped sources of compounds for use in drug discovery are very valuable.

25 A central core of modern biology is that genetic information resides in a nucleic acid genome, and that the information embodied in such a genome (*i.e.*, the genotype) directs cell function. This occurs through the expression of various genes in the genome of an organism and regulation of the expression of such genes. The expression of genes in a cell or organism defines the cell or organism's physical characteristics (*i.e.*, its phenotype). This is accomplished through the translation of genes into proteins. Determining the biological activity of a protein obtained from an environmental sample 30 can provide valuable information about the role of proteins in the environments. In addition, such information can help in the development of biologics, diagnostics, therapeutics, and compositions for industrial applications.

Accordingly, the present invention provides methods and compositions to access this untapped biodiversity and to rapidly screen for polynucleotides and small molecules of interest utilizing high throughput screening of multiple samples.

5

SUMMARY OF THE INVENTION

The present invention adapts traditional eukaryotic flow cytometry cell sorting systems to high throughput screening for expression clones in prokaryotes. In the present invention, nucleic acid libraries derived from DNA, for example DNA directly isolated from the environment, are screened very rapidly for bioactivities of interest utilizing fluorescence activated cell sorting (FACS). These libraries can contain greater than 10^8 members and can represent single organisms or can represent the genomes of hundreds of different organisms, species or subspecies. In one aspect, the libraries, or clones of the libraries are “biopanned” as a step in the high throughput analysis.

15 Accordingly, in one embodiment, the present invention provides a process for identifying clones having a specified activity of interest, which process comprises (i) generating one or more expression libraries derived from nucleic acid directly isolated from the environment; and (ii) screening said libraries utilizing a high throughput cell analyzer, preferably a fluorescence activated cell sorter, to identify said clones.

20 More particularly, the invention provides a process for identifying clones having a specified activity of interest by (i) generating one or more libraries, *e.g.*, expression libraries, made to contain nucleic acid directly or indirectly isolated from the environment; (ii) exposing said libraries to a particular substrate or substrates of interest; and (iii) screening said exposed libraries utilizing a high throughput cell analyzer, preferably a fluorescence activated cell sorter, to identify clones which react with the substrate or substrates.

25 In another aspect, the invention also provides a process for identifying clones having a specified activity of interest by (i) generating one or more gene libraries derived from nucleic acid directly or indirectly isolated from the environment; and (ii) screening said exposed libraries utilizing an assay requiring a binding event or the covalent

modification of a target, and a high throughput cell analyzer, preferably a fluorescence activated cell sorter, to identify positive clones.

The invention further provides a method of screening for an agent that modulates the activity of a target protein or other cell component (e.g., nucleic acid), wherein the 5 target and a selectable marker are expressed by a recombinant cell, by co-encapsulating the agent in a microenvironment with the recombinant cell expressing the target and detectable marker and detecting the effect of the agent on the activity of the target cell component.

In another embodiment, the invention provides a method for enriching for target 10 DNA sequences containing at least a partial coding region for at least one specified activity in a DNA sample by co-encapsulating a mixture of target DNA obtained from a mixture of organisms with a mixture of DNA probes including a detectable marker and at least a portion of a DNA sequence encoding at least one enzyme having a specified enzyme activity and a detectable marker; incubating the co-encapsulated mixture under 15 such conditions and for such time as to allow hybridization of complementary sequences and screening for the target DNA. Optionally the method further comprises transforming host cells with recovered target DNA to produce an expression library of a plurality of clones.

The invention further provides a method of screening for an agent that modulates 20 the interaction of a first test protein linked to a DNA binding moiety and a second test protein linked to a transcriptional activation moiety by co-encapsulating the agent with the first test protein and second test protein in a suitable microenvironment and determining the ability of the agent to modulate the interaction of the first test protein linked to a DNA binding moiety with the second test protein covalently linked to a 25 transcriptional activation moiety, wherein the agent enhances or inhibits the expression of a detectable protein. Preferably, screening is by FACS analysis.

In yet another aspect, the present invention provides a method for identifying a 30 polynucleotide of interest or a molecule having an activity of interest, including contacting a library containing a plurality of clones which include nucleic acid derived from more than one organism or source, e.g., a mixed population of organisms, including

microorganisms or plant tissue, with at least one oligonucleotide probe labeled with a detectable, *e.g.*, fluorescent, molecule. The detectable molecule changes, *e.g.*, fluoresces, upon interaction of the probe to a target polynucleotide in the library. Clones from the library are then separated with an analyzer that detects the change in the 5 detectable molecule, *e.g.*, fluorescence. The separated clones can be contacted with a reporter system that identifies a polynucleotide encoding a polypeptide or a small molecule of interest, for example, and the clones capable of modulating expression or activity of the reporter system identified thereby identifying a polynucleotide of interest.

In another embodiment, the invention provides a method for identifying a 10 polynucleotide encoding a polypeptide of interest. The method includes co- encapsulating in a microenvironment a plurality of library clones containing DNA obtained from a mixed population of organisms with a mixture of oligonucleotide probes comprising a fluorescence marker and at least a portion of a polynucleotide sequence encoding a polypeptide of interest having a specified bioactivity. The encapsulated 15 clones are incubated under such conditions and for such time as to allow interaction of complementary sequences and clones containing a complement to the oligonucleotide probe encoding the polypeptide of interest identified by separating clones with a fluorescent analyzer that detects fluorescence.

In yet another embodiment, the invention provides a method for high throughput 20 screening of a polynucleotide library for a polynucleotide of interest that encodes a molecule of interest. The method includes contacting a library containing a plurality of clones comprising polynucleotides derived from a mixed population of organisms with a plurality of oligonucleotide probes labeled with a fluorescence molecule wherein said fluorescence molecule fluoresces upon interaction of the probe to a target polynucleotide 25 in the library; separating clones with a fluorescent analyzer that detects fluorescence; contacting the separated clones with a reporter system that identifies a polynucleotide encoding the molecule of interest; and identifying clones capable of modulating expression or activity of the reporter system thereby identifying a polynucleotide of interest.

In another embodiment, the invention provides a method of screening for a polynucleotide encoding an activity of interest. The method includes (a) obtaining polynucleotides from an environmental sample; (b) normalizing the polynucleotides obtained from the sample; (c) generating a library from the normalized polynucleotides; 5 (d) contacting the library with a plurality of oligonucleotide probes comprising a fluorescent marker and at least a portion of a polynucleotide sequence encoding a polypeptide of interest having a specified activity to select library clones positive for a sequence of interest; (e) selecting clones with a fluorescent analyzer that detects fluorescence; (f) contacting the selected clones with a reporter system that identifies a 10 polynucleotide encoding the activity of interest; and (g) identifying clones capable of modulating expression or activity of the reporter system thereby identifying a polynucleotide of interest; wherein the positive clones contain a polynucleotide sequence encoding an activity of interest which is capable of catalyzing the bioactive substrate.

In yet another embodiment, the present invention provides a method for 15 screening polynucleotides, comprising contacting a library of polynucleotides derived from a mixed population of organism with a probe oligonucleotide labeled with a fluorescence molecule, which fluoresce upon binding of the probe to a target polynucleotide of the library, to select library polynucleotides positive for a sequence of interest; separating library members that are positive for the sequence of interest with a 20 fluorescent analyzer that detects fluorescence; expressing the selected polynucleotides to obtain polypeptides; contacting the polypeptides with a reporter system; and identifying polynucleotides encoding polypeptides capable of modulating expression or activity of the reporter system.

In another embodiment, the invention provides a method for obtaining an 25 organism from a mixed population of organisms in a sample. The method includes encapsulating in a microenvironment at least one organism from the sample; incubating the encapsulated organism under such conditions and for such a time to allow the at least one microorganism to grow or proliferate; and sorting the encapsulated organism by flow cytometry to obtain an organism from the sample.

BRIEF DESCRIPTION OF THE FIGURES

Figure 1 illustrates the protocol used in the cell sorting method of the invention to screen for a polynucleotide of interest, in this case using a (library excised into *E. coli*). The clones of interest are isolated by sorting.

5 **Figure 2** shows a microtiter plate where clones or cells are sorted in accordance with the invention. Typically one cell or cells grown within a microdroplet are dispersed per well and grown up as clones.

10 **Figure 3** depicts a co-encapsulation assay. Cells containing library clones are coencapsulated with a substrate or labeled oligonucleotide. Encapsulation can occur in a variety of means, including GMDs, liposomes, and ghost cells. Cells are screened via high throughput screening on a fluorescence analyzer.

15 **Figure 4** depicts a side scatter versus forward scatter graph of FACS sorted gel-microdroplets (GMDs) containing a species of *Streptomyces* which forms unicells. Empty gel-microdroplets are distinguished from free cells and debris, also.

15 **Figure 5** is a depiction of a FACS/Biopanning method described herein and described in Example 3, below.

DETAILED DESCRIPTION OF THE INVENTION

The present invention provides a method for rapid sorting and screening of libraries derived from a mixed population of organisms from, for example, an 20 environmental sample or an uncultivated population of organisms. In one embodiment, gene libraries are generated, clones are either exposed to a substrate or substrate(s) of interest, or hybridized to a fluorescence labeled probe having a sequence corresponding to a sequence of interest and positive clones are identified and isolated via fluorescence activated cell sorting. Cells can be viable or non-viable during the process or at the end 25 of the process, as nucleic acids encoding a positive activity can be isolated and cloned utilizing techniques well known in the art.

This invention differs from fluorescence activated cell sorting, as normally performed, in several aspects. Previously, FACS machines have been employed in

studies focused on the analyses of eukaryotic and prokaryotic cell lines and cell culture processes. FACS has also been utilized to monitor production of foreign proteins in both eukaryotes and prokaryotes to study, for example, differential gene expression. The detection and counting capabilities of the FACS system have been applied in these 5 examples. However, FACS has never previously been employed in a discovery process to screen for and recover bioactivities in prokaryotes. Furthermore, the present invention does not require cells to survive, as do previously described technologies, since the desired nucleic acid (recombinant clones) can be obtained from alive or dead cells. For example, the cells only need to be viable long enough to contain, carry or 10 synthesize a complementary nucleic acid sequence to be detected, and can thereafter be either viable or non-viable cells so long as the complementary sequence remains intact. The present invention also solves problems that would have been associated with detection and sorting of *E. coli* expressing recombinant enzymes, and recovering encoding nucleic acids. Additionally, the present invention includes within its 15 embodiments any apparatus capable of detecting fluorescent wavelengths associated with biological material, such apparatuses are defined herein as fluorescent analyzers (one example of which is a FACS apparatus).

The use of a culture-independent approach to directly clone genes encoding novel enzymes from, for example, an environmental sample allows one to access 20 untapped resources of biodiversity. In one embodiment, the invention is based on the construction of “environmental libraries” which represent the collective genomes of naturally occurring organisms archived in cloning vectors that can be propagated in suitable prokaryotic hosts. Because the cloned DNA is initially extracted directly from environmental samples, the libraries are not limited to the small fraction of prokaryotes 25 that can be grown in pure culture. Additionally, a normalization of the environmental DNA present in these samples could allow more equal representation of the DNA from all of the species present in the original sample. This can dramatically increase the efficiency of finding interesting genes from minor constituents of the sample which may be under-represented by several orders of magnitude compared to the dominant species.

30 Prior to the present invention, the evaluation of complex environmental expression libraries was rate limiting. The present invention allows the rapid screening

of complex environmental libraries, containing, for example, genes from thousands of different organisms. The benefits of the present invention can be seen, for example, in screening a complex environmental sample. Screening of a complex sample previously required one to use labor intensive methods to screen several million clones to cover the 5 genomic biodiversity. The invention represents an extremely high-throughput screening method which allows one to assess this enormous number of clones. The method disclosed herein allows the screening anywhere from about 30 million to about 200 million clones per hour for a desired nucleic acid sequence or biological activity. This allows the thorough screening of environmental libraries for clones expressing novel 10 biomolecules.

The invention provides methods and composition whereby one can screen, sort or identify a polynucleotide sequence, polypeptide, or molecule of interest from a mixed population of organisms (e.g., organisms present in an environmental sample) based on 15 polynucleotide sequences present in the sample. Thus, the invention provides methods and compositions useful in screening organisms for a desired biological activity or biological sequence and to assist in obtaining sequences of interest that can further be used in directed evolution, molecular biology, biotechnology and industrial applications. By screening and identifying the nucleic acid sequences present in the sample, the invention increases the repertoire of available sequences that can be used for the 20 development of diagnostics, therapeutics or molecules for industrial applications. Accordingly, the methods of the invention can identify novel nucleic acid sequences encoding proteins or polypeptides having a desired biological activity.

Flow cytometry has been used in cloning and selection of variants from existing cell clones. This selection, however, has required stains that diffuse through cells 25 passively, rapidly and irreversibly, with no toxic effects or other influences on metabolic or physiological processes. Since, typically, flow sorting has been used to study animal cell culture performance, physiological state of cells, and the cell cycle, one goal of cell sorting has been to keep the cells viable during and after sorting.

There currently are no reports in the literature of screening and discovery of 30 polynucleotide sequence in libraries by fluorescence activated cell sorting. Furthermore

there are no reports of recovering DNA encoding bioactivities screened by FACS and additionally screening for a bioactivity of interest. The present invention provides these methods to allow the extremely rapid screening of viable or non-viable cells to recover desirable activities and the nucleic acid encoding those activities.

5 Fluorescence and other forms of staining have been employed for microbial discrimination and identification, and in the analysis of the interaction of drugs and antibiotics with microbial cells. Flow cytometry has been used in aquatic biology, where autofluorescence of photosynthetic pigments are used in the identification of algae or DNA stains are used to quantify and count marine populations (Davey and Kell, 1996).

10 Diaper and Edwards used flow cytometry to detect viable bacteria after staining with a range of fluorogenic esters including fluorescein diacetate (FDA) derivatives and CemChrome B, a stain sold commercially for the detection of viable bacteria in suspension (Diaper and Edwards, 1994). Labeled antibodies and oligonucleotide probes can also been used for these purposes.

15 Papers have been published describing the application of flow cytometry to the detection of native and recombinant enzymatic activities in eukaryotes. Betz *et al.* studied native (non-recombinant) lipase production by the eukaryote, *Rhizopus arrhizus* with flow cytometry. They found that spore suspensions of the mold were heterogeneous as judged by light-scattering data obtained with excitation at 633 nm, and they sorted clones of the subpopulations into the wells of microtiter plates. After 20 germination and growth, lipase production was automatically assayed (turbidimetrically) in the microtiter plates, and a representative set of the most active were reisolated, cultured, and assayed conventionally (Betz *et al.*, 1984). The ability of flow cytometry to make measurements on single cells means that individual cells with high levels of expression (*e.g.*, due to gene amplification or higher plasmid copy number) could be 25 detected.

30 Cells with chromogenic or fluorogenic substrates yield colored and fluorescent products, respectively. Previously, it had been thought that the flow cytometry-fluorescence activated cell sorter approaches could be of benefit only for the analysis of cells that contain intracellularly, or are normally physically associated with, the

enzymatic activity of a molecule of interest. On this basis, one could only use fluorogenic reagents which could penetrate the cell and which are thus potentially cytotoxic. In addition, gel microdroplets (GMDs) can be used during FACS sorting and culturing. The use of GMDs containing (physically) single cells which can take up 5 nutrients, secrete products, and grow to form colonies is useful in the present invention. The diffusional properties of GMDs may be made such that sufficient extracellular product remains associated with each individual GMD, so as to permit flow cytometric analysis and cell sorting on the basis of concentration of secreted molecule within each microdroplet. Beads have also been used to isolate mutants growing at different rates, 10 and to analyze antibody secretion by hybridoma cells and the nutrient sensitivity of hybridoma cells.

15 The GMD technology has had significance in amplifying the signals available in flow cytometric analysis, and in permitting the screening and sorting of microbial strains in strain improvement and isolation programs. GMD or other related technologies can be used in the present invention to localize, sort as well as amplify signals in the high throughput screening of recombinant libraries. Cell viability during the screening is not an issue or concern since nucleic acid can be recovered from the microdroplet.

20 Different types of encapsulation strategies and compounds or polymers can be used with the present invention. For instance, high temperature agaroses can be employed for making microdroplets stable at high temperatures, allowing stable encapsulation of cells subsequent to heat-kill steps utilized to remove all background activities when screening for thermostable bioactivities. Encapsulation can be in beads, high temperature agaroses, gel microdroplets, cells, such as ghost red blood cells or macrophages, liposomes, or any other means of encapsulating and localizing molecules.

25 For example, methods of preparing liposomes have been described (*i.e.*, U.S. Patent No.'s 5,653,996, 5393530 and 5,651,981), as well as the use of liposomes to encapsulate a variety of molecules U.S. Patent No.'s 5,595,756, 5,605,703, 5,627,159, 5,652,225, 5,567,433, 4,235,871, 5,227,170). Entrapment of proteins, viruses, 30 bacteria and DNA in erythrocytes during endocytosis has been described, as well (Journal of Applied Biochemistry 4, 418-435 (1982)). Erythrocytes employed as

carriers *in vitro* or *in vivo* for substances entrapped during hypo-osmotic lysis or dielectric breakdown of the membrane have also been described (reviewed in Ihler, G. M. (1983) *J. Pharm. Ther.*). These techniques are useful in the present invention to encapsulate samples for screening.

5

“Microenvironment”, as used herein, is any molecular structure which provides an appropriate environment for facilitating the interactions necessary for the method of the invention. An environment suitable for facilitating molecular interactions include, for example, liposomes. Liposomes can be prepared from a variety of lipids including 10 phospholipids, glycolipids, steroids, long-chain alkyl esters; *e.g.*, alkyl phosphates, fatty acid esters; *e.g.*, lecithin, fatty amines and the like. A mixture of fatty material may be employed such a combination of neutral steroid, a charge amphiphile and a phospholipid. Illustrative examples of phospholipids include lecithin, sphingomyelin and dipalmitoylphos-phatidylcholine. Representative steroids include cholesterol, 15 cholestanol and lanosterol. Representative charged amphiphilic compounds generally contain from 12-30 carbon atoms. Mono- or dialkyl phosphate esters, or alkyl amines; *e.g.*, dicetyl phosphate, stearyl amine, hexadecyl amine, dilauryl phosphate, and the like.

20

As used herein and in the appended claims, the singular forms “a,” “and,” and “the” include plural referents unless the context clearly dictates otherwise. Thus, for example, reference to “a clone” includes a plurality of clones and reference to “the nucleic acid sequence” generally includes reference to one or more nucleic acid sequences and equivalents thereof known to those skilled in the art, and so forth.

25

Unless defined otherwise, all technical and scientific terms used herein have the same meaning as commonly understood to one of ordinary skill in the art to which the invention belongs. Although any methods, devices and materials similar or equivalent to those described herein can be used in the practice or testing of the invention, the preferred methods, devices and materials are now described.

30

All publications mentioned herein are incorporated herein by reference in full for the purpose of describing and disclosing the databases, proteins, and methodologies,

which are described in the publications which might be used in connection with the presently described invention. The publications discussed above and throughout the text are provided solely for their disclosure prior to the filing date of the present application. Nothing herein is to be construed as an admission that the inventors are not entitled to 5 antedate such disclosure by virtue of prior invention.

An “amino acid” is a molecule having the structure wherein a central carbon atom (the α -carbon atom) is linked to a hydrogen atom, a carboxylic acid group (the carbon atom of which is referred to herein as a “carboxyl carbon atom”), an amino group (the nitrogen atom of which is referred to herein as an “amino nitrogen atom”), and a 10 side chain group, R. When incorporated into a peptide, polypeptide, or protein, an amino acid loses one or more atoms of its amino acid carboxylic groups in the dehydration reaction that links one amino acid to another. As a result, when incorporated into a protein, an amino acid is referred to as an “amino acid residue.”

“Protein” or “polypeptide” refers to any polymer of two or more individual 15 amino acids (whether or not naturally occurring) linked via a peptide bond, and occurs when the carboxyl carbon atom of the carboxylic acid group bonded to the α -carbon of one amino acid (or amino acid residue) becomes covalently bound to the amino nitrogen atom of amino group bonded to the α -carbon of an adjacent amino acid. The term “protein” is understood to include the terms “polypeptide” and “peptide” (which, at 20 times may be used interchangeably herein) within its meaning. In addition, proteins comprising multiple polypeptide subunits (e.g., DNA polymerase III, RNA polymerase II) or other components (for example, an RNA molecule, as occurs in telomerase) will also be understood to be included within the meaning of “protein” as used herein. Similarly, fragments of proteins and polypeptides are also within the scope of the 25 invention and may be referred to herein as “proteins.”

A particular amino acid sequence of a given protein (*i.e.*, the polypeptide’s “primary structure,” when written from the amino-terminus to carboxy-terminus) is determined by the nucleotide sequence of the coding portion of a mRNA, which is in turn specified by genetic information, typically genomic DNA (including organelle 30 DNA, *e.g.*, mitochondrial or chloroplast DNA). Thus, determining the sequence of a

gene assists in predicting the primary sequence of a corresponding polypeptide and more particular the role or activity of the polypeptide or proteins encoded by that gene or polynucleotide sequence.

The term "isolated" means altered "by the hand of man" from its natural state; 5 *i.e.*, if it occurs in nature, it has been changed or removed from its original environment, or both. For example, a naturally occurring polynucleotide or a polypeptide naturally present in a living animal, a biological sample or an environmental sample in its natural state is not "isolated", but the same polynucleotide or polypeptide separated from the coexisting materials of its natural state is "isolated", as the term is employed herein.

10 Such polynucleotides, when introduced into host cells in culture or in whole organisms, still would be isolated, as the term is used herein, because they would not be in their naturally occurring form or environment. Similarly, the polynucleotides and polypeptides may occur in a composition, such as a media formulation (solutions for introduction of polynucleotides or polypeptides, for example, into cells or compositions 15 or solutions for chemical or enzymatic reactions).

"Polynucleotide" or "nucleic acid sequence" refers to a polymeric form of nucleotides. In some instances a polynucleotide refers to a sequence that is not immediately contiguous with either of the coding sequences with which it is immediately contiguous (one on the 5' end and one on the 3' end) in the naturally occurring genome of the organism from which it is derived. The term therefore includes, 20 for example, a recombinant DNA which is incorporated into a vector; into an autonomously replicating plasmid or virus; or into the genomic DNA of a prokaryote or eukaryote, or which exists as a separate molecule (*e.g.*, a cDNA) independent of other sequences. The nucleotides of the invention can be ribonucleotides, 25 deoxyribonucleotides, or modified forms of either nucleotide. A polynucleotides as used herein refers to, among others, single-and double-stranded DNA, DNA that is a mixture of single- and double-stranded regions, single- and double-stranded RNA, and RNA that is mixture of single- and double-stranded regions, hybrid molecules comprising DNA and RNA that may be single-stranded or, more typically, double-stranded or a mixture of 30 single- and double-stranded regions.

In addition, polynucleotide as used herein refers to triple-stranded regions comprising RNA or DNA or both RNA and DNA. The strands in such regions may be from the same molecule or from different molecules. The regions may include all of one or more of the molecules, but more typically involve only a region of some of the 5 molecules. One of the molecules of a triple-helical region often is an oligonucleotide. The term polynucleotide encompasses genomic DNA or RNA (depending upon the organism, *i.e.*, RNA genome of viruses), as well as mRNA encoded by the genomic DNA, and cDNA.

As mentioned above, there is currently a need in the biotechnology and chemical 10 industry for molecules that can optimally carry out biological or chemical processes (*e.g.*, enzymes). Identifying novel enzymes in an environmental sample is one solution to this problem. By rapidly identifying polypeptides having an activity of interest and polynucleotides encoding the polypeptide of interest the invention provides methods, compositions and sources for the development of biologics, diagnostics, therapeutics, 15 and compositions for industrial applications.

All classes of molecules and compounds that are utilized in both established and emerging chemical, pharmaceutical, textile, food and feed, detergent markets must meet stringent economical and environmental standards. The synthesis of polymers, pharmaceuticals, natural products and agrochemicals is often hampered by expensive 20 processes which produce harmful byproducts and which suffer from poor or inefficient catalysis. Enzymes, for example, have a number of remarkable advantages which can overcome these problems in catalysis: they act on single functional groups, they distinguish between similar functional groups on a single molecule, and they distinguish between enantiomers. Moreover, they are biodegradable and function at very low mole 25 fractions in reaction mixtures. Because of their chemo-, regio- and stereospecificity, enzymes present a unique opportunity to optimally achieve desired selective transformations. These are often extremely difficult to duplicate chemically, especially in single-step reactions. The elimination of the need for protection groups, selectivity, the ability to carry out multi-step transformations in a single reaction vessel, along with 30 the concomitant reduction in environmental burden, has led to the increased demand for enzymes in chemical and pharmaceutical industries. Enzyme-based processes have been

gradually replacing many conventional chemical-based methods. A current limitation to more widespread industrial use is primarily due to the relatively small number of commercially available enzymes. Only ~300 enzymes (excluding DNA modifying enzymes) are at present commercially available from the > 3000 non DNA-modifying enzyme activities thus far described.

The use of enzymes for technological applications also may require performance under demanding industrial conditions. This includes activities in environments or on substrates for which the currently known arsenal of enzymes was not evolutionarily selected. However, the natural environment provides extreme conditions including, for example, extremes in temperature and pH. A number of organisms have adapted to these conditions due in part to selection for polypeptides than can withstand these extremes.

Enzymes have evolved by selective pressure to perform very specific biological functions within the milieu of a living organism, under conditions of temperature, pH and salt concentration. For the most part, the non-DNA modifying enzyme activities thus far described have been isolated from mesophilic organisms, which represent a very small fraction of the available phylogenetic diversity. The dynamic field of biocatalysis takes on a new dimension with the help of enzymes isolated from microorganisms that thrive in extreme environments. For example, such enzymes must function at temperatures above 100°C in terrestrial hot springs and deep sea thermal vents, at temperatures below 0°C in arctic waters, in the saturated salt environment of the Dead Sea, at pH values around 0 in coal deposits and geothermal sulfur-rich springs, or at pH values greater than 11 in sewage sludge. Environmental samples obtained, for example, from extreme conditions containing organisms, polynucleotides or polypeptides (*e.g.*, enzymes) open a new field in biocatalysis. By rapidly screening for polynucleotides encoding polypeptides of interest, the invention provides not only a source of materials for the development of biologics, therapeutics, and enzymes for industrial applications, but also provides a new materials for further processing by, for example, directed evolution and mutagenesis to develop molecules or polypeptides modified for particular activity or conditions.

In addition to the need for new enzymes for industrial use, there has been a dramatic increase in the need for bioactive compounds with novel activities. This demand has arisen largely from changes in worldwide demographics coupled with the clear and increasing trend in the number of pathogenic organisms that are resistant to 5 currently available antibiotics. For example, while there has been a surge in demand for antibacterial drugs in emerging nations with young populations, countries with aging populations, such as the U.S., require a growing repertoire of drugs against cancer, diabetes, arthritis and other debilitating conditions. The death rate from infectious diseases has increased 58% between 1980 and 1992 and it has been estimated that the 10 emergence of antibiotic resistant microbes has added in excess of \$30 billion annually to the cost of health care in the U.S. alone. (Adams *et al.*, *Chemical and Engineering News*, 1995; Amann *et al.*, *Microbiological Reviews*, 59, 1995). As a response to this trend pharmaceutical companies have significantly increased their screening of microbial diversity for compounds with unique activities or specificity. Accordingly, the invention 15 can be used to obtain and identify polynucleotides and related sequence specific information from, for example, infectious microorganisms present in the environment such as, for example, in the gut of various macroorganisms.

In another embodiment, the methods and compositions of the invention provide for the identification of lead drug compounds present in an environmental sample. The 20 methods of the invention provide the ability to mine the environment for novel drugs or identify related drugs contained in different microorganisms. There are several common sources of lead compounds (drug candidates), including natural product collections, synthetic chemical collections, and synthetic combinatorial chemical libraries, such as nucleotides, peptides, or other polymeric molecules that have been identified or 25 developed as a result of environmental mining. Each of these sources has advantages and disadvantages. The success of programs to screen these candidates depends largely on the number of compounds entering the programs, and pharmaceutical companies have to date screened hundred of thousands of synthetic and natural compounds in search of lead compounds. Unfortunately, the ratio of novel to previously-discovered 30 compounds has diminished with time. The discovery rate of novel lead compounds has not kept pace with demand despite the best efforts of pharmaceutical companies. There exists a strong need for accessing new sources of potential drug candidates.

Accordingly, the invention provides a rapid and efficient method to identify and characterize environmental samples that may contain novel drug compounds.

The majority of bioactive compounds currently in use are derived from soil microorganisms. Many microbes inhabiting soils and other complex ecological 5 communities produce a variety of compounds that increase their ability to survive and proliferate. These compounds are generally thought to be nonessential for growth of the organism and are synthesized with the aid of genes involved in intermediary metabolism hence their name – “secondary metabolites”. Secondary metabolites that influence the growth or survival of other organisms are known as “bioactive” compounds and serve as 10 key components of the chemical defense arsenal of both micro- and macro-organisms.

Humans have exploited these compounds for use as antibiotics, antiinfectives and other bioactive compounds with activity against a broad range of prokaryotic and eukaryotic pathogens. Approximately 6,000 bioactive compounds of microbial origin have been characterized, with more than 60% produced by the gram positive soil bacteria of the 15 genus *Streptomyces*. (Barnes *et al.*, *Proc.Nat. Acad. Sci. U.S.A.*, 91, 1994). Of these, at least 70 are currently used for biomedical and agricultural applications. The largest class of bioactive compounds, the polyketides, include a broad range of antibiotics, immunosuppressants and anticancer agents which together account for sales of over \$5 billion per year.

20 Despite the seemingly large number of available bioactive compounds, it is clear that one of the greatest challenges facing modern biomedical science is the proliferation of antibiotic resistant pathogens. Because of their short generation time and ability to readily exchange genetic information, pathogenic microbes have rapidly evolved and disseminated resistance mechanisms against virtually all classes of antibiotic 25 compounds. For example, there are virulent strains of the human pathogens *Staphylococcus* and *Streptococcus* that can now be treated with but a single antibiotic, vancomycin, and resistance to this compound will require only the transfer of a single gene, *vanA*, from resistant *Enterococcus* species for this to occur. (Bateson *et al.*, *System. Appl. Microbiol.*, 12, 1989). When this crucial need for novel antibacterial 30 compounds is superimposed on the growing demand for enzyme inhibitors, immunosuppressants and anti-cancer agents it becomes readily apparent why

pharmaceutical companies have stepped up their screening of microbial samples for bioactive compounds.

The invention provides methods of identifying a nucleic acid sequence encoding a polypeptide having either known or unknown function. For example, much of the 5 diversity in microbial genomes results from the rearrangement of gene clusters in the genome of microorganisms. These gene clusters can be present across species or phylogenetically related with other organisms.

For example, bacteria and many eukaryotes have a coordinated mechanism for regulating genes whose products are involved in related processes. The genes are 10 clustered, in structures referred to as "gene clusters," on a single chromosome and are transcribed together under the control of a single regulatory sequence, including a single promoter which initiates transcription of the entire cluster. The gene cluster, the promoter, and additional sequences that function in regulation altogether are referred to as an "operon" and can include up to 20 or more genes, usually from 2 to 6 genes. Thus, 15 a gene cluster is a group of adjacent genes that are either identical or related, usually as to their function.

Some gene families consist of identical members. Clustering is a prerequisite for maintaining identity between genes, although clustered genes are not necessarily identical. Gene clusters range from extremes where a duplication is generated to 20 adjacent related genes to cases where hundreds of identical genes lie in a tandem array. Sometimes no significance is discernable in a repetition of a particular gene. A principal example of this is the expressed duplicate insulin genes in some species, whereas a single insulin gene is adequate in other mammalian species.

Further, gene clusters undergo continual reorganization and, thus, the ability to 25 create heterogeneous libraries of gene clusters from, for example, bacterial or other prokaryote sources is valuable in determining sources of novel proteins, particularly including enzymes such as, for example, the polyketide synthases that are responsible for the synthesis of polyketides having a vast array of useful activities. Other types of proteins that are the product(s) of gene clusters are also contemplated, including, for 30 example, antibiotics, antivirals, antitumor agents and regulatory proteins, such as insulin.

As an example, polyketide synthases enzymes fall in a gene cluster. Polyketides are molecules which are an extremely rich source of bioactivities, including antibiotics (such as tetracyclines and erythromycin), anti-cancer agents (daunomycin), immunosuppressants (FK506 and rapamycin), and veterinary products (monensin).

5 Many polyketides (produced by polyketide synthases) are valuable as therapeutic agents. Polyketide synthases are multifunctional enzymes that catalyze the biosynthesis of a huge variety of carbon chains differing in length and patterns of functionality and cyclization. Polyketide synthase genes fall into gene clusters and at least one type (designated type I) of polyketide synthases have large size genes and enzymes, 10 complicating genetic manipulation and *in vitro* studies of these genes/proteins.

The ability to select and combine desired components from a library of polyketides and postpolyketide biosynthesis genes for generation of novel polyketides for study is appealing. The method(s) of the present invention make it possible to, and facilitate the cloning of, novel polyketide synthases, since one can generate gene banks 15 with clones containing large inserts (especially when using the f-factor based vectors), which facilitates cloning of gene clusters.

For example, a gene cluster can be ligated into a vector containing an expression regulatory sequences which can control and regulate the production of a detectable protein or protein-related array activity from the ligated gene clusters. Use of vectors 20 which have an exceptionally large capacity for exogenous nucleic acid introduction are particularly appropriate for use with such gene clusters and are described by way of example herein to include the f-factor (or fertility factor) of *E. coli*. This f-factor of *E. coli* is a plasmid which affects high-frequency transfer of itself during conjugation and is ideal to achieve and stably propagate large nucleic acid fragments, such as gene clusters 25 from mixed microbial samples.

The nucleic acid isolated or derived from these samples (e.g., a mixed population of microorganisms) can preferably be inserted into a vector or a plasmid prior to screening of the polynucleotides. Such vectors or plasmids are typically those containing expression regulatory sequences, including promoters, enhancers and the like.

Accordingly, the invention provides novel systems to clone and screen mixed populations of organisms present, for example, in an environmental samples, for polynucleotides of interest, enzymatic activities and bioactivities of interest *in vitro*. The method(s) of the invention allow the cloning and discovery of novel bioactive molecules 5 *in vitro*, and in particular novel bioactive molecules derived from uncultivated or cultivated samples. Large size gene clusters, genes and gene fragments can be cloned, sequenced and screened using the method(s) of the invention. Unlike previous strategies, the method(s) of the invention allow one to clone, screen and identify polynucleotides and the polypeptides encoded by these polynucleotides *in vitro* from a 10 wide range of environmental samples.

The invention allows one to screen for and identify polynucleotide sequences from complex environmental samples. DNA libraries obtained from these samples can be created from cell free samples, so long as the sample contains nucleic acid sequences, or from samples containing cellular organisms or viral particles. The organisms from 15 which the libraries may be prepared include prokaryotic microorganisms, such as *Eubacteria* and *Archaeobacteria*, lower eukaryotic microorganisms such as fungi, algae and protozoa, as well as mixed populations of plants, plant spores and pollen. The organisms may be cultured organisms or uncultured organisms obtained from environmental samples and includes extremophiles, such as thermophiles, 20 hyperthermophiles, psychrophiles and psychrotrophs.

Sources of nucleic acids used to construct a DNA library can be obtained from environmental samples, such as, but not limited to, microbial samples obtained from Arctic and Antarctic ice, water or permafrost sources, materials of volcanic origin, materials from soil or plant sources in tropical areas, droppings from various organisms 25 including mammals, invertebrates, as well as dead and decaying matter *etc*. Thus, for example, nucleic acids may be recovered from either a cultured or non-cultured organism and used to produce an appropriate DNA library (e.g., a recombinant expression library) for subsequent determination of the identity of the particular polynucleotide sequence or screening for enzyme activity.

The following outlines a general procedure for producing libraries from both culturable and non-culturable organisms as well as mixed population of organisms, which libraries can be probed, sequenced or screened to select therefrom nucleic acid sequences having an identified, desired or predicted biological activity (e.g., an enzymatic activity).

5 As used herein an environmental sample is any sample containing organisms or polynucleotides or a combination thereof. Thus, an environmental sample can be obtained from any number of sources (as described above), including, for example, insect feces, soil, water, *etc.* Any source of nucleic acids in purified or non-purified

10 form can be utilized as starting material. Thus, the nucleic acids may be obtained from any source which is contaminated by an organism or from any sample containing cells. The environmental sample can be an extract from any bodily sample such as blood, urine, spinal fluid, tissue, vaginal swab, stool, amniotic fluid or buccal mouthwash from any mammalian organism. For non-mammalian (e.g., invertebrates) organisms the

15 sample can be a tissue sample, salivary sample, fecal material or material in the digestive tract of the organism. An environmental sample also includes samples obtained from extreme environments including, for example, hot sulfur pools, volcanic vents, and frozen tundra. In addition, the sample can come from a variety of sources. For example, in horticulture and agricultural testing the sample can be a plant, fertilizer, soil, liquid or

20 other horticultural or agricultural product; in food testing the sample can be fresh food or processed food (for example infant formula, seafood, fresh produce and packaged food); and in environmental testing the sample can be liquid, soil, sewage treatment, sludge and any other sample in the environment which is considered or suspected of containing an organism or polynucleotides.

25 When the sample is a mixture of material (e.g., a mixed population of organisms), for example, blood, soil and sludge, it can be treated with an appropriate reagent which is effective to open the cells and expose or separate the strands of nucleic acids. Although not necessary, this lysing and nucleic acid denaturing step will allow cloning, amplification or sequencing to occur more readily. Further, if desired, the mixed

30 population can be cultured prior to analysis in order to purify a particular population and thus obtaining a purer sample. However, this is not necessary. For example, culturing of

organisms in the sample can include culturing the organisms in microdroplets and separating the cultured microdroplets with a cell sorter into individual wells of a multi-well tissue culture plate from which further processing may be performed.

Accordingly, the sample comprises nucleic acids from, for example, a diverse and mixed population of organisms (e.g., microorganisms present in the gut of an insect). Nucleic acids are isolated from the sample using any number of methods for DNA and RNA isolation. Such nucleic acid isolation methods are commonly performed in the art. Where the nucleic acid is RNA, the RNA can be reversed transcribed to DNA using primers known in the art. Where the DNA is genomic DNA, the DNA can be sheared using, for example, a 25 gauge needle.

The nucleic acids are then cloned into an appropriate vector. The vector used will depend upon whether the DNA is to be expressed, amplified, sequenced or manipulated in any number of ways known in the art (see, for example, U.S. Patent No. 6,022,716 which discloses high throughput sequencing vectors). Cloning techniques are known in the art or can be developed by one skilled in the art, without undue experimentation. The choice of a vector will also depend on the size of the polynucleotide sequence and the host cell to be employed in the methods of the invention. Thus, the vector used in the invention may be plasmids, phages, cosmids, phagemids, viruses (e.g., retroviruses, parainfluenzavirus, herpesviruses, reoviruses, paramyxoviruses, and the like), or selected portions thereof (e.g., coat protein, spike glycoprotein, capsid protein). For example, cosmids and phagemids are typically used where the specific nucleic acid sequence to be analyzed or modified is large because these vectors are able to stably propagate large polynucleotides.

The vector containing the cloned DNA sequence can then be amplified by plating (*i.e.*, clonal amplification) or transfecting a suitable host cell with the vector (*e.g.*, a phage on an *E. coli* host). Alternatively (or subsequently to amplification), the cloned DNA sequence is used to prepare a library for screening by transforming a suitable organism. Hosts, known in the art are transformed by artificial introduction of the vectors containing the target nucleic acid by inoculation under conditions conducive for such transformation. One could transform with double stranded circular or linear nucleic

acid or there may also be instances where one would transform with single stranded circular or linear nucleic acid sequences. By transform or transformation is meant a permanent or transient genetic change induced in a cell following incorporation of new DNA (*i.e.*, DNA exogenous to the cell). Where the cell is a mammalian cell, a
5 permanent genetic change is generally achieved by introduction of the DNA into the genome of the cell. A transformed cell or host cell generally refers to a cell (*e.g.*, prokaryotic or eukaryotic) into which (or into an ancestor of which) has been introduced, by means of recombinant DNA techniques, a DNA molecule not normally present in the host organism.

10 A particularly type of vector for use in the invention contains an f-factor origin replication. The f-factor (or fertility factor) in *E. coli* is a plasmid which effects high frequency transfer of itself during conjugation and less frequent transfer of the bacterial chromosome itself. In a particular embodiment cloning vectors referred to as "fosmids" or bacterial artificial chromosome (BAC) vectors are used. These are derived from *E. coli* f-factor which is able to stably integrate large segments of DNA. When integrated
15 with DNA from a mixed uncultured environmental sample, this makes it possible to achieve large genomic fragments in the form of a stable "environmental DNA library."

20 The nucleic acids derived from a mixed population or sample may be inserted into the vector by a variety of procedures. In general, the nucleic acid sequence is inserted into an appropriate restriction endonuclease site(s) by procedures known in the art. Such procedures and others are deemed to be within the scope of those skilled in the art. A typical cloning scenario may have the DNA "blunted" with an appropriate nuclelease (*e.g.*, Mung Bean Nuclease), methylated with, for example, *EcoR I* Methylase and ligated to *EcoR I* linkers GGAATTCC (SEQ ID NO:1). The linkers are then
25 digested with an *EcoR I* Restriction Endonuclease and the DNA size fractionated (*e.g.*, using a sucrose gradient). The resulting size fractionated DNA is then ligated into a suitable vector for sequencing, screening or expression (*e.g.*, a lambda vector and packaged using an *in vitro* lambda packaging extract).

30 Transformation of a host cell with recombinant DNA may be carried out by conventional techniques as are well known to those skilled in the art. Where the host is

prokaryotic, such as *E. coli*, competent cells which are capable of DNA uptake can be prepared from cells harvested after exponential growth phase and subsequently treated by the CaCl₂ method by procedures well known in the art. Alternatively, MgCl₂ or RbCl can be used. Transformation can also be performed after forming a protoplast of the host 5 cell or by electroporation.

When the host is a eukaryote, methods of transfection or transformation with DNA include calcium phosphate co-precipitates, conventional mechanical procedures such as microinjection, electroporation, insertion of a plasmid encased in liposomes, or virus vectors, as well as others known in the art, may be used. Eukaryotic cells can also 10 be cotransfected with a second foreign DNA molecule encoding a selectable marker, such as the herpes simplex thymidine kinase gene. Another method is to use a eukaryotic viral vector, such as simian virus 40 (SV40) or bovine papilloma virus, to transiently infect or transform eukaryotic cells and express the protein. (Eukaryotic Viral Vectors, Cold Spring Harbor Laboratory, Gluzman ed., 1982). The eukaryotic cell may 15 be a yeast cell (e.g., *Saccharomyces cerevisiae*), an insect cell (e.g., *Drosophila sp.*) or may be a mammalian cell, including a human cell.

Eukaryotic systems, and mammalian expression systems, allow for post-translational modifications of expressed mammalian proteins to occur. Eukaryotic cells which possess the cellular machinery for processing of the primary transcript, 20 glycosylation, phosphorylation, and, advantageously secretion of the gene product should be used. Such host cell lines may include, but are not limited to, CHO, VERO, BHK, HeLa, COS, MDCK, Jurkat, HEK-293, and WI38.

Biopanning

After the expression libraries have been generated one can perform “biopanning” of the libraries prior to expression screening. The “biopanning” procedure refers to a process for identifying clones having a specified biological activity by screening for

5 sequence homology in the library of clones, using at least one probe DNA comprising at least a portion of a DNA sequence encoding a polypeptide having the specified biological activity; and detecting interactions with the probe DNA to a substantially complementary sequence in a clone. Clones (either viable or non-viable) are then separated by a fluorescence analyzer (e.g., a FACS apparatus).

10 The probe DNA used to probe for the target DNA of interest contained in clones prepared from polynucleotides in a mixed population of organisms can be a full-length coding region sequence or a partial coding region sequence of DNA for a known bioactivity. The sequence of the probe can be generated by synthetic or recombinant means and can be based upon computer based sequencing programs or biological

15 sequences present in a clone. The DNA library can be probed using mixtures of probes comprising at least a portion of the DNA sequence encoding a known bioactivity having a desired activity. These probes or probe libraries are preferably single-stranded. The probes that are particularly suitable are those derived from DNA encoding bioactivities having an activity similar or identical to the specified bioactivity which is to be screened.

20 In another embodiment, the polynucleotides are contained in clones, the clones having been prepared from nucleic acid sequences of a mixed population of organisms, wherein the nucleic acid sequences are used to prepare a DNA library of the mixed population of organisms. The DNA library is screened for a sequence of interest by transfecting a host cell containing the library with at least one labeled nucleic acid

25 sequence which is all or a portion of a DNA sequence encoding a bioactivity having a desirable activity and separating the library clones containing the desirable sequence by fluorescent based analysis.

30 In another embodiment, *in vivo* biopanning may be performed utilizing a FACS-based machine. Complex gene libraries are constructed with vectors which contain elements which stabilize transcribed RNA. For example, the inclusion of

sequences which result in secondary structures such as hairpins which are designed to flank the transcribed regions of the RNA would serve to enhance their stability, thus increasing their half life within the cell. The probe molecules used in the biopanning process consist of oligonucleotides labeled with reporter molecules that only fluoresce upon binding of the probe to a target molecule. Various dyes or stains well known in the art, for example those described in "Practical Flow Cytometry", 1995 Wiley-Liss, Inc., Howard M. Shapiro, M.D., can be used to intercalate or associate with nucleic acid in order to "label" the oligonucleotides. These probes are introduced into the recombinant cells of the library using one of several transformation methods. The probe molecules interact or hybridize to the transcribed target mRNA or DNA resulting in DNA/RNA heteroduplex molecules or DNA/DNA duplex molecules. Binding of the probe to a target will yield a fluorescent signal which is detected and sorted by the FACS machine during the screening process.

The probe DNA should be at least about 10 bases and preferably at least 15 bases. In one embodiment, an entire coding region of one part of a pathway may be employed as a probe. Where the probe is hybridized to the target DNA in an *in vitro* system, conditions for the hybridization in which target DNA is selectively isolated by the use of at least one DNA probe will be designed to provide a hybridization stringency of at least about 50% sequence identity, more particularly a stringency providing for a sequence identity of at least about 70%. Hybridization techniques for probing a microbial DNA library to isolate target DNA of potential interest are well known in the art and any of those which are described in the literature are suitable for use herein. Prior to fluorescence sorting the clones may be viable or non-viable. For example, in one embodiment, the cells are fixed with paraformaldehyde prior to sorting.

Once viable or non-viable clones containing a sequence substantially complementary to the probe DNA are separated by a fluorescence analyzer, polynucleotides present in the separated clones may be further manipulated. In some instances, it may be desirable to perform an amplification of the target DNA that has been isolated. In this embodiment, the target DNA is separated from the probe DNA after isolation. In one embodiment, the clone can be grown to expand the clonal

population. Alternatively, the host cell is lysed and the target DNA amplified. It is then amplified before being used to transform a new host (e.g., subcloning). Long PCR (Barnes, W M, Proc. Natl. Acad. Sci, USA, Mar. 15, 1994) can be used to amplify large DNA fragments (e.g., 35 kb). Numerous amplification methodologies are now well known in the art.

5 Where the target DNA is identified *in vitro*, the selected DNA is then used for preparing a library for further processing and screening by transforming a suitable organism. Hosts, particularly those specifically identified herein as preferred, are transformed by artificial introduction of a vector containing a target DNA by inoculation 10 under conditions conducive for such transformation.

10 The resultant libraries (enriched for a polynucleotide of interest) can then be screened for clones which display an activity of interest. Clones can be shuttled in alternative hosts for expression of active compounds, or screened using methods described herein.

15 Having prepared a multiplicity of clones from DNA selectively isolated via FACS-based hybridization, such clones are screened for a specific activity to identify clones having a specified characteristic.

20 The screening for activity may be effected on individual expression clones or may be initially effected on a mixture of expression clones to ascertain whether or not the mixture has one or more specified activities. If the mixture has a specified activity, then the individual clones may be rescreened for such activity or for a more specific activity.

25 Prior to, subsequent to or as an alternative to the *in vivo* biopanning described above is an encapsulation techniques such as GMDs, which may be employed to localize at least one clone in one location for growth or screening by a fluorescent analyzer (e.g. FACS). The separated at least one clone contained in the GMD may then be cultured to expand the number of clones or screened on a FACS machine to identify clones containing a sequence of interest as described above, which can then be broken out into individual clones to be screened again on a FACS machine to identify positive individual

clones. Screening in this manner using a FACS machine is described in patent application Ser. No. 08/876,276, filed June 16, 1997. Thus, for example, if a clone has a desirable activity, then the individual clones may be recovered and rescreened utilizing a FACS machine to determine which of such clones has the specified desirable activity.

5 Further, it is possible to combine some or all of the above embodiments such that a normalization step is performed prior to generation of the expression library, the expression library is then generated, the expression library so generated is then biopanned, and the biopanned expression library is then screened using a high throughput cell sorting and screening instrument. Thus there are a variety of options, 10 including: (i) generating the library and then screening it; (ii) normalize the target DNA, generate the expression library and screen it; (iii) normalize, generate the library, biopan and screen; or (iv) generate, biopan and screen the library.

15 The library may, for example, be screened for a specified enzyme activity. For example, the enzyme activity screened for may be one or more of the six IUB classes; oxidoreductases, transferases, hydrolases, lyases, isomerases and ligases. The recombinant enzymes which are determined to be positive for one or more of the IUB classes may then be rescreened for a more specific enzyme activity.

20 Alternatively, the library may be screened for a more specialized enzyme activity. For example, instead of generically screening for hydrolase activity, the library may be screened for a more specialized activity, *i.e.* the type of bond on which the hydrolase acts. Thus, for example, the library may be screened to ascertain those hydrolases which act on one or more specified chemical functionalities, such as: (a) 25 amide (peptide bonds), *i.e.* proteases; (b) ester bonds, *i.e.* esterases and lipases; (c) acetals, *i.e.*, glycosidases *etc.*

30 As described with respect to one of the above aspects, the invention provides a process for activity screening of clones containing selected DNA derived from a mixed population of organisms or more than one organism.

Biopanning polynucleotides from a mixed population of organisms by separating the clones or polynucleotides positive for sequence of interest with a fluorescent analyzer that detects fluorescence, to select polynucleotides or clones containing polynucleotides positive for a sequence of interest, and screening the selected clones or 5 polynucleotides for specified bioactivity. In one embodiment, the polynucleotides are contained in clones having been prepared by recovering DNA of a microorganism, which DNA is selected by hybridization to at least one DNA sequence which is all or a portion of a DNA sequence encoding a bioactivity having a desirable activity.

In another embodiment, a DNA library derived from a microorganism is 10 subjected to a selection procedure to select therefrom DNA which hybridizes to one or more probe DNA sequences which is all or a portion of a DNA sequence encoding an activity having a desirable activity by:

(a) contacting a DNA library with a fluorescent labeled DNA probe under 15 conditions permissive of hybridization so as to produce a double-stranded complex of probe and members of the DNA library.

Screening

The present invention offers the ability to screen for many types of bioactivities. For instance, the ability to select and combine desired components from a library of 20 polyketides and postpolyketide biosynthesis genes for generation of novel polyketides for study is appealing. The method(s) of the present invention make it possible to and facilitate the cloning of novel polyketide synthases, and other relevant pathways or genes encoding commercially relevant secondary metabolites, since one can generate gene banks with clones containing large inserts (especially when using vectors which can accept large inserts, such as the f-factor based vectors), which facilitates cloning of gene 25 clusters.

The biopanning approach described above can be used to create libraries enriched with clones carrying sequences substantially homologous to a given probe sequence. Using this approach libraries containing clones with inserts of up to 40 kbp can be enriched approximately 1,000 fold after each round of panning. This enables one

to reduce the number of clones to be screened after 1 round of biopanning enrichment. This approach can be applied to create libraries enriched for clones carrying sequence of interest related to a bioactivity of interest, for example, polyketide sequences.

Hybridization screening using high density filters or biopanning has proven an efficient approach to detect homologues of pathways containing genes of interest to discover novel bioactive molecules that may have no known counterparts. Once a polynucleotide of interest is enriched in a library of clones it may be desirable to screen for an activity. For example, it may be desirable to screen for the expression of small molecule ring structures or "backbones". Because the genes encoding these polycyclic structures can often be expressed in *E. coli* the small molecule backbone can be manufactured albeit in an inactive form. Bioactivity is conferred upon transferring the molecule or pathway to an appropriate host that expresses the requisite glycosylation and methylation genes that can modify or "decorate" the structure to its active form. Thus, inactive ring compounds, recombinantly expressed in *E. coli* are detected to identify clones which are then shuttled to a metabolically rich host, such as *Streptomyces*, for subsequent production of the bioactive molecule. The use of high throughput robotic systems allows the screening of hundreds of thousands of clones in multiplexed arrays in microtiter dishes.

One approach to detect and enrich for clones carrying these structures is to use FACS screening, a procedure described and exemplified in U.S. Ser. No. 08/876,276, filed June 16, 1997. Polycyclic ring compounds typically have characteristic fluorescent spectra when excited by ultraviolet light. Thus, clones expressing these structures can be distinguished from background using a sufficiently sensitive detection method. High throughput FACS screening can be utilized to screen for small molecule backbones in *E. coli* libraries. Commercially available FACS machines are capable of screening up to 100,000 clones per second for UV active molecules. These clones can be sorted for further FACS screening or the resident plasmids can be extracted and shuttled to *Streptomyces* for activity screening.

In an alternate screening approach, after shuttling to *Streptomyces* hosts, organic extracts from candidate clones can be tested for bioactivity by susceptibility screening

against test organisms such as *Staphylococcus aureus*, *E. coli*, or *Saccharomyces cervisiae*. FACS screening can be used in this approach by co-encapsulating clones with the test organism.

An alternative to the above-mentioned screening methods provided by the 5 present invention is an approach termed "mixed extract" screening. The "mixed extract" screening approach takes advantage of the fact that the accessory genes needed to confer activity upon the polycyclic backbones are expressed in metabolically rich hosts, such as *Streptomyces*, and that the enzymes can be extracted and combined with the backbones extracted from *E. coli* clones to produce the bioactive compound *in vitro*. Enzyme 10 extract preparations from metabolically rich hosts, such as *Streptomyces* strains, at various growth stages are combined with pools of organic extracts from *E. coli* libraries and then evaluated for bioactivity.

Another approach to detect activity in the *E. coli* clones is to screen for genes 15 that can convert bioactive compounds to different forms. For example, a recombinant enzyme was recently discovered that can convert the low value daunomycin to the higher value doxorubicin. Similar enzyme pathways are being sought to convert penicillins to cephalosporins.

Screening may be carried out to detect a specified enzyme activity by procedures 20 known in the art. For example, enzyme activity may be screened for one or more of the six IUB classes; oxidoreductases, transferases, hydrolases, lyases, isomerases and ligases. The recombinant enzymes which are determined to be positive for one or more of the IUB classes may then be rescreened for a more specific enzyme activity.

Alternatively, the library may be screened for a more specialized enzyme activity. For 25 example, instead of generically screening for hydrolase activity, the library may be screened for a more specialized activity, *i.e.* the type of bond on which the hydrolase acts. Thus, for example, the library may be screened to ascertain those hydrolases which act on one or more specified chemical functionalities, such as: (a) amide (peptide bonds), *i.e.* proteases; (b) ester bonds, *i.e.* esterases and lipases; (c) acetals, *i.e.*, glycosidases.

FACS screening can also be used to detect expression of UV fluorescent 30 molecules in metabolically rich hosts, such as *Streptomyces*. Recombinant oxytetracylin

retains its diagnostic red fluorescence when produced heterologously in *S. lividans* TK24. Pathway clones, which can be sorted by FACS, can thus be screened for polycyclic molecules in a high throughput fashion.

Recombinant bioactive compounds can also be screened *in vivo* using “two-hybrid” systems, which can detect enhancers and inhibitors of protein-protein or other interactions such as those between transcription factors and their activators, or receptors and their cognate targets. In this embodiment, both the small molecule pathway and the GFP reporter construct are co-expressed. Clones altered in GFP expression can then be sorted by FACS and the pathway clone isolated for characterization.

As indicated, common approaches to drug discovery involve screening assays in which disease targets (macromolecules implicated in causing a disease) are exposed to potential drug candidates which are tested for therapeutic activity. In other approaches, whole cells or organisms that are representative of the causative agent of the disease, such as bacteria or tumor cell lines, are exposed to the potential candidates for screening purposes. Any of these approaches can be employed with the present invention.

The present invention also allows for the transfer of cloned pathways derived from uncultivated samples into metabolically rich hosts for heterologous expression and downstream screening for bioactive compounds of interest using a variety of screening approaches briefly described above.

20 *Recovering Desirable Bioactivities*

After viable or non-viable cells, each containing a different expression clone from the gene library are screened, and positive clones are recovered, DNA can be isolated from positive clones utilizing techniques well known in the art. The DNA can then be amplified either *in vivo* or *in vitro* by utilizing any of the various amplification techniques known in the art. *In vivo* amplification would include transformation of the clone(s) or subclone(s) into a viable host, followed by growth of the host. *In vitro* amplification can be performed using techniques such as the polymerase chain reaction. Once amplified the identified sequences can be “evolved” or sequenced.

Evolution

One advantage afforded by present invention is the ability to manipulate the identified polynucleotides to generate and select for encoded variants with altered activity or specificity.

Clones found to have the bioactivity for which the screen was performed can be
5 subjected to directed mutagenesis to develop new bioactivities with desired properties or to develop modified bioactivities with particularly desired properties that are absent or less pronounced in the wild-type activity, such as stability to heat or organic solvents. Any of the known techniques for directed mutagenesis are applicable to the invention. For example, particularly preferred mutagenesis techniques for use in accordance with
10 the invention include those described below.

Alternatively, it may be desirable to variegate a polynucleotide sequence obtained, identified or cloned as described herein. Such variegation can modify the polynucleotide sequence in order to modify (*e.g.*, increase or decrease) the encoded polypeptide's activity, specificity, affinity, function, *etc.* DNA shuffling can be used to
15 increase variation in a particular sample. DNA shuffling is meant to indicate recombination between substantially homologous but non-identical sequences, in some embodiments DNA shuffling may involve crossover via non-homologous recombination, such as via cer/lox and/or flp/frt systems and the like (see, for example, U.S. Patent No. 5,939,250, issued to Dr. Jay Short on August 17, 1999, and assigned to
20 Diversa Corporation, the disclosure of which is incorporated herein by reference). Various methods for shuffling, mutating or variegating polynucleotide sequences are discussed below.

Nucleic acid shuffling is a method for *in vitro* or *in vivo* homologous recombination of pools of shorter or smaller polynucleotides to produce a
25 polynucleotide or polynucleotides. Mixtures of related nucleic acid sequences or polynucleotides are subjected to sexual PCR to provide random polynucleotides, and reassembled to yield a library or mixed population of recombinant hybrid nucleic acid molecules or polynucleotides.

In contrast to cassette mutagenesis, only shuffling and error-prone PCR allow one to mutate a pool of sequences blindly (without sequence information other than primers).

5 The advantage of the mutagenic shuffling of the invention over error-prone PCR alone for repeated selection can best be explained as follows. Consider DNA shuffling as compared with error-prone PCR (not sexual PCR). The initial library of selected pooled sequences can consist of related sequences of diverse origin or can be derived by any type of mutagenesis (including shuffling) of a single gene. A
10 collection of selected sequences is obtained after the first round of activity selection. Shuffling allows the free combinatorial association of all of the related sequences, for example.

15 This method differs from error-prone PCR, in that it is an inverse chain reaction. In error-prone PCR, the number of polymerase start sites and the number of molecules grows exponentially. However, the sequence of the polymerase start sites and the sequence of the molecules remains essentially the same. In contrast, in nucleic acid reassembly or shuffling of random polynucleotides the number of start sites and the number (but not size) of the random polynucleotides decreases over time.
20 For polynucleotides derived from whole plasmids the theoretical endpoint is a single, large concatemeric molecule.

25 Since cross-overs occur at regions of homology, recombination will primarily occur between members of the same sequence family. This discourages combinations of sequences that are grossly incompatible (e.g., having different activities or specificities). It is contemplated that multiple families of sequences can be shuffled in the same reaction. Further, shuffling generally conserves the relative order.

30 Rare shufflants will contain a large number of the best molecules (e.g., highest activity or specificity) and these rare shufflants may be selected based on their superior activity or specificity.

A pool of 100 different polypeptide sequences can be permuted in up to 10^3 different ways. This large number of permutations cannot be represented in a single library of DNA sequences. Accordingly, it is contemplated that multiple cycles of DNA shuffling and selection may be required depending on the length of the sequence and the sequence diversity desired.

5 Error-prone PCR, in contrast, keeps all the selected sequences in the same relative orientation, generating a much smaller mutant cloud.

10 The template polynucleotide which may be used in the methods of the invention may be DNA or RNA. It may be of various lengths depending on the size of the gene or shorter or smaller polynucleotide to be recombined or reassembled. Preferably, the template polynucleotide is from 50 bp to 50 kb. It is contemplated that entire vectors containing the nucleic acid encoding the protein of interest can be used 15 in the methods of the invention, and in fact have been successfully used.

The template polynucleotide may be obtained by amplification using the PCR reaction (USPN 4,683,202 and USPN 4,683,195) or other amplification or cloning methods. However, the removal of free primers from the PCR products before 20 subjecting them to pooling of the PCR products and sexual PCR may provide more efficient results. Failure to adequately remove the primers from the original pool before sexual PCR can lead to a low frequency of crossover clones.

25 The template polynucleotide often is double-stranded. A double-stranded nucleic acid molecule is recommended to ensure that regions of the resulting single-stranded polynucleotides are complementary to each other and thus can hybridize to form a double-stranded molecule.

30 It is contemplated that single-stranded or double-stranded nucleic acid polynucleotides having regions of identity to the template polynucleotide and regions of heterology to the template polynucleotide may be added to the template

polynucleotide, at this step. It is also contemplated that two different but related polynucleotide templates can be mixed at this step.

The double-stranded polynucleotide template and any added double- or 5 single-stranded polynucleotides are subjected to sexual PCR which includes slowing or halting to provide a mixture of from about 5 bp to 5 kb or more. Preferably the size of the random polynucleotides is from about 10 bp to 1000 bp, more preferably the size of the polynucleotides is from about 20 bp to 500 bp.

10 Alternatively, it is also contemplated that double-stranded nucleic acid having multiple nicks may be used in the methods of the invention. A nick is a break in one strand of the double-stranded nucleic acid. The distance between such nicks is preferably 5 bp to 5 kb, more preferably between 10 bp to 1000 bp. This can provide areas of self-priming to produce shorter or smaller polynucleotides to be included 15 with the polynucleotides resulting from random primers, for example.

20 The concentration of any one specific polynucleotide will not be greater than 1% by weight of the total polynucleotides, more preferably the concentration of any one specific nucleic acid sequence will not be greater than 0.1% by weight of the total nucleic acid.

The number of different specific polynucleotides in the mixture will be at least about 100, preferably at least about 500, and more preferably at least about 1000.

25 At this step single-stranded or double-stranded polynucleotides, either synthetic or natural, may be added to the random double-stranded shorter or smaller polynucleotides in order to increase the heterogeneity of the mixture of polynucleotides.

30 It is also contemplated that populations of double-stranded randomly broken polynucleotides may be mixed or combined at this step with the polynucleotides from

the sexual PCR process and optionally subjected to one or more additional sexual PCR cycles.

Where insertion of mutations into the template polynucleotide is desired,
5 single-stranded or double-stranded polynucleotides having a region of identity to the template polynucleotide and a region of heterology to the template polynucleotide may be added in a 20 fold excess by weight as compared to the total nucleic acid, more preferably the single-stranded polynucleotides may be added in a 10 fold excess by weight as compared to the total nucleic acid.

10

Where a mixture of different but related template polynucleotides is desired, populations of polynucleotides from each of the templates may be combined at a ratio of less than about 1:100, more preferably the ratio is less than about 1:40. For example, a backcross of the wild-type polynucleotide with a population of mutated 15 polynucleotide may be desired to eliminate neutral mutations (e.g., mutations yielding an insubstantial alteration in the phenotypic property being selected for). In such an example, the ratio of randomly provided wild-type polynucleotides which may be added to the randomly provided sexual PCR cycle hybrid polynucleotides is approximately 1:1 to about 100:1, and more preferably from 1:1 to 40:1.

20

The mixed population of random polynucleotides are denatured to form single-stranded polynucleotides and then re-annealed. Only those single-stranded polynucleotides having regions of homology with other single-stranded polynucleotides will re-anneal.

25

The random polynucleotides may be denatured by heating. One skilled in the art could determine the conditions necessary to completely denature the double-stranded nucleic acid. Preferably the temperature is from 80 °C to 100 °C, more preferably the temperature is from 90 °C to 96 °C. other methods which may be used 30 to denature the polynucleotides include pressure and pH.

The polynucleotides may be re-annealed by cooling. Preferably the temperature is from 20 °C to 75 °C, more preferably the temperature is from 40 °C to 65 °C. If a high frequency of crossovers is needed based on an average of only 4 consecutive bases of homology, recombination can be forced by using a low 5 annealing temperature, although the process becomes more difficult. The degree of renaturation which occurs will depend on the degree of homology between the population of single-stranded polynucleotides.

Renaturation can be accelerated by the addition of polyethylene glycol 10 (“PEG”) or salt. The salt concentration is preferably from 0 mM to 200 mM, more preferably the salt concentration is from 10 mM to 100 mM. The salt may be KCl or NaCl. The concentration of PEG is preferably from 0% to 20%, more preferably from 5% to 10%.

15 The annealed polynucleotides are next incubated in the presence of a nucleic acid polymerase and dNTP's (*i.e.* dATP, dCTP, DGTP and dTTP). The nucleic acid polymerase may be the Klenow fragment, the Taq polymerase or any other DNA polymerase known in the art.

20 The approach to be used for the assembly depends on the minimum degree of homology that should still yield crossovers. If the areas of identity are large, Taq polymerase can be used with an annealing temperature of between 45-65 °C. If the areas of identity are small, Klenow polymerase can be used with an annealing temperature of between 20-30 °C. One skilled in the art could vary the temperature of 25 annealing to increase the number of cross-overs achieved.

The polymerase may be added to the random polynucleotides prior to annealing, simultaneously with annealing or after annealing.

30 The cycle of denaturation, renaturation and incubation in the presence of polymerase is referred to herein as shuffling or reassembly of the nucleic acid. This

cycle is repeated for a desired number of times. Preferably the cycle is repeated from 2 to 50 times, more preferably the sequence is repeated from 10 to 40 times.

The resulting nucleic acid is a larger double-stranded polynucleotide of from 5 about 50 bp to about 100 kb, preferably the larger polynucleotide is from 500 bp to 50 kb.

This larger polynucleotides may contain a number of copies of a polynucleotide having the same size as the template polynucleotide in tandem. This 10 concatemeric polynucleotide is then denatured into single copies of the template polynucleotide. The result will be a population of polynucleotides of approximately the same size as the template polynucleotide. The population will be a mixed population where single or double-stranded polynucleotides having an area of identity and an area of heterology have been added to the template polynucleotide prior to 15 shuffling. These polynucleotides are then cloned into the appropriate vector and the ligation mixture used to transform bacteria.

It is contemplated that the single polynucleotides may be obtained from the larger concatemeric polynucleotide by amplification of the single polynucleotide prior 20 to cloning by a variety of methods including PCR (USPN 4,683,195 and USPN 4,683,202), rather than by digestion of the concatemer.

The vector used for cloning is not critical provided that it will accept a polynucleotide of the desired size. If expression of the particular polynucleotide is 25 desired, the cloning vehicle should further comprise transcription and translation signals next to the site of insertion of the polynucleotide to allow expression of the polynucleotide in the host cell.

The resulting bacterial population will include a number of recombinant 30 polynucleotides having random mutations. This mixed population may be tested to identify the desired recombinant polynucleotides. The method of selection will depend on the polynucleotide desired.

For example, if a polynucleotide, identified by the methods of described herein, encodes a protein with a first binding affinity, subsequent mutated (e.g., shuffled) sequences having an increased binding efficiency to a ligand may be 5 desired. In such a case the proteins expressed by each of the portions of the polynucleotides in the population or library may be tested for their ability to bind to the ligand by methods known in the art (*i.e.* panning, affinity chromatography). If a polynucleotide which encodes for a protein with increased drug resistance is desired, the proteins expressed by each of the polynucleotides in the population or library may 10 be tested for their ability to confer drug resistance to the host organism. One skilled in the art, given knowledge of the desired protein, could readily test the population to identify polynucleotides which confer the desired properties onto the protein.

It is contemplated that one skilled in the art could use a phage display system 15 in which fragments of the protein are expressed as fusion proteins on the phage surface (Pharmacia, Milwaukee WI). The recombinant DNA molecules are cloned into the phage DNA at a site which results in the transcription of a fusion protein a portion of which is encoded by the recombinant DNA molecule. The phage containing the recombinant nucleic acid molecule undergoes replication and 20 transcription in the cell. The leader sequence of the fusion protein directs the transport of the fusion protein to the tip of the phage particle. Thus the fusion protein which is partially encoded by the recombinant DNA molecule is displayed on the phage particle for detection and selection by the methods described above.

It is further contemplated that a number of cycles of nucleic acid shuffling 25 may be conducted with polynucleotides from a sub-population of the first population, which sub-population contains DNA encoding the desired recombinant protein. In this manner, proteins with even higher binding affinities or enzymatic activity could be achieved.

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It is also contemplated that a number of cycles of nucleic acid shuffling may be conducted with a mixture of wild-type polynucleotides and a sub-population of

nucleic acid from the first or subsequent rounds of nucleic acid shuffling in order to remove any silent mutations from the sub-population.

Any source of nucleic acid, in a purified form can be utilized as the starting 5 nucleic acid. Thus the process may employ DNA or RNA including messenger RNA, which DNA or RNA may be single or double stranded. In addition, a DNA-RNA hybrid which contains one strand of each may be utilized. The nucleic acid sequence may be of various lengths depending on the size of the nucleic acid sequence to be mutated. Preferably the specific nucleic acid sequence is from 50 to 50000 base pairs. 10 It is contemplated that entire vectors containing the nucleic acid encoding the protein of interest may be used in the methods of the invention.

Any specific nucleic acid sequence can be used to produce the population of hybrids by the present process. It is only necessary that a small population of hybrid 15 sequences of the specific nucleic acid sequence exist or be available for the present process.

A population of specific nucleic acid sequences having mutations may be created by a number of different methods. Mutations may be created by error-prone 20 PCR. Error-prone PCR uses low-fidelity polymerization conditions to introduce a low level of point mutations randomly over a long sequence. Alternatively, mutations can be introduced into the template polynucleotide by oligonucleotide-directed mutagenesis. In oligonucleotide-directed mutagenesis, a short sequence of the polynucleotide is removed from the polynucleotide using restriction enzyme digestion 25 and is replaced with a synthetic polynucleotide in which various bases have been altered from the original sequence. The polynucleotide sequence can also be altered by chemical mutagenesis. Chemical mutagens include, for example, sodium bisulfite, nitrous acid, hydroxylamine, hydrazine or formic acid. other agents which are analogues of nucleotide precursors include nitrosoguanidine, 5-bromouracil, 30 2-aminopurine, or acridine. Generally, these agents are added to the PCR reaction in place of the nucleotide precursor thereby mutating the sequence. Intercalating agents such as proflavine, acriflavine, quinacrine and the like can also be used. Random

mutagenesis of the polynucleotide sequence can also be achieved by irradiation with X-rays or ultraviolet light. Generally, plasmid polynucleotides so mutagenized are introduced into *E. coli* and propagated as a pool or library of hybrid plasmids.

5 Alternatively, a small mixed population of specific nucleic acids may be found in nature in that they may consist of different alleles of the same gene or the same gene from different related species (*i.e.*, cognate genes). Alternatively, they may be related DNA sequences found within one species, for example, the immunoglobulin genes.

10

Once a mixed population of specific nucleic acid sequences is generated, the polynucleotides can be used directly or inserted into an appropriate cloning vector, using techniques well-known in the art.

15

The choice of vector depends on the size of the polynucleotide sequence and the host cell to be employed in the methods of the invention. The templates of the invention may be plasmids, phages, cosmids, phagemids, viruses (*e.g.*, retroviruses, parainfluenzavirus, herpesviruses, reoviruses, paramyxoviruses, and the like), or selected portions thereof (*e.g.*, coat protein, spike glycoprotein, capsid protein). For 20 example, cosmids and phagemids are preferred where the specific nucleic acid sequence to be mutated is larger because these vectors are able to stably propagate large polynucleotides.

25

If a mixed population of the specific nucleic acid sequence is cloned into a vector it can be clonally amplified. Utility can be readily determined by screening expressed polypeptides.

30

The DNA shuffling method of the invention can be performed blindly on a pool of unknown sequences. By adding to the reassembly mixture oligonucleotides (with ends that are homologous to the sequences being reassembled) any sequence mixture can be incorporated at any specific position into another sequence mixture. Thus, it is contemplated that mixtures of synthetic oligonucleotides, PCR

polynucleotides or even whole genes can be mixed into another sequence library at defined positions. The insertion of one sequence (mixture) is independent from the insertion of a sequence in another part of the template. Thus, the degree of recombination, the homology required, and the diversity of the library can be

5 independently and simultaneously varied along the length of the reassembled DNA.

Shuffling requires the presence of homologous regions separating regions of diversity. Scaffold-like protein structures may be particularly suitable for shuffling. The conserved scaffold determines the overall folding by self-association, while

10 displaying relatively unrestricted loops that mediate the specific binding. Examples of such scaffolds are the immunoglobulin beta-barrel, and the four-helix bundle which are well-known in the art. This shuffling can be used to create scaffold-like proteins with various combinations of mutated sequences for binding.

15 *In vitro Shuffling*

The equivalents of some standard genetic matings may also be performed by shuffling *in vitro*. For example, a “molecular backcross” can be performed by repeatedly mixing the hybrid's nucleic acid with the wild-type nucleic acid while

20 selecting for the mutations of interest. As in traditional breeding, this approach can be used to combine phenotypes from different sources into a background of choice. It is useful, for example, for the removal of neutral mutations that affect unselected characteristics (e.g., immunogenicity). Thus it can be useful to determine which mutations in a protein are involved in the enhanced biological activity and which are

25 not, an advantage which cannot be achieved by error-prone mutagenesis or cassette mutagenesis methods.

Large, functional genes can be assembled correctly from a mixture of small random polynucleotides. This reaction may be of use for the reassembly of genes

30 from the highly fragmented DNA of fossils. In addition random nucleic acid fragments from fossils may be combined with polynucleotides from similar genes from related species.

It is also contemplated that the method of the invention can be used for the *in vitro* amplification of a whole genome from a single cell as is needed for a variety of research and diagnostic applications. DNA amplification by PCR typically includes sequences of about 40 kb. Amplification of a whole genome such as that of *E. coli* (5, 000 kb) by PCR would require about 250 primers yielding 125 forty kb polynucleotides. On the other hand, random production of polynucleotides of the genome with sexual PCR cycles, followed by gel purification of small polynucleotides will provide a multitude of possible primers. Use of this mix of random small polynucleotides as primers in a PCR reaction alone or with the whole genome as the template should result in an inverse chain reaction with the theoretical endpoint of a single concatamer containing many copies of the genome.

A 100 fold amplification in the copy number and an average polynucleotide size of greater than 50 kb may be obtained when only random polynucleotides are used. It is thought that the larger concatamer is generated by overlap of many smaller polynucleotides. The quality of specific PCR products obtained using synthetic primers will be indistinguishable from the product obtained from unamplified DNA. It is expected that this approach will be useful for the mapping of genomes.

The polynucleotide to be shuffled can be produced as random or non-random polynucleotides, at the discretion of the practitioner. Moreover, the invention provides a method of shuffling that is applicable to a wide range of polynucleotide sizes and types, including the step of generating polynucleotide monomers to be used as building blocks in the reassembly of a larger polynucleotide. For example, the building blocks can be fragments of genes or they can be comprised of entire genes or gene pathways, or any combination thereof.

In vivo Shuffling

In an embodiment of *in vivo* shuffling, a mixed population of a specific nucleic acid sequence is introduced into bacterial or eukaryotic cells under conditions such that at least two different nucleic acid sequences are present in each host cell.

The polynucleotides can be introduced into the host cells by a variety of different methods. The host cells can be transformed with the smaller polynucleotides using methods known in the art, for example treatment with calcium chloride. If the polynucleotides are inserted into a phage genome, the host cell can be transfected with the recombinant phage genome having the specific nucleic acid sequences.

5 Alternatively, the nucleic acid sequences can be introduced into the host cell using electroporation, transfection, lipofection, biolistics, conjugation, and the like.

In general, in this embodiment, specific nucleic acid sequences will be present

10 in vectors which are capable of stably replicating the sequence in the host cell. In addition, it is contemplated that the vectors will encode a marker gene such that host cells having the vector can be selected. This ensures that the mutated specific nucleic acid sequence can be recovered after introduction into the host cell. However, it is contemplated that the entire mixed population of the specific nucleic acid sequences

15 need not be present on a vector sequence. Rather only a sufficient number of sequences need be cloned into vectors to ensure that after introduction of the polynucleotides into the host cells each host cell contains one vector having at least one specific nucleic acid sequence present therein. It is also contemplated that rather than having a subset of the population of the specific nucleic acids sequences cloned

20 into vectors, this subset may be already stably integrated into the host cell.

It has been found that when two polynucleotides which have regions of identity are inserted into the host cells homologous recombination occurs between the two polynucleotides. Such recombination between the two mutated specific nucleic acid sequences will result in the production of double or triple hybrids in some

25 situations.

It has also been found that the frequency of recombination is increased if some of the mutated specific nucleic acid sequences are present on linear nucleic acid molecules. Therefore, in a one embodiment, some of the specific nucleic acid sequences are present on linear polynucleotides.

After transformation, the host cell transformants are placed under selection to identify those host cell transformants which contain mutated specific nucleic acid sequences having the qualities desired. For example, if increased resistance to a particular drug is desired then the transformed host cells may be subjected to 5 increased concentrations of the particular drug and those transformants producing mutated proteins able to confer increased drug resistance will be selected. If the enhanced ability of a particular protein to bind to a receptor is desired, then expression of the protein can be induced from the transformants and the resulting protein assayed in a ligand binding assay by methods known in the art to identify that 10 subset of the mutated population which shows enhanced binding to the ligand. Alternatively, the protein can be expressed in another system to ensure proper processing.

Once a subset of the first recombined specific nucleic acid sequences 15 (daughter sequences) having the desired characteristics are identified, they are then subject to a second round of recombination. In the second cycle of recombination, the recombined specific nucleic acid sequences may be mixed with the original mutated specific nucleic acid sequences (parent sequences) and the cycle repeated as described above. In this way a set of second recombined specific nucleic acids sequences can 20 be identified which have enhanced characteristics or encode for proteins having enhanced properties. This cycle can be repeated a number of times as desired.

It is also contemplated that in the second or subsequent recombination cycle, a 25 backcross can be performed. A molecular backcross can be performed by mixing the desired specific nucleic acid sequences with a large number of the wild-type sequence, such that at least one wild-type nucleic acid sequence and a mutated nucleic acid sequence are present in the same host cell after transformation. Recombination with the wild-type specific nucleic acid sequence will eliminate those neutral 30 mutations that may affect unselected characteristics such as immunogenicity but not the selected characteristics.

In another embodiment of the invention, it is contemplated that during the first round a subset of specific nucleic acid sequences can be generated as smaller polynucleotides by slowing or halting their PCR amplification prior to introduction into the host cell. The size of the polynucleotides must be large enough to contain 5 some regions of identity with the other sequences so as to homologously recombine with the other sequences. The size of the polynucleotides will range from 0.03 kb to 100 kb more preferably from 0.2 kb to 10 kb. It is also contemplated that in subsequent rounds, all of the specific nucleic acid sequences other than the sequences selected from the previous round may be utilized to generate PCR polynucleotides 10 prior to introduction into the host cells.

The shorter polynucleotide sequences can be single-stranded or double-stranded. The reaction conditions suitable for separating the strands of nucleic acid are well known in the art.

15 The steps of this process can be repeated indefinitely, being limited only by the number of possible hybrids which can be achieved.

Therefore, the initial pool or population of mutated template nucleic acid is 20 cloned into a vector capable of replicating in a bacteria such as *E. coli*. The particular vector is not essential, so long as it is capable of autonomous replication in *E. coli*. In a one embodiment, the vector is designed to allow the expression and production of any protein encoded by the mutated specific nucleic acid linked to the vector. It is also preferred that the vector contain a gene encoding for a selectable marker.

25 The population of vectors containing the pool of mutated nucleic acid sequences is introduced into the *E. coli* host cells. The vector nucleic acid sequences may be introduced by transformation, transfection or infection in the case of phage. The concentration of vectors used to transform the bacteria is such that a number of 30 vectors is introduced into each cell. Once present in the cell, the efficiency of homologous recombination is such that homologous recombination occurs between the various vectors. This results in the generation of hybrids (daughters) having a

combination of mutations which differ from the original parent mutated sequences. The host cells are then clonally replicated and selected for the marker gene present on the vector. Only those cells having a plasmid will grow under the selection. The host cells which contain a vector are then tested for the presence of favorable mutations.

5

Once a particular daughter mutated nucleic acid sequence has been identified which confers the desired characteristics, the nucleic acid is isolated either already linked to the vector or separated from the vector. This nucleic acid is then mixed with the first or parent population of nucleic acids and the cycle is repeated.

10

The parent mutated specific nucleic acid population, either as polynucleotides or cloned into the same vector is introduced into the host cells already containing the daughter nucleic acids. Recombination is allowed to occur in the cells and the next generation of recombinants, or granddaughters are selected by the methods described 15 above. This cycle can be repeated a number of times until the nucleic acid or peptide having the desired characteristics is obtained. It is contemplated that in subsequent cycles, the population of mutated sequences which are added to the hybrids may come from the parental hybrids or any subsequent generation.

20

In an alternative embodiment, the invention provides a method of conducting a "molecular" backcross of the obtained recombinant specific nucleic acid in order to eliminate any neutral mutations. Neutral mutations are those mutations which do not confer onto the nucleic acid or peptide the desired properties. Such mutations may however confer on the nucleic acid or peptide undesirable characteristics.

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Accordingly, it is desirable to eliminate such neutral mutations. The method of the invention provide a means of doing so.

In this embodiment, after the hybrid nucleic acid, having the desired characteristics, is obtained by the methods of the embodiments, the nucleic acid, the 30 vector having the nucleic acid or the host cell containing the vector and nucleic acid is isolated.

The nucleic acid or vector is then introduced into the host cell with a large excess of the wild-type nucleic acid. The nucleic acid of the hybrid and the nucleic acid of the wild-type sequence are allowed to recombine. The resulting recombinants are placed under the same selection as the hybrid nucleic acid. Only those

5 recombinants which retained the desired characteristics will be selected. Any silent mutations which do not provide the desired characteristics will be lost through recombination with the wild-type DNA. This cycle can be repeated a number of times until all of the silent mutations are eliminated.

Exonuclease-Mediated Reassembly

In a another embodiment, the invention provides for a method for shuffling, assembling, reassembling, recombining, and/or concatenating at least two 5 polynucleotides to form a progeny polynucleotide (e.g., a chimeric progeny polynucleotide that can be expressed to produce a polypeptide or a gene pathway). In a particular embodiment, a double stranded polynucleotide (e.g., two single stranded sequences hybridized to each other as hybridization partners) is treated with an exonuclease to liberate nucleotides from one of the two strands, leaving the remaining 10 strand free of its original partner so that, if desired, the remaining strand may be used to achieve hybridization to another partner.

In a particular aspect, a double stranded polynucleotide end (that may be part of - or connected to - a polynucleotide or a nonpolynucleotide sequence) is subjected 15 to a source of exonuclease activity. Serviceable sources of exonuclease activity may be an enzyme with 3' exonuclease activity, an enzyme with 5' exonuclease activity, an enzyme with both 3' exonuclease activity and 5' exonuclease activity, and any combination thereof. An exonuclease can be used to liberate nucleotides from one or both ends of a linear double stranded polynucleotide, and from one to all ends of a 20 branched polynucleotide having more than two ends.

By contrast, a non-enzymatic step may be used to shuffle, assemble, reassemble, recombine, and/or concatenate polynucleotide building blocks that is comprised of subjecting a working sample to denaturing (or "melting") conditions 25 (for example, by changing temperature, pH, and /or salinity conditions) so as to melt a working set of double stranded polynucleotides into single polynucleotide strands. For shuffling, it is desirable that the single polynucleotide strands participate to some extent in annealment with different hybridization partners (*i.e.* and not merely revert to exclusive reannealment between what were former partners before the denaturation 30 step). The presence of the former hybridization partners in the reaction vessel, however, does not preclude, and may sometimes even favor, reannealment of a single stranded polynucleotide with its former partner, to recreate an original double stranded polynucleotide.

In contrast to this non-enzymatic shuffling step comprised of subjecting double stranded polynucleotide building blocks to denaturation, followed by annealment, the invention further provides an exonuclease-based approach requiring 5 no denaturation – rather, the avoidance of denaturing conditions and the maintenance of double stranded polynucleotide substrates in annealed (*i.e.* non-denatured) state are necessary conditions for the action of exonucleases (*e.g.*, exonuclease III and red alpha gene product). Additionally, in contrast, the generation of single stranded polynucleotide sequences capable of hybridizing to other single stranded 10 polynucleotide sequences is the result of covalent cleavage – and hence sequence destruction - in one of the hybridization partners. For example, an exonuclease III enzyme may be used to enzymatically liberate 3' terminal nucleotides in one hybridization strand (to achieve covalent hydrolysis in that polynucleotide strand); and this favors hybridization of the remaining single strand to a new partner (since its 15 former partner was subjected to covalent cleavage).

It is particularly appreciated that enzymes can be discovered, optimized (*e.g.*, engineered by directed evolution), or both discovered and optimized specifically for the instantly disclosed approach that have more optimal rates and/or more highly 20 specific activities &/or greater lack of unwanted activities. In fact it is expected that the invention may encourage the discovery and/or development of such designer enzymes.

Furthermore, it is appreciated that one can protect the end of a double stranded 25 polynucleotide or render it susceptible to a desired enzymatic action of a serviceable exonuclease as necessary. For example, a double stranded polynucleotide end having a 3' overhang is not susceptible to the exonuclease action of exonuclease III. However, it may be rendered susceptible to the exonuclease action of exonuclease III by a variety of means; for example, it may be blunted by treatment with a polymerase, 30 cleaved to provide a blunt end or a 5' overhang, joined (ligated or hybridized) to another double stranded polynucleotide to provide a blunt end or a 5' overhang,

hybridized to a single stranded polynucleotide to provide a blunt end or a 5' overhang, or modified by any of a variety of means).

According to one aspect, an exonuclease may be allowed to act on one or on 5 both ends of a linear double stranded polynucleotide and proceed to completion, to near completion, or to partial completion. When the exonuclease action is allowed to go to completion, the result will be that the length of each 5' overhang will be extend far towards the middle region of the polynucleotide in the direction of what might be considered a "rendezvous point" (which may be somewhere near the polynucleotide 10 midpoint). Ultimately, this results in the production of single stranded polynucleotides (that can become dissociated) that are each about half the length of the original double stranded polynucleotide.

Thus this exonuclease-mediated approach is serviceable for shuffling, 15 assembling and/or reassembling, recombining, and concatenating polynucleotide building blocks, which polynucleotide building blocks can be up to ten bases long or tens of bases long or hundreds of bases long or thousands of bases long or tens of thousands of bases long or hundreds of thousands of bases long or millions of bases long or even longer.

20 Substrates for an exonuclease may be generated by subjecting a double stranded polynucleotide to fragmentation. Fragmentation may be achieved by mechanical means (e.g., shearing, sonication, *etc.*), by enzymatic means (e.g., using restriction enzymes), and by any combination thereof. Fragments of a larger 25 polynucleotide may also be generated by polymerase-mediated synthesis.

Additional examples of enzymes with exonuclease activity include red-alpha and venom phosphodiesterases. Red alpha (*redα*) gene product (also referred to as lambda exonuclease) is of bacteriophage λ origin. Red alpha gene product acts 30 processively from 5'-phosphorylated termini to liberate mononucleotides from duplex DNA (Takahashi & Kobayashi, 1990). Venom phosphodiesterases (Laskowski, 1980) is capable of rapidly opening supercoiled DNA.

Non-stochastic Ligation Reassembly

In one aspect, the present invention provides a non-stochastic method termed synthetic ligation reassembly (SLR), that is somewhat related to stochastic shuffling, 5 save that the nucleic acid building blocks are not shuffled or concatenated or chimerized randomly, but rather are assembled non-stochastically.

The SLR method does not depend on the presence of a high level of homology between polynucleotides to be shuffled. The invention can be used to non- 10 stochastically generate libraries (or sets) of progeny molecules comprised of over 10^{100} different chimeras. Conceivably, SLR can even be used to generate libraries comprised of over 10^{1000} different progeny chimeras.

Thus, in one aspect, the invention provides a non-stochastic method of 15 producing a set of finalized chimeric nucleic acid molecules having an overall assembly order that is chosen by design, which method is comprised of the steps of generating by design a plurality of specific nucleic acid building blocks having serviceable mutually compatible ligatable ends, and assembling these nucleic acid building blocks, such that a designed overall assembly order is achieved.

20 The mutually compatible ligatable ends of the nucleic acid building blocks to be assembled are considered to be “serviceable” for this type of ordered assembly if they enable the building blocks to be coupled in predetermined orders. Thus, in one aspect, the overall assembly order in which the nucleic acid building blocks can be 25 coupled is specified by the design of the ligatable ends and, if more than one assembly step is to be used, then the overall assembly order in which the nucleic acid building blocks can be coupled is also specified by the sequential order of the assembly step(s). In a one embodiment of the invention, the annealed building pieces are treated with an enzyme, such as a ligase (e.g., T4 DNA ligase) to achieve covalent bonding of the 30 building pieces.

In a another embodiment, the design of nucleic acid building blocks is obtained upon analysis of the sequences of a set of progenitor nucleic acid templates

that serve as a basis for producing a progeny set of finalized chimeric nucleic acid molecules. These progenitor nucleic acid templates thus serve as a source of sequence information that aids in the design of the nucleic acid building blocks that are to be mutagenized, *i.e.* chimerized or shuffled.

5

In one exemplification, the invention provides for the chimerization of a family of related genes and their encoded family of related products. In a particular exemplification, the encoded products are enzymes. As a representative list of families of enzymes which may be mutagenized in accordance with the aspects of the 10 present invention, there may be mentioned, the following enzymes and their functions: Lipase/Esterase, Protease, Glycosidase/Glycosyl, transferase, Phosphatase/Kinase, Mono/Dioxygenase, Haloperoxidase, Lignin, peroxidase/Diarylpropane peroxidase, Epoxide hydrolase, Nitrile hydratase/nitrilase, 15 Transaminase, Amidase/Acylase. These exemplifications, while illustrating certain specific aspects of the invention, do not portray the limitations or circumscribe the scope of the disclosed invention.

Thus according to one aspect of the invention, the sequences of a plurality of progenitor nucleic acid templates identified using the methods of the invention are 20 aligned in order to select one or more demarcation points, which demarcation points can be located at an area of homology. The demarcation points can be used to delineate the boundaries of nucleic acid building blocks to be generated. Thus, the demarcation points identified and selected in the progenitor molecules serve as potential chimerization points in the assembly of the progeny molecules.

25

Typically a serviceable demarcation point is an area of homology (comprised of at least one homologous nucleotide base) shared by at least two progenitor templates, but the demarcation point can be an area of homology that is shared by at 30 least half of the progenitor templates, at least two thirds of the progenitor templates, at least three fourths of the progenitor templates, and preferably at almost all of the progenitor templates. Even more preferably still a serviceable demarcation point is an area of homology that is shared by all of the progenitor templates.

In a preferred embodiment, the ligation reassembly process is performed exhaustively in order to generate an exhaustive library. In other words, all possible ordered combinations of the nucleic acid building blocks are represented in the set of 5 finalized chimeric nucleic acid molecules. At the same time, the assembly order (*i.e.* the order of assembly of each building block in the 5' to 3' sequence of each finalized chimeric nucleic acid) in each combination is by design (or non-stochastic). Because of the non-stochastic nature of the invention, the possibility of unwanted side products is greatly reduced.

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In another preferred embodiment, the invention provides that, the ligation reassembly process is performed systematically, for example in order to generate a systematically compartmentalized library, with compartments that can be screened systematically, *e.g.*, one by one. In other words the invention provides that, through 15 the selective and judicious use of specific nucleic acid building blocks, coupled with the selective and judicious use of sequentially stepped assembly reactions, an experimental design can be achieved where specific sets of progeny products are made in each of several reaction vessels. This allows a systematic examination and screening procedure to be performed. Thus, it allows a potentially very large number 20 of progeny molecules to be examined systematically in smaller groups.

Because of its ability to perform chimerizations in a manner that is highly flexible yet exhaustive and systematic as well, particularly when there is a low level of homology among the progenitor molecules, the instant invention provides for the 25 generation of a library (or set) comprised of a large number of progeny molecules. Because of the non-stochastic nature of the instant ligation reassembly invention, the progeny molecules generated preferably comprise a library of finalized chimeric nucleic acid molecules having an overall assembly order that is chosen by design. In a particularly embodiment, such a generated library is comprised of greater than 10^3 30 to greater than 10^{1000} different progeny molecular species.

In one aspect, a set of finalized chimeric nucleic acid molecules, produced as described is comprised of a polynucleotide encoding a polypeptide. According to one embodiment, this polynucleotide is a gene, which may be a man-made gene. According to another embodiment, this polynucleotide is a gene pathway, which may 5 be a man-made gene pathway. The invention provides that one or more man-made genes generated by the invention may be incorporated into a man-made gene pathway, such as pathway operable in a eukaryotic organism (including a plant).

In another exemplification, the synthetic nature of the step in which the 10 building blocks are generated allows the design and introduction of nucleotides (e.g., one or more nucleotides, which may be, for example, codons or introns or regulatory sequences) that can later be optionally removed in an *in vitro* process (e.g., by mutagenesis) or in an *in vivo* process (e.g., by utilizing the gene splicing ability of a host organism). It is appreciated that in many instances the introduction of these 15 nucleotides may also be desirable for many other reasons in addition to the potential benefit of creating a serviceable demarcation point.

Thus, according to another embodiment, the invention provides that a nucleic acid building block can be used to introduce an intron. Thus, the invention provides 20 that functional introns may be introduced into a man-made gene of the invention. The invention also provides that functional introns may be introduced into a man-made gene pathway of the invention. Accordingly, the invention provides for the generation of a chimeric polynucleotide that is a man-made gene containing one (or more) artificially introduced intron(s).

25

Accordingly, the invention also provides for the generation of a chimeric polynucleotide that is a man-made gene pathway containing one (or more) artificially introduced intron(s). Preferably, the artificially introduced intron(s) are functional in 30 one or more host cells for gene splicing much in the way that naturally-occurring introns serve functionally in gene splicing. The invention provides a process of producing man-made intron-containing polynucleotides to be introduced into host organisms for recombination and/or splicing.

A man-made genes produced using the invention can also serve as a substrate for recombination with another nucleic acid. Likewise, a man-made gene pathway produced using the invention can also serve as a substrate for recombination with another nucleic acid. In a preferred instance, the recombination is facilitated by, or occurs at, areas of homology between the man-made intron-containing gene and a nucleic acid with serves as a recombination partner. In a particularly preferred instance, the recombination partner may also be a nucleic acid generated by the invention, including a man-made gene or a man-made gene pathway. Recombination may be facilitated by or may occur at areas of homology that exist at the one (or more) artificially introduced intron(s) in the man-made gene.

The synthetic ligation reassembly method of the invention utilizes a plurality of nucleic acid building blocks, each of which preferably has two ligatable ends. The two ligatable ends on each nucleic acid building block may be two blunt ends (*i.e.* each having an overhang of zero nucleotides), or preferably one blunt end and one overhang, or more preferably still two overhangs.

A serviceable overhang for this purpose may be a 3' overhang or a 5' overhang. Thus, a nucleic acid building block may have a 3' overhang or alternatively a 5' overhang or alternatively two 3' overhangs or alternatively two 5' overhangs. The overall order in which the nucleic acid building blocks are assembled to form a finalized chimeric nucleic acid molecule is determined by purposeful experimental design and is not random.

According to one preferred embodiment, a nucleic acid building block is generated by chemical synthesis of two single-stranded nucleic acids (also referred to as single-stranded oligos) and contacting them so as to allow them to anneal to form a double-stranded nucleic acid building block.

A double-stranded nucleic acid building block can be of variable size. The sizes of these building blocks can be small or large. Preferred sizes for building block

range from 1 base pair (not including any overhangs) to 100,000 base pairs (not including any overhangs). Other preferred size ranges are also provided, which have lower limits of from 1 bp to 10,000 bp (including every integer value in between), and upper limits of from 2 bp to 100, 000 bp (including every integer value in between).

5

Many methods exist by which a double-stranded nucleic acid building block can be generated that is serviceable for the invention; and these are known in the art and can be readily performed by the skilled artisan.

10 According to one embodiment, a double-stranded nucleic acid building block is generated by first generating two single stranded nucleic acids and allowing them to anneal to form a double-stranded nucleic acid building block. The two strands of a double-stranded nucleic acid building block may be complementary at every nucleotide apart from any that form an overhang; thus containing no mismatches, 15 apart from any overhang(s). According to another embodiment, the two strands of a double-stranded nucleic acid building block are complementary at fewer than every nucleotide apart from any that form an overhang. Thus, according to this embodiment, a double-stranded nucleic acid building block can be used to introduce codon degeneracy. Preferably the codon degeneracy is introduced using the site- 20 saturation mutagenesis described herein, using one or more N,N,G/T cassettes or alternatively using one or more N,N,N cassettes.

25 The *in vivo* recombination method of the invention can be performed blindly on a pool of unknown hybrids or alleles of a specific polynucleotide or sequence. However, it is not necessary to know the actual DNA or RNA sequence of the specific polynucleotide.

30 The approach of using recombination within a mixed population of genes can be useful for the generation of any useful proteins, for example, interleukin I, antibodies, tPA and growth hormone. This approach may be used to generate proteins having altered specificity or activity. The approach may also be useful for the generation of hybrid nucleic acid sequences, for example, promoter regions, introns,

exons, enhancer sequences, 31 untranslated regions or 51 untranslated regions of genes. Thus this approach may be used to generate genes having increased rates of expression. This approach may also be useful in the study of repetitive DNA sequences. Finally, this approach may be useful to mutate ribozymes or aptamers.

5 *End Selection*

The invention provides a method for selecting a subset of polynucleotides from a starting set of polynucleotides, which method is based on the ability to discriminate one or more selectable features (or selection markers) present anywhere 10 in a working polynucleotide, so as to allow one to perform selection for (positive selection) and/or against (negative selection) each selectable polynucleotide. In a preferred aspect, a method is provided termed end-selection, which method is based on the use of a selection marker located in part or entirely in a terminal region of a selectable polynucleotide, and such a selection marker may be termed an “end-selection marker”. 15

End-selection may be based on detection of naturally occurring sequences or on detection of sequences introduced experimentally (including by any mutagenesis procedure mentioned herein and not mentioned herein) or on both, even within the 20 same polynucleotide. An end-selection marker can be a structural selection marker or a functional selection marker or both a structural and a functional selection marker. An end-selection marker may be comprised of a polynucleotide sequence or of a polypeptide sequence or of any chemical structure or of any biological or biochemical tag, including markers that can be selected using methods based on the detection of 25 radioactivity, of enzymatic activity, of fluorescence, of any optical feature, of a magnetic property (e.g., using magnetic beads), of immunoreactivity, and of hybridization.

End-selection may be applied in combination with any method for performing 30 mutagenesis. Such mutagenesis methods include, but are not limited to, methods described herein (*supra* and *infra*). Such methods include, by way of non-limiting exemplification, any method that may be referred herein or by others in the art by any

of the following terms: “saturation mutagenesis”, “shuffling”, “recombination”, “re-assembly”, “error-prone PCR”, “assembly PCR”, “sexual PCR”, “crossover PCR”, “oligonucleotide primer-directed mutagenesis”, “recursive (and/or exponential) ensemble mutagenesis (see Arkin and Youvan, 1992)”, “cassette mutagenesis”, “*in vivo* mutagenesis”, and “*in vitro* mutagenesis”. Moreover, end-selection may be performed on molecules produced by any mutagenesis and/or amplification method (see, *e.g.*, Arnold, 1993; Caldwell and Joyce, 1992; Stemmer, 1994) following which method it is desirable to select for (including to screen for the presence of) desirable progeny molecules.

10

In addition, end-selection may be applied to a polynucleotide apart from any mutagenesis method. In a one embodiment, end-selection, as provided herein, can be used in order to facilitate a cloning step, such as a step of ligation to another polynucleotide (including ligation to a vector). The invention thus provides for end-15 selection as a serviceable means to facilitate library construction, selection and/or enrichment for desirable polynucleotides, and cloning in general.

In a another embodiment, end-selection can be based on (positive) selection for a polynucleotide; alternatively end-selection can be based on (negative) selection 20 against a polynucleotide; and alternatively still, end-selection can be based on both (positive) selection for, and on (negative) selection against, a polynucleotide. End-selection, along with other methods of selection and/or screening, can be performed in an iterative fashion, with any combination of like or unlike selection and/or screening methods and serviceable mutagenesis methods, all of which can be performed in an 25 iterative fashion and in any order, combination, and permutation. It is also appreciated that end-selection may also be used to select a polynucleotide in a: circular (*e.g.*, a plasmid or any other circular vector or any other polynucleotide that is partly circular), and/or branched, and/or modified or substituted with any chemical group or moiety.

30

In one non-limiting aspect, end-selection of a linear polynucleotide is performed using a general approach based on the presence of at least one end-

selection marker located at or near a polynucleotide end or terminus (that can be either a 5' end or a 3' end). In one particular non-limiting exemplification, end-selection is based on selection for a specific sequence at or near a terminus such as, but not limited to, a sequence recognized by an enzyme that recognizes a

5 polynucleotide sequence. An enzyme that recognizes and catalyzes a chemical modification of a polynucleotide is referred to herein as a polynucleotide-acting enzyme. In a preferred embodiment, serviceable polynucleotide-acting enzymes are exemplified non-exclusively by enzymes with polynucleotide-cleaving activity, enzymes with polynucleotide-methylating activity, enzymes with polynucleotide-

10 ligating activity, and enzymes with a plurality of distinguishable enzymatic activities (including non-exclusively, *e.g.*, both polynucleotide-cleaving activity and polynucleotide-ligating activity).

It is appreciated that relevant polynucleotide-acting enzymes include any

15 enzymes identifiable by one skilled in the art (*e.g.*, commercially available) or that may be developed in the future, though currently unavailable, that are serviceable for generating a ligation compatible end, preferably a sticky end, in a polynucleotide. It may be preferable to use restriction sites that are not contained, or alternatively that are not expected to be contained, or alternatively that are unlikely to be contained

20 (*e.g.*, when sequence information regarding a working polynucleotide is incomplete) internally in a polynucleotide to be subjected to end-selection. It is recognized that methods (*e.g.*, mutagenesis methods) can be used to remove unwanted internal restriction sites. It is also appreciated that a partial digestion reaction (*i.e.* a digestion reaction that proceeds to partial completion) can be used to achieve digestion at a

25 recognition site in a terminal region while sparing a susceptible restriction site that occurs internally in a polynucleotide and that is recognized by the same enzyme. In one aspect, partial digest are useful because it is appreciated that certain enzymes show preferential cleavage of the same recognition sequence depending on the location and environment in which the recognition sequence occurs.

30

It is also appreciated that protection methods can be used to selectively protect specified restriction sites (*e.g.*, internal sites) against unwanted digestion by enzymes

that would otherwise cut a working polypeptide in response to the presence of those sites; and that such protection methods include modifications such as methylations and base substitutions (*e.g.*, U instead of T) that inhibit an unwanted enzyme activity.

5 In another embodiment of the invention, a serviceable end-selection marker is a terminal sequence that is recognized by a polynucleotide-acting enzyme that recognizes a specific polynucleotide sequence. In one aspect of the invention, serviceable polynucleotide-acting enzymes also include other enzymes in addition to classic type II restriction enzymes. According to this preferred aspect of the
10 invention, serviceable polynucleotide-acting enzymes also include gyrases (*e.g.*, topoisomerases), helicases, recombinases, relaxases, and any enzymes related thereto.

It is appreciated that, end-selection can be used to distinguish and separate parental template molecules (*e.g.*, to be subjected to mutagenesis) from progeny molecules
15 (*e.g.*, generated by mutagenesis). For example, a first set of primers, lacking in a topoisomerase I recognition site, can be used to modify the terminal regions of the parental molecules (*e.g.*, in polymerase-based amplification). A different second set of primers (*e.g.*, having a topoisomerase I recognition site) can then be used to generate mutated progeny molecules (*e.g.*, using any polynucleotide chimerization method, such as
20 interrupted synthesis, template-switching polymerase-based amplification, or interrupted synthesis; or using saturation mutagenesis; or using any other method for introducing a topoisomerase I recognition site into a mutagenized progeny molecule) from the amplified template molecules. The use of topoisomerase I-based end-selection can then facilitate, not only discernment, but selective topoisomerase I-based ligation of the desired progeny
25 molecules.

It is appreciated that an end-selection approach using topoisomerase-based nicking and ligation has several advantages over previously available selection methods. In sum, this approach allows one to achieve direction cloning (including
30 expression cloning).

Peptide Display Methods

The present method can be used to shuffle, by *in vitro* and/or *in vivo* recombination by any of the disclosed methods, and in any combination,

5 polynucleotide sequences selected by peptide display methods, wherein an associated polynucleotide encodes a displayed peptide which is screened for a phenotype (*e.g.*, for affinity for a predetermined receptor (ligand).

An increasingly important aspect of bio-pharmaceutical drug development and
10 molecular biology is the identification of peptide structures, including the primary amino acid sequences, of peptides or peptidomimetics that interact with biological macromolecules. One method of identifying peptides that possess a desired structure or functional property, such as binding to a predetermined biological macromolecule (*e.g.*, a receptor), involves the screening of a large library of peptides for individual
15 library members which possess the desired structure or functional property conferred by the amino acid sequence of the peptide.

In addition to direct chemical synthesis methods for generating peptide libraries, several recombinant DNA methods also have been reported. One type
20 involves the display of a peptide sequence, antibody, or other protein on the surface of a bacteriophage particle or cell. Generally, in these methods each bacteriophage particle or cell serves as an individual library member displaying a single species of displayed peptide in addition to the natural bacteriophage or cell protein sequences. Each bacteriophage or cell contains the nucleotide sequence information encoding the
25 particular displayed peptide sequence; thus, the displayed peptide sequence can be ascertained by nucleotide sequence determination of an isolated library member.

A well-known peptide display method involves the presentation of a peptide sequence on the surface of a filamentous bacteriophage, typically as a fusion with a
30 bacteriophage coat protein. The bacteriophage library can be incubated with an immobilized, predetermined macromolecule or small molecule (*e.g.*, a receptor) so that bacteriophage particles which present a peptide sequence that binds to the

immobilized macromolecule can be differentially partitioned from those that do not present peptide sequences that bind to the predetermined macromolecule. The bacteriophage particles (*i.e.*, library members) which are bound to the immobilized macromolecule are then recovered and replicated to amplify the selected 5 bacteriophage sub-population for a subsequent round of affinity enrichment and phage replication. After several rounds of affinity enrichment and phage replication, the bacteriophage library members that are thus selected are isolated and the nucleotide sequence encoding the displayed peptide sequence is determined, thereby identifying the sequence(s) of peptides that bind to the predetermined macromolecule (*e.g.*, 10 receptor). Such methods are further described in PCT patent publications WO 91/17271, WO 91/18980, WO 91/19818 and WO 93/08278.

The present invention also provides random, pseudorandom, and defined sequence framework peptide libraries and methods for generating and screening those 15 libraries to identify useful compounds (*e.g.*, peptides, including single-chain antibodies) that bind to receptor molecules or epitopes of interest or gene products that modify peptides or RNA in a desired fashion. The random, pseudorandom, and defined sequence framework peptides are produced from libraries of peptide library members that comprise displayed peptides or displayed single-chain antibodies 20 attached to a polynucleotide template from which the displayed peptide was synthesized. The mode of attachment may vary according to the specific embodiment of the invention selected, and can include encapsulation in a phage particle or incorporation in a cell.

25 A significant advantage of the present invention is that no prior information regarding an expected ligand structure is required to isolate peptide ligands or antibodies of interest. The peptide identified can have biological activity, which is meant to include at least specific binding affinity for a selected receptor molecule and, in some instances, will further include the ability to block the binding of other 30 compounds, to stimulate or inhibit metabolic pathways, to act as a signal or messenger, to stimulate or inhibit cellular activity, and the like.

The invention also provides a method for shuffling a pool of polynucleotide sequences identified by the methods of the invention and selected by affinity screening a library of polysomes displaying nascent peptides (including single-chain antibodies) for library members which bind to a predetermined receptor (e.g., a 5 mammalian proteinaceous receptor such as, for example, a peptidergic hormone receptor, a cell surface receptor, an intracellular protein which binds to other protein(s) to form intracellular protein complexes such as hetero-dimers and the like) or epitope (e.g., an immobilized protein, glycoprotein, oligosaccharide, and the like).

10 Polynucleotide sequences selected in a first selection round (typically by affinity selection for binding to a receptor (e.g., a ligand)) by any of these methods are pooled and the pool(s) is/are shuffled by *in vitro* and/or *in vivo* recombination to produce a shuffled pool comprising a population of recombined selected polynucleotide sequences. The recombined selected polynucleotide sequences are 15 subjected to at least one subsequent selection round. The polynucleotide sequences selected in the subsequent selection round(s) can be used directly, sequenced, and/or subjected to one or more additional rounds of shuffling and subsequent selection. Selected sequences can also be back-crossed with polynucleotide sequences encoding neutral sequences (*i.e.*, having insubstantial functional effect on binding), such as for 20 example by back-crossing with a wild-type or naturally-occurring sequence substantially identical to a selected sequence to produce native-like functional peptides, which may be less immunogenic. Generally, during back-crossing subsequent selection is applied to retain the property of binding to the predetermined receptor (ligand).

25 Prior to or concomitant with the shuffling of selected sequences, the sequences can be mutagenized. In one embodiment, selected library members are cloned in a prokaryotic vector (e.g., plasmid, phagemid, or bacteriophage) wherein a collection of individual colonies (or plaques) representing discrete library members are produced. 30 Individual selected library members can then be manipulated (e.g., by site-directed mutagenesis, cassette mutagenesis, chemical mutagenesis, PCR mutagenesis, and the like) to generate a collection of library members representing a kernal of sequence

diversity based on the sequence of the selected library member. The sequence of an individual selected library member or pool can be manipulated to incorporate random mutation, pseudorandom mutation, defined kernal mutation (*i.e.*, comprising variant and invariant residue positions and/or comprising variant residue positions which can 5 comprise a residue selected from a defined subset of amino acid residues), codon-based mutation, and the like, either segmentally or over the entire length of the individual selected library member sequence. The mutagenized selected library members are then shuffled by *in vitro* and/or *in vivo* recombinatorial shuffling as disclosed herein.

10

The invention also provides peptide libraries comprising a plurality of individual library members of the invention, wherein (1) each individual library member of said plurality comprises a sequence produced by shuffling of a pool of selected sequences, and (2) each individual library member comprises a variable 15 peptide segment sequence or single-chain antibody segment sequence which is distinct from the variable peptide segment sequences or single-chain antibody sequences of other individual library members in said plurality (although some library members may be present in more than one copy per library due to uneven amplification, stochastic probability, or the like).

20

The invention also provides a product-by-process, wherein selected polynucleotide sequences having (or encoding a peptide having) a predetermined binding specificity are formed by the process of: (1) screening a displayed peptide or displayed single-chain antibody library against a predetermined receptor (*e.g.*, ligand) 25 or epitope (*e.g.*, antigen macromolecule) and identifying and/or enriching library members which bind to the predetermined receptor or epitope to produce a pool of selected library members, (2) shuffling by recombination the selected library members (or amplified or cloned copies thereof) which binds the predetermined epitope and has been thereby isolated and/or enriched from the library to generate a 30 shuffled library, and (3) screening the shuffled library against the predetermined receptor (*e.g.*, ligand) or epitope (*e.g.*, antigen macromolecule) and identifying and/or

enriching shuffled library members which bind to the predetermined receptor or epitope to produce a pool of selected shuffled library members.

Antibody Display and Screening Methods

5 The present method can be used to shuffle, by *in vitro* and/or *in vivo* recombination by any of the disclosed methods, and in any combination, polynucleotide sequences selected by antibody display methods, wherein an associated polynucleotide encodes a displayed antibody which is screened for a phenotype (e.g., for affinity for binding a predetermined antigen (ligand)).

10 Various molecular genetic approaches have been devised to capture the vast immunological repertoire represented by the extremely large number of distinct variable regions which can be present in immunoglobulin chains. The naturally-occurring germ line immunoglobulin heavy chain locus is composed of 15 separate tandem arrays of variable segment genes located upstream of a tandem array of diversity segment genes, which are themselves located upstream of a tandem array of joining (i) region genes, which are located upstream of the constant region genes. During B lymphocyte development, V-D-J rearrangement occurs wherein a heavy chain variable region gene (VH) is formed by rearrangement to form a fused D 20 segment followed by rearrangement with a V segment to form a V-D-J joined product gene which, if productively rearranged, encodes a functional variable region (VH) of a heavy chain. Similarly, light chain loci rearrange one of several V segments with one of several J segments to form a gene encoding the variable region (VL) of a light chain.

25 The vast repertoire of variable regions possible in immunoglobulins derives in part from the numerous combinatorial possibilities of joining V and i segments (and, in the case of heavy chain loci, D segments) during rearrangement in B cell development. Additional sequence diversity in the heavy chain variable regions arises 30 from non-uniform rearrangements of the D segments during V-D-J joining and from N region addition. Further, antigen-selection of specific B cell clones selects for higher affinity variants having non-germline mutations in one or both of the heavy

and light chain variable regions; a phenomenon referred to as “affinity maturation” or “affinity sharpening”. Typically, these “affinity sharpening” mutations cluster in specific areas of the variable region, most commonly in the complementarity-determining regions (CDRs).

5

In order to overcome many of the limitations in producing and identifying high-affinity immunoglobulins through antigen-stimulated B cell development (*i.e.*, immunization), various prokaryotic expression systems have been developed that can be manipulated to produce combinatorial antibody libraries which may be screened 10 for high-affinity antibodies to specific antigens. Recent advances in the expression of antibodies in *Escherichia coli* and bacteriophage systems (see “alternative peptide display methods”, *infra*) have raised the possibility that virtually any specificity can be obtained by either cloning antibody genes from characterized hybridomas or by de novo selection using antibody gene libraries (*e.g.*, from Ig cDNA).

15

Combinatorial libraries of antibodies have been generated in bacteriophage lambda expression systems which may be screened as bacteriophage plaques or as colonies of lysogens (Huse *et al.*, 1989); Caton and Koprowski, 1990; Mullinax *et al.*, 1990; Persson *et al.*, 1991). Various embodiments of bacteriophage antibody display 20 libraries and lambda phage expression libraries have been described (Kang *et al.*, 1991; Clackson *et al.*, 1991; McCafferty *et al.*, 1990; Burton *et al.*, 1991; Hoogenboom *et al.*, 1991; Chang *et al.*, 1991; Breitling *et al.*, 1991; Marks *et al.*, 1991, p. 581; Barbas *et al.*, 1992; Hawkins and Winter, 1992; Marks *et al.*, 1992, p. 25 779; Marks *et al.*, 1992, p. 16007; and Lowman *et al.*, 1991; Lerner *et al.*, 1992; all incorporated herein by reference). Typically, a bacteriophage antibody display library is screened with a receptor (*e.g.*, polypeptide, carbohydrate, glycoprotein, nucleic acid) that is immobilized (*e.g.*, by covalent linkage to a chromatography resin to enrich for reactive phage by affinity chromatography) and/or labeled (*e.g.*, to screen plaque or colony lifts).

30

One particularly advantageous approach has been the use of so-called single-chain fragment variable (scfv) libraries (Marks *et al.*, 1992, p. 779; Winter and

Milstein, 1991; Clackson *et al.*, 1991; Marks *et al.*, 1991, p. 581; Chaudhary *et al.*, 1990; Chiswell *et al.*, 1992; McCafferty *et al.*, 1990; and Huston *et al.*, 1988).

Various embodiments of scfv libraries displayed on bacteriophage coat proteins have been described.

5

Beginning in 1988, single-chain analogues of Fv fragments and their fusion proteins have been reliably generated by antibody engineering methods. The first step generally involves obtaining the genes encoding VH and VL domains with desired binding properties; these V genes may be isolated from a specific hybridoma cell line, 10 selected from a combinatorial V-gene library, or made by V gene synthesis. The single-chain Fv is formed by connecting the component V genes with an oligonucleotide that encodes an appropriately designed linker peptide, such as (Gly-Gly-Gly-Gly-Ser) or equivalent linker peptide(s). The linker bridges the C-terminus of the first V region and N-terminus of the second, ordered as either 15 VH-linker-VL or VL-linker-VH'. In principle, the scfv binding site can faithfully replicate both the affinity and specificity of its parent antibody combining site.

Thus, scfv fragments are comprised of VH and VL domains linked into a single polypeptide chain by a flexible linker peptide. After the scfv genes are 20 assembled, they are cloned into a phagemid and expressed at the tip of the M13 phage (or similar filamentous bacteriophage) as fusion proteins with the bacteriophage PIII (gene 3) coat protein. Enriching for phage expressing an antibody of interest is accomplished by panning the recombinant phage displaying a population scfv for binding to a predetermined epitope (e.g., target antigen, receptor).

25

The linked polynucleotide of a library member provides the basis for replication of the library member after a screening or selection procedure, and also provides the basis for the determination, by nucleotide sequencing, of the identity of the displayed peptide sequence or VH and VL amino acid sequence. The displayed 30 peptide (s) or single-chain antibody (e.g., scfv) and/or its VH and VL domains or their CDRs can be cloned and expressed in a suitable expression system. Often polynucleotides encoding the isolated VH and VL domains will be ligated to

polynucleotides encoding constant regions (CH and CL) to form polynucleotides encoding complete antibodies (e.g., chimeric or fully-human), antibody fragments, and the like. Often polynucleotides encoding the isolated CDRs will be grafted into polynucleotides encoding a suitable variable region framework (and optionally constant regions) to form polynucleotides encoding complete antibodies (e.g., humanized or fully-human), antibody fragments, and the like. Antibodies can be used to isolate preparative quantities of the antigen by immunoaffinity chromatography. Various other uses of such antibodies are to diagnose and/or stage disease (e.g., neoplasia) and for therapeutic application to treat disease, such as for example: neoplasia, autoimmune disease, AIDS, cardiovascular disease, infections, and the like.

Various methods have been reported for increasing the combinatorial diversity of a scfv library to broaden the repertoire of binding species (idiotype spectrum). The use of PCR has permitted the variable regions to be rapidly cloned either from a specific hybridoma source or as a gene library from non-immunized cells, affording combinatorial diversity in the assortment of VH and VL cassettes which can be combined. Furthermore, the VH and VL cassettes can themselves be diversified, such as by random, pseudorandom, or directed mutagenesis. Typically, VH and VL cassettes are diversified in or near the complementarity-determining regions (CDRS), often the third CDR, CDR3. Enzymatic inverse PCR mutagenesis has been shown to be a simple and reliable method for constructing relatively large libraries of scfv site-directed hybrids (Stemmer *et al.*, 1993), as has error-prone PCR and chemical mutagenesis (Deng *et al.*, 1994). Riechmann (Riechmann *et al.*, 1993) showed semi-rational design of an antibody scfv fragment using site-directed randomization by degenerate oligonucleotide PCR and subsequent phage display of the resultant scfv hybrids. Barbas (Barbas *et al.*, 1992) attempted to circumvent the problem of limited repertoire sizes resulting from using biased variable region sequences by randomizing the sequence in a synthetic CDR region of a human tetanus toxoid-binding Fab.

CDR randomization has the potential to create approximately 1×10^{20} CDRs for the heavy chain CDR3 alone, and a roughly similar number of variants of the heavy chain CDR1 and CDR2, and light chain CDR1-3 variants. Taken individually

or together, the combination possibilities of CDR randomization of heavy and/or light chains requires generating a prohibitive number of bacteriophage clones to produce a clone library representing all possible combinations, the vast majority of which will be non-binding. Generation of such large numbers of primary transformants is not feasible with current transformation technology and bacteriophage display systems. For example, Barbas (Barbas *et al.*, 1992) only generated 5×10^7 transformants, which represents only a tiny fraction of the potential diversity of a library of thoroughly randomized CDRS.

Despite these substantial limitations, bacteriophage display of scfv have already yielded a variety of useful antibodies and antibody fusion proteins. A bispecific single chain antibody has been shown to mediate efficient tumor cell lysis (Gruber *et al.*, 1994). Intracellular expression of an anti-Rev scfv has been shown to inhibit HIV-1 virus replication *in vitro* (Duan *et al.*, 1994), and intracellular expression of an anti-p21rar, scfv has been shown to inhibit meiotic maturation of *Xenopus* oocytes (Biocca *et al.*, 1993). Recombinant scfv which can be used to diagnose HIV infection have also been reported, demonstrating the diagnostic utility of scfv (Lilley *et al.*, 1994). Fusion proteins wherein an scFv is linked to a second polypeptide, such as a toxin or fibrinolytic activator protein, have also been reported (Holvost *et al.*, 1992; Nicholls *et al.*, 1993).

If it were possible to generate scfv libraries having broader antibody diversity and overcoming many of the limitations of conventional CDR mutagenesis and randomization methods which can cover only a very tiny fraction of the potential sequence combinations, the number and quality of scfv antibodies suitable for therapeutic and diagnostic use could be vastly improved. To address this, the *in vitro* and *in vivo* shuffling methods of the invention are used to recombine CDRs which have been obtained (typically via PCR amplification or cloning) from nucleic acids obtained from selected displayed antibodies. Such displayed antibodies can be displayed on cells, on bacteriophage particles, on polysomes, or any suitable antibody display system wherein the antibody is associated with its encoding nucleic acid(s). In a variation, the CDRs are initially obtained from mRNA (or cDNA) from

antibody-producing cells (e.g., plasma cells/splenocytes from an immunized wild-type mouse, a human, or a transgenic mouse capable of making a human antibody as in WO 92/03918, WO 93/12227, and WO 94/25585), including hybridomas derived therefrom.

5

Polynucleotide sequences selected in a first selection round (typically by affinity selection for displayed antibody binding to an antigen (e.g., a ligand) by any of these methods are pooled and the pool(s) is/are shuffled by *in vitro* and/or *in vivo* recombination, especially shuffling of CDRs (typically shuffling heavy chain CDRs with other heavy chain CDRs and light chain CDRs with other light chain CDRs) to produce a shuffled pool comprising a population of recombined selected polynucleotide sequences. The recombined selected polynucleotide sequences are expressed in a selection format as a displayed antibody and subjected to at least one subsequent selection round. The polynucleotide sequences selected in the subsequent selection round(s) can be used directly, sequenced, and/or subjected to one or more additional rounds of shuffling and subsequent selection until an antibody of the desired binding affinity is obtained. Selected sequences can also be back-crossed with polynucleotide sequences encoding neutral antibody framework sequences (*i.e.*, having insubstantial functional effect on antigen binding), such as for example by back-crossing with a human variable region framework to produce human-like sequence antibodies. Generally, during back-crossing subsequent selection is applied to retain the property of binding to the predetermined antigen.

Alternatively, or in combination with the noted variations, the valency of the target epitope may be varied to control the average binding affinity of selected scfv library members. The target epitope can be bound to a surface or substrate at varying densities, such as by including a competitor epitope, by dilution, or by other method known to those in the art. A high density (valency) of predetermined epitope can be used to enrich for scfv library members which have relatively low affinity, whereas a low density (valency) can preferentially enrich for higher affinity scfv library members.

For generating diverse variable segments, a collection of synthetic oligonucleotides encoding random, pseudorandom, or a defined sequence kernal set of peptide sequences can be inserted by ligation into a predetermined site (e.g., a CDR). Similarly, the sequence diversity of one or more CDRs of the single-chain antibody cassette(s) can be expanded by mutating the CDR(s) with site-directed mutagenesis, CDR-replacement, and the like. The resultant DNA molecules can be propagated in a host for cloning and amplification prior to shuffling, or can be used directly (i.e., may avoid loss of diversity which may occur upon propagation in a host cell) and the selected library members subsequently shuffled.

10

Displayed peptide/polynucleotide complexes (library members) which encode a variable segment peptide sequence of interest or a single-chain antibody of interest are selected from the library by an affinity enrichment technique. This is accomplished by means of a immobilized macromolecule or epitope specific for the peptide sequence of interest, such as a receptor, other macromolecule, or other epitope species. Repeating the affinity selection procedure provides an enrichment of library members encoding the desired sequences, which may then be isolated for pooling and shuffling, for sequencing, and/or for further propagation and affinity enrichment.

20

The library members without the desired specificity are removed by washing. The degree and stringency of washing required will be determined for each peptide sequence or single-chain antibody of interest and the immobilized predetermined macromolecule or epitope. A certain degree of control can be exerted over the binding characteristics of the nascent peptide/DNA complexes recovered by adjusting the conditions of the binding incubation and the subsequent washing. The temperature, pH, ionic strength, divalent cations concentration, and the volume and duration of the washing will select for nascent peptide/DNA complexes within particular ranges of affinity for the immobilized macromolecule. Selection based on slow dissociation rate, which is usually predictive of high affinity, is often the most practical route. This may be done either by continued incubation in the presence of a saturating amount of free predetermined macromolecule, or by increasing the volume, number, and length of the washes. In each case, the rebinding of dissociated nascent

peptide/DNA or peptide/RNA complex is prevented, and with increasing time, nascent peptide/DNA or peptide/RNA complexes of higher and higher affinity are recovered.

5 Additional modifications of the binding and washing procedures may be applied to find peptides with special characteristics. The affinities of some peptides are dependent on ionic strength or cation concentration. This is a useful characteristic for peptides that will be used in affinity purification of various proteins when gentle conditions for removing the protein from the peptides are required.

10

One variation involves the use of multiple binding targets (multiple epitope species, multiple receptor species), such that a scfv library can be simultaneously screened for a multiplicity of scfv which have different binding specificities. Given that the size of a scfv library often limits the diversity of potential scfv sequences, it is typically desirable to use scfv libraries of as large a size as possible. The time and economic considerations of generating a number of very large polysome scFv-display libraries can become prohibitive. To avoid this substantial problem, multiple predetermined epitope species (receptor species) can be concomitantly screened in a single library, or sequential screening against a number of epitope species can be used. In one variation, multiple target epitope species, each encoded on a separate bead (or subset of beads), can be mixed and incubated with a polysome-display scfv library under suitable binding conditions. The collection of beads, comprising multiple epitope species, can then be used to isolate, by affinity selection, scfv library members. Generally, subsequent affinity screening rounds can include the same mixture of beads, subsets thereof, or beads containing only one or two individual epitope species. This approach affords efficient screening, and is compatible with laboratory automation, batch processing, and high throughput screening methods.

30 A variety of techniques can be used in the present invention to diversify a peptide library or single-chain antibody library, or to diversify, prior to or concomitant with shuffling, around variable segment peptides found in early rounds of panning to have sufficient binding activity to the predetermined macromolecule or

epitope. In one approach, the positive selected peptide/polynucleotide complexes (those identified in an early round of affinity enrichment) are sequenced to determine the identity of the active peptides. Oligonucleotides are then synthesized based on these active peptide sequences, employing a low level of all bases incorporated at 5 each step to produce slight variations of the primary oligonucleotide sequences. This mixture of (slightly) degenerate oligonucleotides is then cloned into the variable segment sequences at the appropriate locations. This method produces systematic, controlled variations of the starting peptide sequences, which can then be shuffled. It requires, however, that individual positive nascent peptide/polynucleotide complexes 10 be sequenced before mutagenesis, and thus is useful for expanding the diversity of small numbers of recovered complexes and selecting variants having higher binding affinity and/or higher binding specificity. In a variation, mutagenic PCR amplification of positive selected peptide/polynucleotide complexes (especially of the variable region sequences, the amplification products of which are shuffled *in vitro* 15 and/or *in vivo* and one or more additional rounds of screening is done prior to sequencing. The same general approach can be employed with single-chain antibodies in order to expand the diversity and enhance the binding affinity/specificity, typically by diversifying CDRs or adjacent framework regions prior to or concomitant with shuffling. If desired, shuffling reactions can be spiked 20 with mutagenic oligonucleotides capable of *in vitro* recombination with the selected library members can be included. Thus, mixtures of synthetic oligonucleotides and PCR produced polynucleotides (synthesized by error-prone or high-fidelity methods) can be added to the *in vitro* shuffling mix and be incorporated into resulting shuffled library members (shufflants).

25

The invention of shuffling enables the generation of a vast library of CDR-variant single-chain antibodies. One way to generate such antibodies is to insert synthetic CDRs into the single-chain antibody and/or CDR randomization prior to or concomitant with shuffling. The sequences of the synthetic CDR cassettes are 30 selected by referring to known sequence data of human CDR and are selected in the discretion of the practitioner according to the following guidelines: synthetic CDRs will have at least 40 percent positional sequence identity to known CDR sequences,

and preferably will have at least 50 to 70 percent positional sequence identity to known CDR sequences. For example, a collection of synthetic CDR sequences can be generated by synthesizing a collection of oligonucleotide sequences on the basis of naturally-occurring human CDR sequences listed in Kabat (Kabat *et al.*, 1991); the 5 pool (s) of synthetic CDR sequences are calculated to encode CDR peptide sequences having at least 40 percent sequence identity to at least one known naturally-occurring human CDR sequence. Alternatively, a collection of naturally-occurring CDR sequences may be compared to generate consensus sequences so that amino acids used at a residue position frequently (*i.e.*, in at least 5 percent of known CDR 10 sequences) are incorporated into the synthetic CDRs at the corresponding position(s). Typically, several (*e.g.*, 3 to about 50) known CDR sequences are compared and observed natural sequence variations between the known CDRs are tabulated, and a collection of oligonucleotides encoding CDR peptide sequences encompassing all or most permutations of the observed natural sequence variations is synthesized. For 15 example but not for limitation, if a collection of human VH CDR sequences have carboxy-terminal amino acids which are either Tyr, Val, Phe, or Asp, then the pool(s) of synthetic CDR oligonucleotide sequences are designed to allow the carboxy-terminal CDR residue to be any of these amino acids. In some embodiments, residues other than those which naturally-occur at a residue position in the collection 20 of CDR sequences are incorporated: conservative amino acid substitutions are frequently incorporated and up to 5 residue positions may be varied to incorporate non-conservative amino acid substitutions as compared to known naturally-occurring CDR sequences. Such CDR sequences can be used in primary library members (prior to first round screening) and/or can be used to spike *in vitro* shuffling reactions of 25 selected library member sequences. Construction of such pools of defined and/or degenerate sequences will be readily accomplished by those of ordinary skill in the art.

The collection of synthetic CDR sequences comprises at least one member 30 that is not known to be a naturally-occurring CDR sequence. It is within the discretion of the practitioner to include or not include a portion of random or pseudorandom sequence corresponding to N region addition in the heavy chain CDR;

the N region sequence ranges from 1 nucleotide to about 4 nucleotides occurring at V-D and D-J junctions. A collection of synthetic heavy chain CDR sequences comprises at least about 100 unique CDR sequences, typically at least about 1,000 unique CDR sequences, preferably at least about 10,000 unique CDR sequences,
5 frequently more than 50,000 unique CDR sequences; however, usually not more than about 1×10^6 unique CDR sequences are included in the collection, although occasionally 1×10^7 to 1×10^8 unique CDR sequences are present, especially if conservative amino acid substitutions are permitted at positions where the conservative amino acid substituent is not present or is rare (*i.e.*, less than 0.1 percent)
10 in that position in naturally-occurring human CDRS. In general, the number of unique CDR sequences included in a library should not exceed the expected number of primary transformants in the library by more than a factor of 10. Such single-chain antibodies generally bind of about at least 1×10^6 M⁻¹, preferably with an affinity of about at least 5×10^7 M⁻¹, more preferably with an affinity of at least 1×10^8 M⁻¹ to
15 1×10^9 M⁻¹ or more, sometimes up to 1×10^{10} M⁻¹ or more. Frequently, the predetermined antigen is a human protein, such as for example a human cell surface antigen (*e.g.*, CD4, CD8, IL-2 receptor, EGF receptor, PDGF receptor), other human biological macromolecule (*e.g.*, thrombomodulin, protein C, carbohydrate antigen, sialyl Lewis antigen, Lselectin), or nonhuman disease associated macromolecule (*e.g.*,
20 bacterial LPS, virion capsid protein or envelope glycoprotein) and the like.

High affinity single-chain antibodies of the desired specificity can be engineered and expressed in a variety of systems. For example, scfv have been produced in plants (Firek *et al.*, 1993) and can be readily made in prokaryotic systems
25 (Owens and Young, 1994; Johnson and Bird, 1991). Furthermore, the single-chain antibodies can be used as a basis for constructing whole antibodies or various fragments thereof (Kettleborough *et al.*, 1994). The variable region encoding sequence may be isolated (*e.g.*, by PCR amplification or subcloning) and spliced to a sequence encoding a desired human constant region to encode a human sequence
30 antibody more suitable for human therapeutic uses where immunogenicity is preferably minimized. The polynucleotide(s) having the resultant fully human

encoding sequence(s) can be expressed in a host cell (e.g., from an expression vector in a mammalian cell) and purified for pharmaceutical formulation.

Once expressed, the antibodies, individual mutated immunoglobulin chains, 5 mutated antibody fragments, and other immunoglobulin polypeptides of the invention can be purified according to standard procedures of the art, including ammonium sulfate precipitation, fraction column chromatography, gel electrophoresis and the like (see, generally, Scopes, 1982). Once purified, partially or to homogeneity as desired, the polypeptides may then be used therapeutically or in developing and performing 10 assay procedures, immunofluorescent stainings, and the like (see, generally, Lefkovits and Pernis, 1979 and 1981; Lefkovits, 1997).

The antibodies generated by the method of the present invention can be used for diagnosis and therapy. By way of illustration and not limitation, they can be used 15 to treat cancer, autoimmune diseases, or viral infections. For treatment of cancer, the antibodies will typically bind to an antigen expressed preferentially on cancer cells, such as erbB-2, CEA, CD33, and many other antigens and binding members well known to those skilled in the art.

Two-Hybrid Based Screening Assays

Shuffling can also be used to recombinatorially diversify a pool of selected library members obtained by screening a two-hybrid screening system to identify 5 library members which bind a predetermined polypeptide sequence. The selected library members are pooled and shuffled by *in vitro* and/or *in vivo* recombination. The shuffled pool can then be screened in a yeast two hybrid system to select library members which bind said predetermined polypeptide sequence (e.g., an SH2 domain) or which bind an alternate predetermined polypeptide sequence (e.g., an SH2 10 domain from another protein species).

An approach to identifying polypeptide sequences which bind to a predetermined polypeptide sequence has been to use a so-called “two-hybrid” system wherein the predetermined polypeptide sequence is present in a fusion protein (Chien 15 *et al.*, 1991). This approach identifies protein-protein interactions *in vivo* through reconstitution of a transcriptional activator (Fields and Song, 1989), the yeast Gal4 transcription protein. Typically, the method is based on the properties of the yeast Gal4 protein, which consists of separable domains responsible for DNA-binding and transcriptional activation. Polynucleotides encoding two hybrid proteins, one 20 consisting of the yeast Gal4 DNA-binding domain fused to a polypeptide sequence of a known protein and the other consisting of the Gal4 activation domain fused to a polypeptide sequence of a second protein, are constructed and introduced into a yeast host cell. Intermolecular binding between the two fusion proteins reconstitutes the Gal4 DNA-binding domain with the Gal4 activation domain, which leads to the 25 transcriptional activation of a reporter gene (e.g., *lacZ*, *HIS3*) which is operably linked to a Gal4 binding site. Typically, the two-hybrid method is used to identify novel polypeptide sequences which interact with a known protein (Silver and Hunt, 1993; Durfee *et al.*, 1993; Yang *et al.*, 1992; Luban *et al.*, 1993; Hardy *et al.*, 1992; Bartel *et* 30 *al.*, 1993; and Vojtek *et al.*, 1993). However, variations of the two-hybrid method have been used to identify mutations of a known protein that affect its binding to a second known protein (Li and Fields, 1993; Lalo *et al.*, 1993; Jackson *et al.*, 1993; and Madura *et al.*, 1993). Two-hybrid systems have also been used to identify

interacting structural domains of two known proteins (Bardwell *et al.*, 1993; Chakrabarty *et al.*, 1992; Staudinger *et al.*, 1993; and Milne and Weaver 1993) or domains responsible for oligomerization of a single protein (Iwabuchi *et al.*, 1993; Bogerd *et al.*, 1993). Variations of two-hybrid systems have been used to study the *in vivo* activity of a proteolytic enzyme (Dasmahapatra *et al.*, 1992). Alternatively, an *E. coli*/BCCP interactive screening system (Germino *et al.*, 1993; Guarente, 1993) can be used to identify interacting protein sequences (*i.e.*, protein sequences which heterodimerize or form higher order heteromultimers). Sequences selected by a two-hybrid system can be pooled and shuffled and introduced into a two-hybrid system for one or more subsequent rounds of screening to identify polypeptide sequences which bind to the hybrid containing the predetermined binding sequence. The sequences thus identified can be compared to identify consensus sequence(s) and consensus sequence kernels.

One microgram samples of template DNA are obtained and treated with U.V. light to cause the formation of dimers, including TT dimers, particularly purine dimers. U.V. exposure is limited so that only a few photoproducts are generated per gene on the template DNA sample. Multiple samples are treated with U.V. light for varying periods of time to obtain template DNA samples with varying numbers of dimers from U.V. exposure.

A random priming kit which utilizes a non-proofreading polymerase (for example, Prime-It II Random Primer Labeling kit by Stratagene Cloning Systems) is utilized to generate different size polynucleotides by priming at random sites on templates which are prepared by U.V. light (as described above) and extending along the templates. The priming protocols such as described in the Prime-It II Random Primer Labeling kit may be utilized to extend the primers. The dimers formed by U.V. exposure serve as a roadblock for the extension by the non-proofreading polymerase. Thus, a pool of random size polynucleotides is present after extension with the random primers is finished.

The invention is further directed to a method for generating a selected mutant polynucleotide sequence (or a population of selected polynucleotide sequences) typically in the form of amplified and/or cloned polynucleotides, whereby the selected polynucleotide sequences(s) possess at least one desired phenotypic characteristic

5 (e.g., encodes a polypeptide, promotes transcription of linked polynucleotides, binds a protein, and the like) which can be selected for. One method for identifying hybrid polypeptides that possess a desired structure or functional property, such as binding to a predetermined biological macromolecule (e.g., a receptor), involves the screening of a large library of polypeptides for individual library members which possess the

10 desired structure or functional property conferred by the amino acid sequence of the polypeptide.

In one embodiment, the present invention provides a method for generating libraries of displayed polypeptides or displayed antibodies suitable for affinity interaction screening or phenotypic screening. The method comprises (1) obtaining a first plurality of selected library members comprising a displayed polypeptide or displayed antibody and an associated polynucleotide encoding said displayed polypeptide or displayed antibody, and obtaining said associated polynucleotides or copies thereof wherein said associated polynucleotides comprise a region of

15 substantially identical sequences, optimally introducing mutations into said polynucleotides or copies, (2) pooling the polynucleotides or copies, (3) producing smaller or shorter polynucleotides by interrupting a random or particularized priming and synthesis process or an amplification process, and (4) performing amplification, preferably PCR amplification, and optionally mutagenesis to homologously

20 recombine the newly synthesized polynucleotides.

25

It is an object of the invention to provide a process for producing hybrid polynucleotides which express a useful hybrid polypeptide by a series of steps comprising:

30 (a) producing polynucleotides by interrupting a polynucleotide amplification or synthesis process with a means for blocking or interrupting the amplification or synthesis process and thus providing a plurality of smaller or shorter

polynucleotides due to the replication of the polynucleotide being in various stages of completion;

- (b) adding to the resultant population of single- or double-stranded polynucleotides one or more single- or double-stranded oligonucleotides, wherein said 5 added oligonucleotides comprise an area of identity in an area of heterology to one or more of the single- or double-stranded polynucleotides of the population;
- (c) denaturing the resulting single- or double-stranded oligonucleotides to produce a mixture of single-stranded polynucleotides, optionally separating the shorter or smaller polynucleotides into pools of polynucleotides having various 10 lengths and further optionally subjecting said polynucleotides to a PCR procedure to amplify one or more oligonucleotides comprised by at least one of said polynucleotide pools;
- (d) incubating a plurality of said polynucleotides or at least one pool of said polynucleotides with a polymerase under conditions which result in annealing of 15 said single-stranded polynucleotides at regions of identity between the single-stranded polynucleotides and thus forming of a mutagenized double-stranded polynucleotide chain;
- (e) optionally repeating steps (c) and (d);
- (f) expressing at least one hybrid polypeptide from said polynucleotide 20 chain, or chains; and
- (g) screening said at least one hybrid polypeptide for a useful activity.

In a preferred aspect of the invention, the means for blocking or interrupting the amplification or synthesis process is by utilization of uv light, DNA adducts, DNA binding proteins.

25

In one embodiment of the invention, the DNA adducts, or polynucleotides comprising the DNA adducts, are removed from the polynucleotides or polynucleotide pool, such as by a process including heating the solution comprising the DNA fragments prior to further processing.

30

Sequencing

The clones enriched for a desired polynucleotide sequence, which are identified as described above, may be sequenced to identify the DNA sequence(s) 5 present in the clone, which sequence information can be used to screen a database for similar sequences or functional characteristics. Thus, in accordance with the present invention it is possible to isolate and identify: (i) DNA having a sequence of interest (e.g., a sequence encoding an enzyme having a specified enzyme activity), (ii) associate the sequence with known or unknown sequence in a database (e.g., database 10 sequence associated with an enzyme having an activity (including the amino acid sequence thereof)), and (iii) produce recombinant enzymes having such activity.

Sequencing may be performed by high through-put sequencing techniques. The exact method of sequencing is not a limiting factor of the invention. Any method useful in identifying the sequence of a particular cloned DNA sequence can be used. In 15 general, sequencing is an adaptation of the natural process of DNA replication. Therefore, a template (e.g., the vector) and primer sequences are used. One general template preparation and sequencing protocol begins with automated picking of bacterial colonies, each of which contains a separate DNA clone which will function as a template for the sequencing reaction. The selected clones are placed into media, and grown 20 overnight. The DNA templates are then purified from the cells and suspended in water. After DNA quantification, high-throughput sequencing is performed using a sequencers, such as Applied Biosystems, Inc., Prism 377 DNA Sequencers. The resulting sequence data can then be used in additional methods, including to search a database or databases.

Database Searches and Alignment Algorithms

25 A number of source databases are available that contain either a nucleic acid sequence and/or a deduced amino acid sequence for use with the invention in identifying or determining the activity encoded by a particular polynucleotide sequence. All or a representative portion of the sequences (e.g., about 100 individual clones) to be tested are used to search a sequence database (e.g., GenBank, PFAM or ProDom), either 30 simultaneously or individually. A number of different methods of performing such

sequence searches are known in the art. The databases can be specific for a particular organism or a collection of organisms. For example, there are databases for the *C. elegans*, *Arabidopsis. sp.*, *M. genitalium*, *M. jannaschii*, *E. coli*, *H. influenzae*, *S. cerevisiae* and others. The sequence data of the clone is then aligned to the sequences in 5 the database or databases using algorithms designed to measure homology between two or more sequences.

Such sequence alignment methods include, for example, BLAST (Altschul *et al.*, 1990), BLITZ (MPsrch) (Sturrock & Collins, 1993), and FASTA (Person & Lipman, 1988). The probe sequence (*e.g.*, the sequence data from the clone) can be any length, 10 and will be recognized as homologous based upon a threshold homology value. The threshold value may be predetermined, although this is not required. The threshold value can be based upon the particular polynucleotide length. To align sequences a number of different procedures can be used. Typically, Smith-Waterman or Needleman-Wunsch algorithms are used. However, as discussed faster procedures such as BLAST, 15 FASTA, PSI-BLAST can be used.

For example, optimal alignment of sequences for aligning a comparison window may be conducted by the local homology algorithm of Smith (Smith and Waterman, Adv Appl Math, 1981; Smith and Waterman, J Teor Biol, 1981; Smith and Waterman, J Mol Biol, 1981; Smith *et al*, J Mol Evol, 1981), by the homology alignment algorithm of 20 Needleman (Needleman and Wuncsch, 1970), by the search of similarity method of Pearson (Pearson and Lipman, 1988), by computerized implementations of these algorithms (GAP, BESTFIT, FASTA, and TFASTA in the Wisconsin Genetics Software Package Release 7.0, Genetics Computer Group, 575 Science Dr., Madison, WI, or the Sequence Analysis Software Package of the Genetics Computer Group, University of 25 Wisconsin, Madison, WI), or by inspection, and the best alignment (*i.e.*, resulting in the highest percentage of homology over the comparison window) generated by the various methods is selected. The similarity of the two sequence (*i.e.*, the probe sequence and the database sequence) can then be predicted.

Such software matches similar sequences by assigning degrees of homology to 30 various deletions, substitutions and other modifications. The terms “homology” and

“identity” in the context of two or more nucleic acids or polypeptide sequences, refer to two or more sequences or subsequences that are the same or have a specified percentage of amino acid residues or nucleotides that are the same when compared and aligned for maximum correspondence over a comparison window or designated region as measured 5 using any number of sequence comparison algorithms or by manual alignment and visual inspection.

For sequence comparison, typically one sequence acts as a reference sequence, to which test sequences are compared. When using a sequence comparison algorithm, test and reference sequences are entered into a computer, subsequence coordinates are 10 designated, if necessary, and sequence algorithm program parameters are designated. Default program parameters can be used, or alternative parameters can be designated. The sequence comparison algorithm then calculates the percent sequence identities for the test sequences relative to the reference sequence, based on the program parameters.

A “comparison window”, as used herein, includes reference to a segment of any 15 one of the number of contiguous positions selected from the group consisting of from 20 to 600, usually about 50 to about 200, more usually about 100 to about 150 in which a sequence may be compared to a reference sequence of the same number of contiguous positions after the two sequences are optimally aligned.

One example of a useful algorithm is BLAST and BLAST 2.0 algorithms, which 20 are described in Altschul *et al.*, Nuc. Acids Res. 25:3389-3402 (1977) and Altschul *et al.*, J. Mol. Biol. 215:403-410 (1990), respectively. Software for performing BLAST analyses is publicly available through the National Center for Biotechnology Information (<http://www.ncbi.nlm.nih.gov/>). This algorithm involves first identifying high scoring sequence pairs (HSPs) by identifying short words of length W in the query sequence, 25 which either match or satisfy some positive-valued threshold score T when aligned with a word of the same length in a database sequence. T is referred to as the neighborhood word score threshold (Altschul *et al.*, *supra*). These initial neighborhood word hits act as seeds for initiating searches to find longer HSPs containing them. The word hits are extended in both directions along each sequence for as far as the cumulative alignment 30 score can be increased. Cumulative scores are calculated using, for nucleotide

sequences, the parameters M (reward score for a pair of matching residues; always >0). The BLAST algorithm parameters W, T, and X determine the sensitivity and speed of the alignment. The BLASTN program (for nucleotide sequences) uses as defaults a wordlength (W) of 11, an expectation (E) of 10, M=5, N=-4 and a comparison of both 5 strands.

The BLAST algorithm also performs a statistical analysis of the similarity between two sequences (see, *e.g.*, Karlin & Altschul, Proc. Natl. Acad. Sci. USA 90:5873 (1993)). One measure of similarity provided by BLAST algorithm is the smallest sum probability (P(N)), which provides an indication of the probability by 10 which a match between two nucleotide sequences would occur by chance. For example, a nucleic acid is considered similar to a references sequence if the smallest sum probability in a comparison of the test nucleic acid to the reference nucleic acid is less than about 0.2, more preferably less than about 0.01, and most preferably less than about 0.001.

15 Sequence homology means that two polynucleotide sequences are homolgous (*i.e.*, on a nucleotide-by-nucleotide basis) over the window of comparison. A percentage of sequence identity or homology is calculated by comparing two optimally aligned sequences over the window of comparison, determining the number of positions at which the identical nucleic acid base (*e.g.*, A, T, C, G, U, or I) occurs in both sequences 20 to yield the number of matched positions, dividing the number of matched positions by the total number of positions in the window of comparison (*i.e.*, the window size), and multiplying the result by 100 to yield the percentage of sequence homology. This substantial homology denotes a characteristic of a polynucleotide sequence, wherein the polynucleotide comprises a sequence having at least 60 percent sequence homology, 25 typically at least 70 percent homology, often 80 to 90 percent sequence homology, and most commonly at least 99 percent sequence homology as compared to a reference sequence of a comparison window of at least 25-50 nucleotides, wherein the percentage of sequence homology is calculated by comparing the reference sequence to the polynucleotide sequence which may include deletions or additions which total 20 30 percent or less of the reference sequence over the window of comparison.

Sequences having sufficient homology can be further identified by any annotations contained in the database, including, for example, species and activity information. Accordingly, in a typical environmental sample, a plurality of nucleic acid sequences will be obtained, cloned, sequenced and corresponding homologous sequences from a database identified. This information provides a profile of the polynucleotides present in the sample, including one or more features associated with the polynucleotide including the organism and activity associated with that sequence or any polypeptide encoded by that sequence based on the database information. As used herein “fingerprint” or “profile” refers to the fact that each sample will have associated with it a set of polynucleotides characteristic of the sample and the environment from which it was derived. Such a profile can include the amount and type of sequences present in the sample, as well as information regarding the potential activities encoded by the polynucleotides and the organisms from which polynucleotides were derived. This unique pattern is each sample’s profile or fingerprint.

In some instances it may be desirable to express a particular cloned polynucleotide sequence once its identity or activity is determined or an suggested identity or activity is associated with the polynucleotide. In such instances the desired clone, if not already cloned into an expression vector, is ligated downstream of a regulatory control element (*e.g.*, a promoter or enhancer) and cloned into a suitable host cell. Expression vectors are commercially available along with corresponding host cells for use in the invention.

As representative examples of expression vectors which may be used there may be mentioned viral particles, baculovirus, phage, plasmids, phagemids, cosmids, phosmids, bacterial artificial chromosomes, viral nucleic acid (*e.g.*, vaccinia, adenovirus, foul pox virus, pseudorabies and derivatives of SV40), P1-based artificial chromosomes, yeast plasmids, yeast artificial chromosomes, and any other vectors specific for specific hosts of interest (such as bacillus, aspergillus, yeast, *etc.*) Thus, for example, the DNA may be included in any one of a variety of expression vectors for expressing a polypeptide. Such vectors include chromosomal, nonchromosomal and synthetic DNA sequences. Large numbers of suitable vectors are known to those of skill in the art, and are commercially available. The following vectors are provided by way of example;

Bacterial: pQE70, pQE60, pQE-9 (Qiagen), psiX174, pBluescript SK, pBluescript KS, pNH8A, pNH16a, pNH18A, pNH46A (Stratagene); pTRC99a, pKK223-3, pKK233-3, pDR540, pRIT5 (Pharmacia); Eukaryotic: pWLNEO, pSV2CAT, pOG44, pXT1, pSG (Stratagene), pSVK3, pBPV, pMSG, pSVL (Pharmacia). However, any other plasmid
5 or vector may be used as long as they are replicable and viable in the host.

The nucleic acid sequence in the expression vector is operatively linked to an appropriate expression control sequence(s) (promoter) to direct mRNA synthesis. Particular named bacterial promoters include lacI, lacZ, T3, T7, gpt, lambda PR, PL and trp. Eukaryotic promoters include CMV immediate early, HSV thymidine kinase, early
10 and late SV40, LTRs from retrovirus, and mouse metallothionein-I. Selection of the appropriate vector and promoter is well within the level of ordinary skill in the art. The expression vector also contains a ribosome binding site for translation initiation and a transcription terminator. The vector may also include appropriate sequences for amplifying expression. Promoter regions can be selected from any desired gene using
15 CAT (chloramphenicol transferase) vectors or other vectors with selectable markers.

In addition, the expression vectors preferably contain one or more selectable marker genes to provide a phenotypic trait for selection of transformed host cells such as dihydrofolate reductase or neomycin resistance for eukaryotic cell culture, or such as tetracycline or ampicillin resistance in *E. coli*.

20 The nucleic acid sequence(s) selected, cloned and sequenced as hereinabove described can additionally be introduced into a suitable host to prepare a library which is screened for the desired enzyme activity. The selected nucleic acid is preferably already in a vector which includes appropriate control sequences whereby a selected nucleic acid encoding an enzyme may be expressed, for detection of the desired activity. The host
25 cell can be a higher eukaryotic cell, such as a mammalian cell, or a lower eukaryotic cell, such as a yeast cell, or the host cell can be a prokaryotic cell, such as a bacterial cell. The selection of an appropriate host is deemed to be within the scope of those skilled in the art from the teachings herein.

30 In some instances it may be desirable to perform an amplification of the nucleic acid sequence present in a sample or a particular clone that has been isolated. In this

embodiment the nucleic acid sequence is amplified by PCR reaction or similar reaction known to those of skill in the art. Commercially available amplification kits are available to carry out such amplification reactions.

In addition, it is important to recognize that the alignment algorithms and
5 searchable database can be implemented in computer hardware, software or a combination thereof. Accordingly, the isolation, processing and identification of nucleic acid sequences and the corresponding polypeptides encoded by those sequence can be implemented in an automated system.

Without further elaboration, it is believed that one skilled in the art can, using
10 the preceding description, utilize the present invention to its fullest extent. The following examples are to be considered illustrative and thus are not limiting of the remainder of the disclosure in any way whatsoever.

Example 1

DNA Isolation and Library Construction

15 The following outlines the procedures used to generate a gene library from a mixed population of organisms.

DNA isolation. DNA is isolated using the IsoQuick Procedure as per manufacturer's instructions (Orca, Research Inc., Bothell, WA). DNA can be
20 normalized according to Example 2 below. Upon isolation the DNA is sheared by pushing and pulling the DNA through a 25G double-hub needle and a 1-cc syringes about 500 times. A small amount is run on a 0.8% agarose gel to make sure the majority of the DNA is in the desired size range (about 3-6 kb).

25 **Blunt-ending DNA.** The DNA is blunt-ended by mixing 45 μ l of 10X Mung Bean Buffer, 2.0 μ l Mung Bean Nuclease (150 u/ μ l) and water to a final volume of 405 μ l. The mixture is incubate at 37⁰C for 15 minutes. The mixture is phenol/chloroform extracted followed by an additional chloroform extraction. One ml of ice cold ethanol is added to the final extract to precipitate the DNA. The DNA
30 is precipitated for 10 minutes on ice. The DNA is removed by centrifugation in a

microcentrifuge for 30 minutes. The pellet is washed with 1 ml of 70% ethanol and repelleted in the microcentrifuge. Following centrifugation the DNA is dried and gently resuspended in 26 μ l of TE buffer.

5 **Methylation of DNA.** The DNA is methylated by mixing 4 μ l of 10X *EcoR I* Methylase Buffer, 0.5 μ l SAM (32 mM), 5.0 μ l *EcoR I* Methylase (40 u/ μ l) and incubating at 37°C, 1 hour. In order to insure blunt ends, add to the methylation reaction: 5.0 μ l of 100 mM MgCl₂, 8.0 μ l of dNTP mix (2.5 mM of each dGTP, dATP, dTTP, dCTP), 4.0 μ l of Klenow (5 u/ μ l) and incubate at 12°C for 30 minutes.

10

After 30 minutes add 450 μ l 1X STE. The mixture is phenol/chloroform extracted once followed by an additional chloroform extraction. One ml of ice cold ethanol is added to the final extract to precipitate the DNA. The DNA is precipitated for 10 minutes on ice. The DNA is removed by centrifugation in a microcentrifuge for 30 minutes. The pellet is washed with 1 ml of 70% ethanol, repelleted in the microcentrifuge and allowed to dry for 10 minutes.

15 **Ligation.** The DNA is ligated by gently resuspending the DNA in 8 μ l *EcoR I* adaptors (from Stratagene's cDNA Synthesis Kit), 1.0 μ l of 10X Ligation Buffer, 1.0 μ l of 10 mM rATP, 1.0 μ l of T4 DNA Ligase (4Wu/ μ l) and incubating at 4°C for 2 days. The ligation reaction is terminated by heating for 30 minutes at 70°C.

20 **Phosphorylation of adaptors.** The adaptor ends are phosphorylated by mixing the ligation reaction with 1.0 μ l of 10X Ligation Buffer, 2.0 μ l of 10mM rATP, 6.0 μ l of H₂O, 1.0 μ l of polynucleotide kinase (PNK) and incubating at 37°C for 30 minutes. After 30 minutes 31 μ l H₂O and 5 ml 10X STE are added to the reaction and the sample is size fractionate on a Sephadex S-500 spin column. The pooled fractions (1-3) are phenol/chloroform extracted once followed by an additional chloroform extraction. The DNA is precipitated by the addition of ice cold ethanol on ice for 10 minutes. The precipitate is pelleted by centrifugation in a microfuge at high speed for 30 minutes. The resulting pellet is washed with 1 ml 70% ethanol, repelleted by centrifugation and allowed to dry for 10 minutes. The sample is

resuspended in 10.5 μ l TE buffer. Do not plate. Instead, ligate directly to lambda arms as above except use 2.5 μ l of DNA and no water.

Sucrose Gradient (2.2 ml) Size Fractionation. Stop ligation by heating the

5 sample to 65°C for 10 minutes. Gently load sample on 2.2 ml sucrose gradient and centrifuge in mini-ultracentrifuge at 45K, 20°C for 4 hours (no brake). Collect fractions by puncturing the bottom of the gradient tube with a 20G needle and allowing the sucrose to flow through the needle. Collect the first 20 drops in a Falcon 2059 tube then collect 10 1-drop fractions (labeled 1-10). Each drop is about 60 μ l in

10 volume. Run 5 μ l of each fraction on a 0.8% agarose gel to check the size. Pool fractions 1-4 (about 10-1.5 kb) and, in a separate tube, pool fractions 5-7 (about 5-0.5 kb). Add 1 ml ice cold ethanol to precipitate and place on ice for 10 minutes. Pellet the precipitate by centrifugation in a microfuge at high speed for 30 minutes. Wash the pellets by resuspending them in 1 ml 70% ethanol and repelleting them by

15 centrifugation in a microfuge at high speed for 10 minutes and dry. Resuspend each pellet in 10 μ l of TE buffer.

Test Ligation to Lambda Arms. Plate assay by spotting 0.5 μ l of the sample on agarose containing ethidium bromide along with standards (DNA samples of known concentration) to get an approximate concentration. View the samples using UV light and estimate concentration compared to the standards. Fraction 1-4 = >1.0 μ g/ μ l. Fraction 5-7 = 500 ng/ μ l.

Prepare the following ligation reactions (5 μ l reactions) and incubate 4°C, overnight:

Sample	H ₂ O	10X Ligase Buffer	10mM rATP	Lambda arms (ZAP)	Insert DNA	T4 DNA Ligase (4 Wu/l)
Fraction 1-4	0.5 μ l	0.5 μ l	0.5 μ l	1.0 μ l	2.0 μ l	0.5 μ l
Fraction 5-7	0.5 μ l	0.5 μ l	0.5 μ l	1.0 μ l	2.0 μ l	0.5 μ l

Test Package and Plate. Package the ligation reactions following

5 manufacturer's protocol. Stop packaging reactions with 500 μ l SM buffer and pool packaging that came from the same ligation. Titer 1.0 μ l of each pooled reaction on appropriate host (OD₆₀₀ = 1.0) [XLI-Blue MRF]. Add 200 μ l host (in mM MgSO₄) to Falcon 2059 tubes, inoculate with 1 μ l packaged phage and incubate at 37°C for 15 minutes. Add about 3 ml 48°C top agar [50ml stock containing 150 μ l IPTG (0.5M) and 300 μ l X-GAL (350 mg/ml)] and plate on 100 mm plates. Incubate the plates at 37°C, overnight.

10

Amplification of Libraries (5.0 x 10⁵ recombinants from each library).

Add 3.0 ml host cells (OD₆₀₀=1.0) to two 50 ml conical tube and inoculate with 2.5 X 15 10⁵ pfu of phage per conical tube. Incubate at 37°C for 20 minutes. Add top agar to each tube to a final volume of 45 ml. Plate each tube across five 150 mm plates. Incubate the plates at 37°C for 6-8 hours or until plaques are about pin-head in size. Overlay the plates with 8-10 ml SM Buffer and place at 4°C overnight (with gentle rocking if possible).

20

Harvest Phage. Recover phage suspension by pouring the SM buffer off each plate into a 50-ml conical tube. Add 3 ml of chloroform, shake vigorously and incubate at room temperature for 15 minutes. Centrifuge the tubes at 2K rpm for 10

minutes to remove cell debris. Pour supernatant into a sterile flask, add 500 μ l chloroform and store at 4°C.

Titer Amplified Library. Make serial dilutions of the harvested phage (for example, 10^{-5} = 1 μ l amplified phage in 1 ml SM Buffer; 10^{-6} = 1 μ l of the 10^{-3} dilution in 1 ml SM Buffer). Add 200 μ l host (in 10 mM MgSO₄) to two tubes. Inoculate one tube with 10 μ l 10^{-6} dilution (10^{-5}). Inoculate the other tube with 1 μ l 10^{-6} dilution (10^{-6}). Incubate at 37°C for 15 minutes. Add about 3 ml 48°C top agar [50ml stock containing 150 μ l IPTG (0.5M) and 375 μ l X-GAL (350 mg/ml)] to each tube and plate on 100 mm plates. Incubate the plates at 37°C, overnight. Excise the ZAP II library to create the pBLUESCRIPT library according to manufacturers protocols (Stratagene).

Example 2

Normalization

Prior to library generation, purified DNA can be normalized. DNA is first fractionated according to the following protocol. A sample composed of genomic DNA is purified on a cesium-chloride gradient. The cesium chloride ($R_f = 1.3980$) solution is filtered through a 0.2 μ m filter and 15 ml is loaded into a 35 ml OptiSeal tube (Beckman). The DNA is added and thoroughly mixed. Ten micrograms of bis-benzimide (Sigma; Hoechst 33258) is added and mixed thoroughly. The tube is then filled with the filtered cesium chloride solution and spun in a VTi50 rotor in a Beckman L8-70 Ultracentrifuge at 33,000 rpm for 72 hours. Following centrifugation, a syringe pump and fractionator (Brandel Model 186) are used to drive the gradient through an ISCO UA-5 UV absorbance detector set to 280 nm. Peaks representing the DNA from the organisms present in an environmental sample are obtained. Eubacterial sequences can be detected by PCR amplification of DNA encoding rRNA from a 10-fold dilution of the *E. coli* peak using the following primers to amplify:

30

Forward primer: 5'-AGAGTTTGATCCTGGCTCAG-3' (SEQ ID NO:2)

Reverse primer: 5'-GGTTACCTTGTACGACTT-3' (SEQ ID NO:3)

Recovered DNA is sheared or enzymatically digested to 3-6 kb fragments. Lone-linker primers are ligated and the DNA is sized selected. Size-selected DNA is amplified by PCR, if necessary.

5

Normalization is then accomplished as follows by resuspending double-stranded DNA sample in hybridization buffer (0.12 M NaH₂PO₄, pH 6.8/0.82 M NaCl/1 mM EDTA/0.1% SDS). The sample is overlaid with mineral oil and denatured by boiling for 10 minutes. Sample is incubated at 68°C for 12-36 hours.

10 Double-stranded DNA is separated from single-stranded DNA according to standard protocols (Sambrook, 1989) on hydroxyapatite at 60°C. The single-stranded DNA fraction is desalted and amplified by PCR. The process is repeated for several more rounds (up to 5 or more).

15

Example 3

FACS/Biopanning

Infection of library lysates into Exp503 *E.coli* strain. 25 ml LB + Tet culture of Exp503 were cultured overnight at 37 C. The next day the culture was centrifuged at 4000 rpm for 10 minutes and the supernatant decanted. 20ml 10mM MgSO₄ was added and the OD₆₀₀ checked. Dilute to OD 1.0.

In order to obtain a good representation of the library, at least 2-fold (and preferably 5-fold) of the library lysate titer was used. For example: Titer of library lysate is 2x10⁶ cfu/ml. Need to plate at least 4x10⁶ cfu. Can plate approx. 500,000 microcolonies/ 150mm LB-Kan plate. Need 8 plates. Can plate 1 ml of reaction/plate- need 8 mls of cells + lysate.

2-fold (ex. 2 ml) of library lysate was mixed with appropriate amount (e.g., 6 ml) of OD 1.0 Exp503. The sample was incubated at 37°C for at least 1 hour. Plated 30 1 ml reaction on 150mm LB-Kan plate x 8 plates and incubated overnight at 30°C.

Harvesting, induction, and fixing of library in Exp503 cells. Scrape all cells from plates into 20 ml LB using a rubber policeman. Dilute cells approx. 1:100 (200 μ l cells/ 20 ml LB) and incubate at 37°C until culture is OD 0.3. Add 1:50 dilution of 20% sterile Glucose and incubate at 37°C until culture is OD 1.0. Add 5 1:100 dilution of 1M MgSO₄. Transfer 5 ml of culture to a fresh tube and the remaining culture can be used as an uninduced control if desired or discarded. Add 10 MOI 5 of CE6 bacteriophage to the remaining 5 ml of culture. (CE6 codes for T7 RNA Polymerase) (e.g., OD 1 = 8×10^8 cells/ml x 5 ml = 4×10^9 cells x MOI 5 = 2×10^{10} bacteriophage needed). Incubate culture + CE6 for 2 hr at 37°C. Cool on ice 15 and centrifuge cells at 4000 rpm for 10 min. Wash with 10 ml PBS. Fix cells in 600 μ l PBS + 1.8 ml fresh, filtered 4% paraformaldehyde. Incubate on ice for 2 hrs. (4% Paraformaldehyde: Heat 8.25 ml PBS in flask at 65°C. Add 100 μ l 1M NaOH and 0.5 g paraformaldehyde (stored at 4°C.) Mix until dissolved. Add 4.15 ml PBS. Cool to 0°C. Adjust pH to 7.2 with 0.5 M NaH₂PO₄. Cool to 0°C. Syringe filter. Use within 20 24 hrs). After fixing, centrifuge at 4000 rpm for 10 min. Resuspend in 1.8 ml PBS and 200 μ l 0.1% NP40. Store at 4°C overnight.

Hybridization of fixed cells. Centrifuge fixed cells at 4000 rpm for 10 min. Resuspend in 1 ml 40 mM Tris pH7.6/ 0.2% NP40. Transfer 100 μ l fixed cells to an 20 eppendorf tube. Centrifuge for 1 min and remove supernatant. Resuspend each reaction in 50 μ l Hybridization buffer (0.9 M NaCl; 20 mM Tris pH7.4; 0.01% SDS; 25% formamide- can be made in advance and stored at -20°C.). Add 0.5 nmol fluorescein-labeled primer to the appropriate reactions. Incubate with rocking at 46°C for 2 hr. (Hybridization temperature may depend on sequence of primer and 25 template.) Add 1 ml wash buffer to each reaction, rinse briefly and centrifuge for 1 min. Discard supernatant. (Wash buffer: 0.9 M NaCl; 20 mM Tris pH 7.4; 0.01% SDS). Add another 1 ml of wash buffer to each reaction, and incubate at 48°C with

rocking for 30 min. Centrifuge and remove supernatant. Visualize cells under microscope using WIB filter.

FACS sorting. Dilute cells in 1 ml PBS. If cells are clumping, sonicate for 20 seconds at 1.5 power. FAC sort the most highly fluorescent single-cells and collect in 0.5 ml PCR strip tubes (approximately one 96-well plate/ library). PCR single-cells with vector specific primers to amplify the insert in each cell. Electrophores all samples on an agarose gel and select samples with single inserts. These can be re-amplified with Biotin-labeled primers, hybridized to insert-specific primers, and examined in an ELISA assay. Positive clones can then be sequenced. Alternatively, the selected samples can be re-amplified with various combinations of insert-specific primers, or sequenced directly.

15

Example 4

Cell Staining Prior to FACS Screening

Gene libraries, including those generated as described in Example 1 and 3, can be further screened for bioactivities of interest on a FACS machine as indicated herein. A screening process begins with staining of the cells with a desirable substrate according to the following example.

A gene library is made from the hyperthermophilic archaeon *Sulfolobus solfataricus* in the λ -ZAPII vector according to the manufacturers instructions (Stratagene Cloning Systems, Inc., La Jolla, CA), and excised into the pBLUESCRIPT plasmid according to the manufacturers instructions (Stratagene).

DNA was isolated using the IsoQuick DNA isolation kit according to the manufacturers instructions (Orca, Inc., Bothell, WA).

To screen for β -galactosidase activity, cells are stained as follows. Cells are cultivated overnight at 37°C in an orbital shaker at 250rpm. Cells are centrifuged to collect about 2×10^7 cells (0.1ml of the culture), resuspended in 1ml of deionized

water, and stained with C₁₂-Fluoroscein-Di- (-D-galactopyranoside (FDG). Briefly, 0.5ml of cells are mixed with 50 μ l C₁₂-FDG staining solution (1mg C₁₂-FDG in 1ml of a mixture of 98% H₂O, 1% DMSO, 1% EtOH) and 50 μ l Propidium iodide (PI) staining solution (50 μ g/ml of distilled water). The sample is incubated in the dark at 5 37°C with shaking at 150rpm for 30 minutes. Cells are then heated to 70°C for 30 minutes (this step can be avoided if sample is not derived from a hyperthermophilic organism).

Example 5

10 **Screening of Expression Libraries by FACS and Recovery of Genetic Information of Sorted Organisms**

The excised λ -ZAP II library is incubated for 2 hours and induced with IPTG. Cells are centrifuged, washed and stained with the desired enzyme substrate, for example C₁₂-Fluoroscein-Di-(-D-galactopyranoside (FDG) as in Example 3. Clones 15 are sorted on a commercially available FACS machine, and positives are collected. Cells are lysed according to standard techniques (Current Protocols in Molecular Biology, 1987) and plasmids are transformed into new host by electroporation using standard techniques. Transformed cells are plated for secondary screening. The procedure is illustrated in Figure 1. Sorted organisms can be grown and plated for 20 secondary screening.

Example 6

Sorting Directly on Microtiter Plates

Cells can be sorted in a FACS instrument directly on microtiter plates in 25 accordance with the present invention. Sorting in this fashion facilitates downstream processing of positive clones.

E.coli cells containing β -galactosidase genes are exposed to a staining solution. These cells are then left to sit on ice for three minutes. For the cell sorting 30 procedure they are diluted 1:100 in deionized water or in Phosphate Buffered Saline solution according to the manufacturers protocols for cell sorting. The cells are then sorted by the FACS instrument into microtiter plates, one cell per well. The sorting

criteria is fluorescein fluorescence indicating β -galactosidase activity or PI for indicating the staining of dead cells (unlike viable cells, dead cells have no membrane potential; hence PI remains in the cell with dead cells and is pumped out with live cells).

5

Table 1

<u>Habitat</u>	<u>Cultured (%)</u>
Seawater	0.001-0.1
Freshwater	0.25
Mesotrophic lake	0.01-1.0
Unpolluted estuarine waters	0.1-3.0
Activated sludge	1.0-15.0
Sediments	0.25
Soil	0.3

Example 6

Production of single cells or fragmented mycelia

10 Inoculate 25ml MYME media (see recipe below) in 250 ml baffled flask with 100 μ l of *Streptomyces* 10712 spore suspension and incubated overnight @ 30°C 250rpm. After 24 hour incubation, transfer 10ml to 50ml conical polypropylene centrifuge tube and centrifuge @ 4,000rpm for 10 minutes @ 25°C. Decant supernatant and resuspend pellet in 10ml 0.05M TES buffer. Sort cells into MYM agar plates (sort 1 cell per drop, 5 cells per drop, 10 cells per drop) and incubate plates at 30°C.

15

20 MYME media (Yang, et.al., 1995 J. Bacteriol. 177(21): 6111-6117) contains: 10.3% sucrose, 1% maltose, 0.5% peptone, 0.3% yeast extract, 0.3% maltose extract, 5mM MgCl₂ and 1% glycine.

While the invention has been described in detail with reference to certain preferred embodiments thereof, it will be understood that modifications and variations are within the spirit and scope of that which is described and claimed.

WHAT IS CLAIMED IS:

1. A method for identifying a bioactivity or a biomolecule of interest, comprising:
 - (a) contacting a library containing a plurality of clones comprising polynucleotides derived from a mixed population of organisms or more than one organism, with at least one oligonucleotide probe labeled with a detectable molecule; and
 - (b) separating clones with an analyzer that detects the detectable molecule.
2. The method of claim 1 further comprising:
 - (a) contacting the separated clones with a reporter system that identifies a polynucleotide encoding a bioactivity or biomolecule of interest; and
 - (b) identifying clones capable of modulating expression or activity of the reporter system thereby identifying a polynucleotide of interest.
3. The method of claim 1, wherein the library is an expression library.
4. The method of claim 1, wherein the detectable molecule is a fluorescent molecule.
5. The method of claim 1, wherein the analyzer is a FACS analyzer.
6. The method of claim 1, wherein the mixed population of organisms is from an environmental sample.
7. The method of claim 1, wherein the mixed population of organisms comprises microorganisms.
8. The method of claim 6, wherein the environmental sample contains extremophiles.

9. The method of claim 8, wherein the extremophiles are selected from the group consisting of hyperthermophiles, psychrophiles, halophiles, psychrotrophs, alkalophiles, and acidophiles.
10. The method of claim 2, wherein the reporter system is a bioactive substrate.
11. The method of claim 10, wherein the bioactive substrate comprises C12FDG.
12. The method of claim 11, wherein the bioactive substrate further comprises a lipophilic tail.
13. The method of claim 1, further comprising prior to (a):
 - (i) obtaining polynucleotides from a mixed population of organisms; and
 - (ii) generating a polynucleotide library.
14. The method of claim 13, further comprising normalizing the polynucleotides prior to generating the library.
15. The method of claim 1, wherein the clones are encapsulated in a microenvironment.
16. The method of claim 15, wherein the microenvironment is selected from beads, high temperature agaroses, gel microdroplets, cells, ghost red blood cells, macrophages, or liposomes.
17. The method of claim 16, wherein the clones are encapsulated in a gel microdroplet.
18. The method of claim 1, wherein the polynucleotide of interest encodes an enzyme.

19. The method of claim 18, wherein the enzyme is selected from the group consisting of lipases, esterases, proteases, glycosidases, glycosyl transferases, phosphatases, kinases, mono- and dioxygenases, haloperoxidases, lignin peroxidases, diarylpropane peroxidases, eposize hydrolases, nitrile hydratases, nitrilases, transaminases, amidases, and acylases.

20. The method of claim 1, wherein the reporter system comprises a detectable label.

21. The method of claim 1, wherein the reporter system comprises a first test protein linked to a DNA binding moiety and a second test protein linked to a transcriptional activation moiety, wherein modulation of the interaction of the first test protein linked to a DNA binding moiety with the second test protein linked to a transcription activation moiety results in a change in the expression of a detectable protein.

22. The method of claim 1, wherein the polynucleotide of interest encodes a small molecule.

23. The method of claim 1, wherein the polynucleotide of interest, or fragments thereof, comprise one or more operons, or portions thereof.

24. The method of claim 23, wherein the operons, or portions thereof, encodes a complete or partial metabolic pathway.

25. The method of claim 24, wherein the operons or portions thereof encoding a complete or partial metabolic pathway encodes polyketide syntheses.

26. The method of claim 1, wherein the fluorescent analyzer is a fluorescence activated cell sorting (FACS) apparatus.

27. A method for identifying a polynucleotide encoding a polypeptide of interest comprising:
co-encapsulating in a microenvironment a plurality of library clones containing DNA obtained from a mixed population of organisms, with a mixture of oligonucleotide probes comprising a fluorescence marker and at least a portion of a polynucleotide sequence encoding a polypeptide of interest having a specified bioactivity under such conditions and for such time as to allow interaction of complementary sequences; and
identifying clones containing a complement to the oligonucleotide probe encoding the polypeptide of interest by separating clones with a fluorescent analyzer that detects fluorescence.

28. A method for high throughput screening of a polynucleotide library for a polynucleotide of interest that encodes a molecule of interest, comprising:
(a) contacting a library containing a plurality of clones comprising polynucleotides derived from a mixed population of organisms with a plurality of oligonucleotide probes labeled with a fluorescence molecule; and
(b) separating clones with a fluorescent analyzer that detects fluorescence.

29. The method of claim 28, further comprising:
(a) contacting the separated clones with a reporter system that identifies a polynucleotide encoding the molecule of interest; and
(b) identifying clones capable of modulating expression or activity of the reporter system thereby identifying a polynucleotide of interest.

30. The method of claim 28, wherein the library is an expression library.

31. The method of claim 28, wherein the mixed population of organisms is from an environmental sample.

32. The method of claim 28, wherein the mixed population of organisms comprises microorganisms.

33. The method of claim 32, wherein the environmental sample contains extremophiles.

34. The method of claim 33, wherein the extremophiles are selected from the group consisting of hyperthermophiles, psychrophiles, halophiles, psychrotrophs, alkalophiles, and acidophiles.

35. The method of claim 29, wherein the reporter system is a bioactive substrate.

36. The method of claim 35, wherein the bioactive substrate comprises C12FDG.

37. The method of claim 36, wherein the bioactive substrate further comprises a lipophilic tail.

38. The method of claim 28, further comprising prior to (a):
(i) obtaining polynucleotides from a mixed population of organisms; and
(ii) generating a polynucleotide library.

39. The method of claim 38, further comprising normalizing the polynucleotides prior to generating the library.

40. The method of claim 28, wherein the clones are encapsulated in a gel microdrop.

41. The method of claim 28, wherein the polynucleotide of interest encodes an enzyme.

42. The method of claim 41, wherein the enzyme is selected from the group consisting of lipases, esterases, proteases, glycosidases, glycosyl transferases,

phosphatases, kinases, mono- and dioxygenases, haloperoxidases, lignin peroxidases, diarylpropane peroxidases, epoxide hydrolases, nitrile hydratases, nitrilases, transaminases, amidases, and acylases.

43. The method of claim 29, wherein the reporter system comprises a detectable label.
44. The method of claim 28, wherein the reporter system comprises a first test protein linked to a DNA binding moiety and a second test protein linked to a transcriptional activation moiety, wherein modulation of the interaction of the first test protein linked to a DNA binding moiety with the second test protein linked to a transcription activation moiety results in a change in the expression of a detectable protein.
45. The method of claim 28, wherein the polynucleotide of interest encodes a small molecule.
46. The method of claim 28, wherein the polynucleotide of interest, or fragments thereof, comprise one or more operons, or portions thereof.
47. The method of claim 46, wherein the operons, or portions thereof, encodes a complete or partial metabolic pathway.
48. The method of claim 47, wherein the operons or portions thereof encoding a complete or partial metabolic pathway encodes polyketide syntheses.
49. The method of claim 28, wherein the fluorescent analyzer is a fluorescence activated cell sorting (FACS) apparatus.
50. The method of claim 28, wherein the plurality of oligonucleotide probes have different nucleic acid sequences.

51. The method of claim 50, wherein the sequences are portions of a polynucleotide encoding a molecule of interest.

52. The method of claim 28, wherein the plurality of oligonucleotide probes have the same nucleic acid sequence.

53. A method of screening for a polynucleotide encoding an activity of interest, comprising:
(a) obtaining polynucleotides from an environmental sample;
(b) normalizing the polynucleotides obtained from the sample;
(c) generating a library from the normalized polynucleotides;
(d) contacting the library with a plurality of oligonucleotide probes comprising a fluorescent marker and at least a portion of a polynucleotide sequence encoding a polypeptide of interest having a specified activity to select library clones positive for a sequence of interest; and
(e) selecting clones with a fluorescent analyzer that detects fluorescence.

54. The method of claim 53, further comprising:
(a) contacting the selected clones with a reporter system that identifies a polynucleotide encoding the activity of interest; and
(b) identifying clones capable of modulating expression or activity of the reporter system thereby identifying a polynucleotide of interest; wherein the positive clones contain a polynucleotide sequence encoding an activity of interest which is capable of catalyzing the bioactive substrate.

55. A method for screening polynucleotides, comprising:
(a) contacting a library of polynucleotides wherein the polynucleotides are derived from a mixed population of organism with a probe oligonucleotide labeled with a fluorescence molecule, which fluoresce

upon binding of the probe to a target polynucleotide of the library, to select library polynucleotides positive for a sequence of interest;

- (b) separating library members that are positive for the sequence of interest with a fluorescent analyzer that detects fluorescence; and
- (c) expressing the selected polynucleotides to obtain polypeptides.

56. The method of claim 55, further comprising:

- (a) contacting the polypeptides with a reporter system; and
- (b) identifying polynucleotides encoding polypeptides capable of modulating expression or activity of the reporter system.

57. A method for obtaining an organism from a mixed population of organisms in a sample comprising:

- (a) encapsulating in a microenvironment at least one organism from the sample;
- (b) incubating the encapsulated at least one organism under such conditions and for such a time to allow the at least one microorganism to grow or proliferate; and
- (c) sorting the encapsulated at least one organism by flow cytometry to obtain an organism from the sample.

58. The method of claim 57, wherein the mixed population of organisms is from an environmental sample.

59. The method of claim 57, wherein the mixed population of organisms comprises microorganisms.

60. The method of claim 58, wherein the environmental sample contains extremophiles.

61. The method of claim 60, wherein the extremophiles are selected from the group consisting of hyperthermophiles, psychrophiles, halophiles, psychrotrophs, alkalophiles, and acidophiles.

ABSTRACT

Provided is a method of screening or enriching a sample containing polynucleotides from a mixed population of organisms. The method includes creating a DNA library from a plurality of nucleic acid sequences of a mixed population of organisms and separating clones containing a polynucleotide sequence of interest on a fluorescence analyzer based upon hybridization of a fluorescence labeled oligonucleotide probe to a target sequence in a clone. The separated or enrich library can then be further process by activity based screening or sequence based screening. In addition, the enriched sequence can be compared to a database and to identify sequences in the database which have homology to a clone in the library thereby obtaining a nucleic acid profile of the mixed population of organisms.

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*Cell sorting to screen for recombinant enzymes
excise λ library into *E.coli**

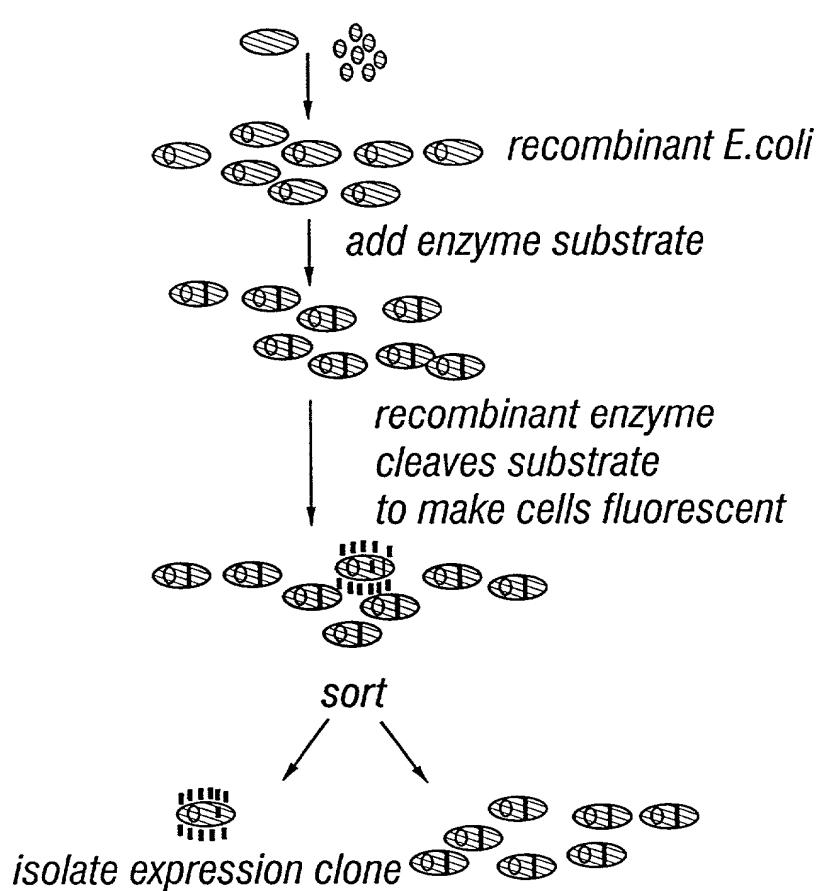


FIG. 1

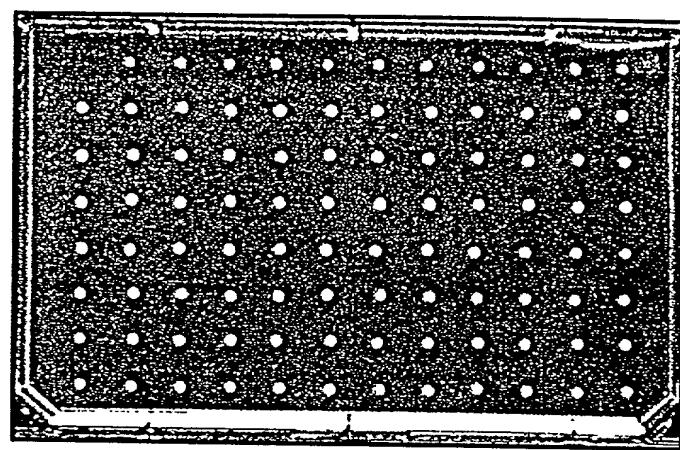
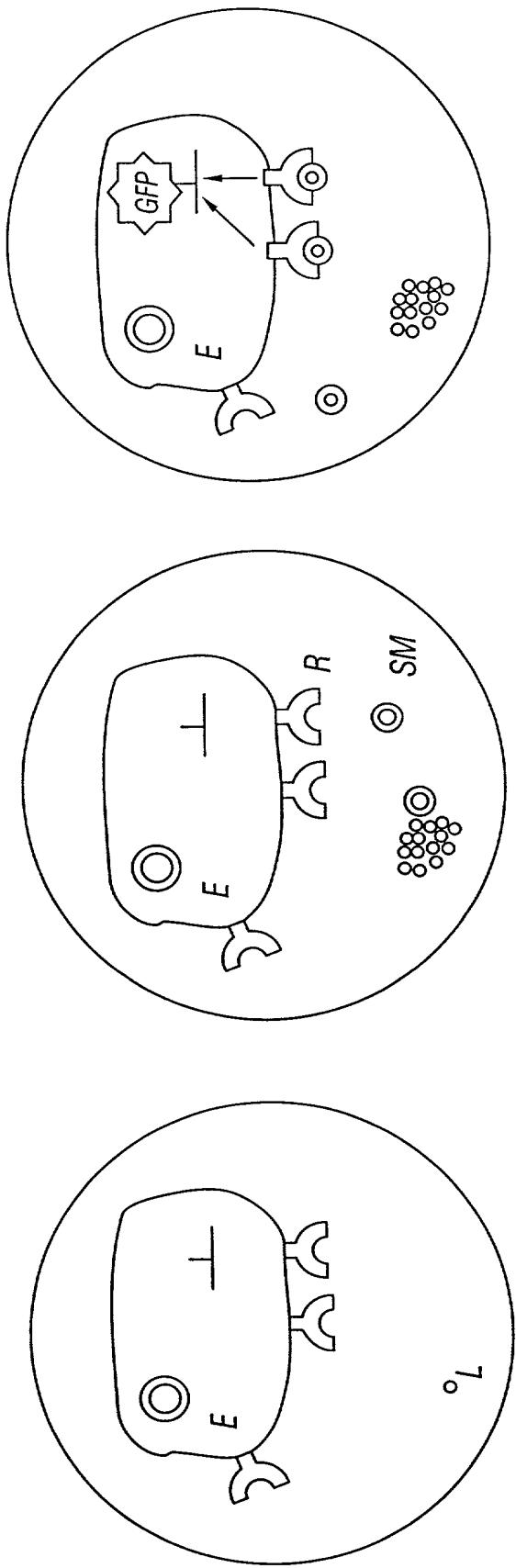


FIG. 2



Receptor binding of small molecule & GFP reporting

Growth and expression of small molecule from host

Co-encapsulation library + Eukaryote

E=Eukaryotic assay organism **L**=Large insert library
GFP= Green fluorescent protein **R**=Eukaryotic receptor
SM=Small molecule

FIG. 3

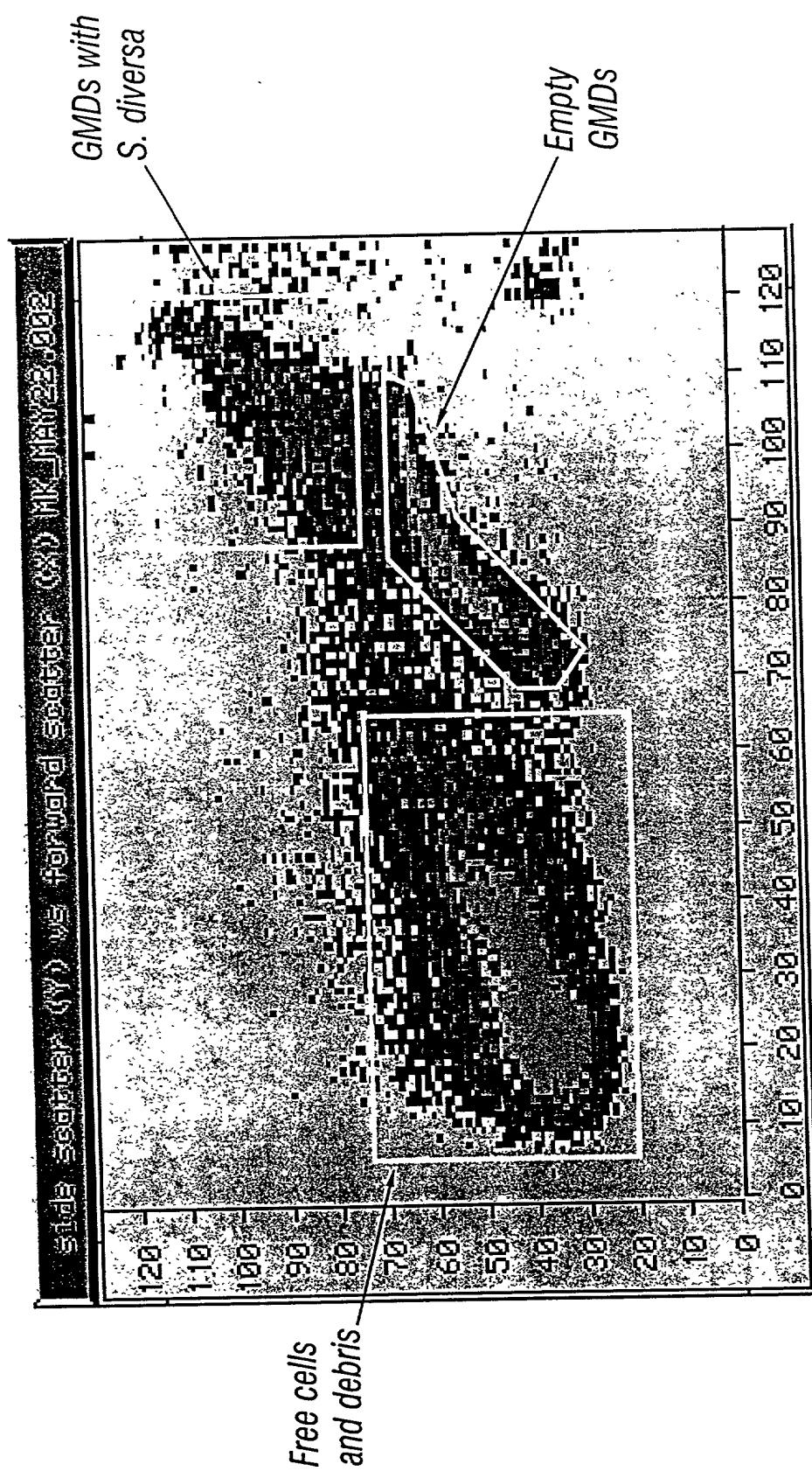


FIG. 4

Whole Cell Hybridization Protocol (+ control)

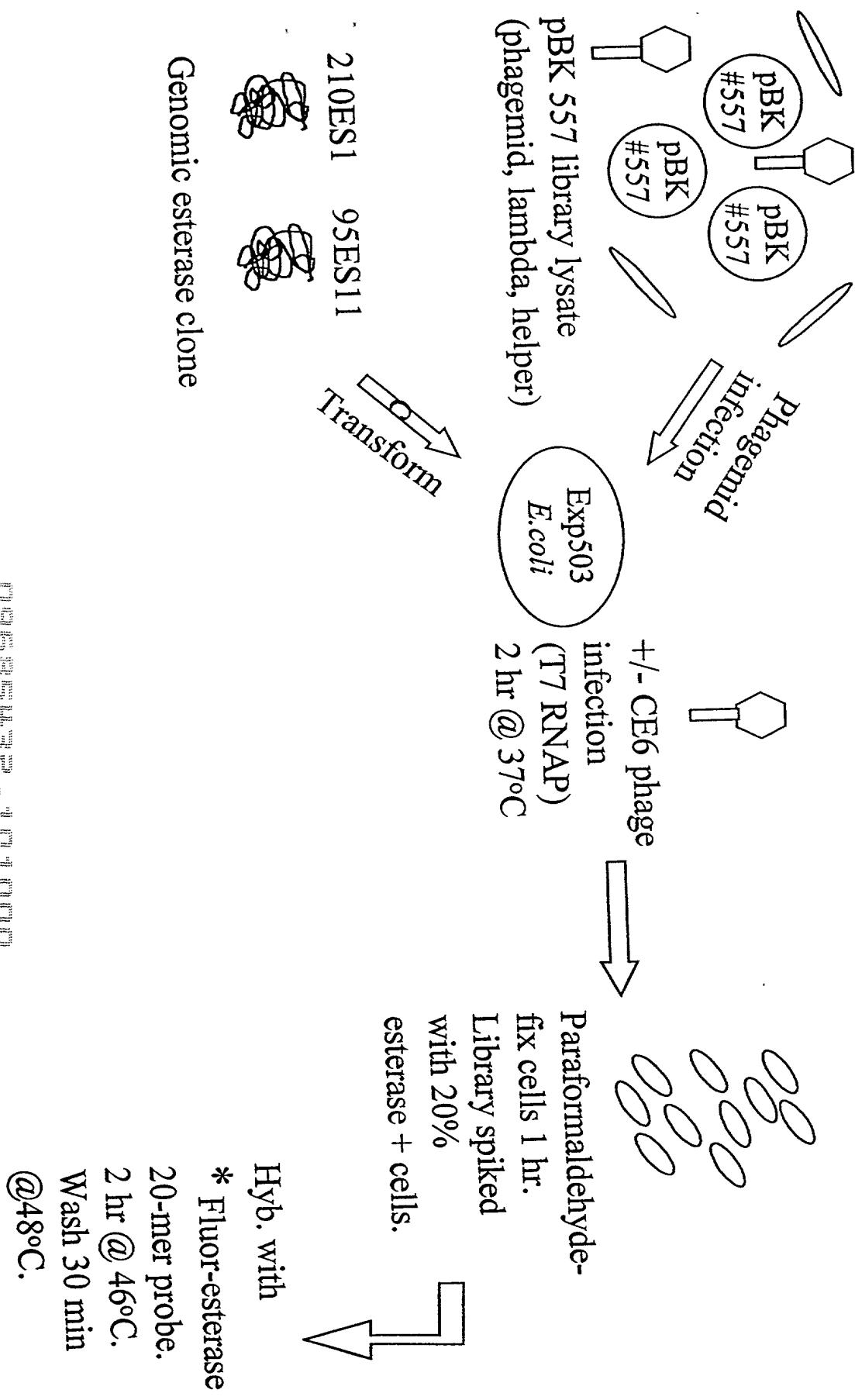


FIG. 5

DECLARATION FOR PATENT APPLICATION

As a below-named inventor, I hereby declare that:

My residence, post office address and citizenship is as stated below next to my name.

I believe that I am the original, first and sole inventor (if only one name is listed below) or an original, first and joint inventor (if plural names are listed below) of the subject matter which is claimed and for which a patent is sought on the invention entitled HIGH THROUGHPUT SCREENING FOR SEQUENCES OF INTEREST, the specification of which

_____ is attached hereto.

X was filed on October 10, 2000 (Attorney Docket No. DIVER1280-3) as U.S. Application Serial No. _____ and was amended on _____ if applicable (the "Application").

I hereby authorize and request insertion of the application serial number of the Application when officially known.

I hereby state that I have reviewed and understand the contents of the above-identified specification, including the claims, as amended by any amendment referred to above.

I acknowledge the duty to disclose to the United States Patent and Trademark Office all information known to me to be material to patentability of the subject matter of the Application as defined in Title 37, Code of Federal Regulations ("C.F.R."), § 1.56.

With respect to the Application, I hereby claim the benefit under 35 U.S.C. Section 120 of any United States application(s), or Section 365(c) of any PCT International application designating the United States, listed below and, insofar as the subject matter of GT\6205513.1

each of the claims of the application is not disclosed in the prior United States or PCT International application in the manner provided by the first paragraph of 35 U.S.C. Section 112, I acknowledge the duty to disclose to the United States Patent and Trademark Office all information known to me to be material to patentability of the subject matter of the Application as defined in Title 37, C.F.R., Section 1.56 which became available between the filing date of the prior application and the national or PCT International filing date of the Application:

<u>09/444,112</u> (Application Serial No.)	<u>November 22, 1999</u> (Filing Date)	<u>Pending</u> (Status)
<u>09/098,206</u> (Application Serial No.)	<u>June 16, 1998</u> (Filing Date)	<u>Pending</u> (Status)
<u>08/876,276</u> (Application Serial No.)	<u>June 16, 1997</u> (Filing Date)	<u>Pending</u>

I hereby claim foreign priority benefits under Title 35, United States Code, § 119 of any foreign application(s) for patent or inventor's certificate or of any PCT international application(s) designating at least one country other than the United States of America listed below and have also identified below any foreign application for patent or inventor's certificate or any PCT international application(s) designating at least one country other than the United States of America filed by me on the same subject matter having a filing date before that of the application(s) of which priority is claimed:

COUNTRY	APPLICATION NO.	FILING DATE	PRIORITY CLAIMED
_____	_____	_____	<input type="checkbox"/> Yes <input type="checkbox"/> No

I hereby declare that all statements made herein of my own knowledge are true and that all statements made on information and belief are believed to be true; and further that these statements were made with the knowledge that willful false statements and the like so made are punishable by fine or imprisonment, or both, under Section 1001 of Title 18 of the

United States Code and that such willful false statements may jeopardize the validity of the application or any patent issued thereon.

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Date: _____

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